Activation of the cyclic AMP cascade as an oncogenic mechanism: the thyroid example

C Maenhaut, PP Roger, S Reuse, JE Dumont

Institut de Recherche Interdisciplinaire, Fuculté de Médecine, Hôpital Erasme, Université Libre de Bruxelles, Campus Erasme, 808 route de Lennik, B-1070 Brussels, Belgium

(Received 26 November 1990; accepted 24 December 1990)

Summary — Three cascades activate thyroid cell proliferation: the EGF-protein tyrosine kinase pathway. the phorbol ester-protein kinase C pathway and the thyrotropin-cyclic AMP pathway. While the first 2 cascades converge early, they remain distinct from the cyclic AMP cascade until very late in G1. The cyclic AMP cascade is characterized by an early and transient expression of c-myc, which may explain why it induces proliferation and differentiation expression. Constitutive activation of this cascade causes growth and hyperfunctioning adenomas. The various possible defects that could lead to such a constitutive activation are discussed.

cyclic AMP / oncogenicity / proliferation / thyroid

Cyclic AMP in cell proliferation

It is probable that the key machinery in the immediate control of the decision of cells to divide will prove to be very general. Indeed, the first studies of complementation of proliferation deficient yeasts with mammalian genes have already allowed the identification in mammalian cells of genes able to perform the same function. One of these genes, $cdc2^+$, has been shown to be a part of the well-known maturation promoting factor involved in the triggering of meiosis in oocytes [1]. On the other hand, the regulatory circuits allowing the control of mammalian cells by extracellular signals vary in importance and significance from one cell type to another, and within one cell type from one species to another. It is therefore highly improbable that the role of these circuits in the control of the decision to divide will be conserved in all mammalian cells. Indeed, examples of opposite effects of the same cascade on proliferation and differentiation in different cell types abound: the phorbol ester protein kinase C pathway, which in the thyroid induces proliferation and dedifferentiation, exerts the opposite effects in keratinocytes. The cyclic AMP cascade which negatively regulates proliferation in fibroblasts and lymphocytes stimulates it in keratinocytes and thyrocytes, etc. Thus the role of a given cascade in the control of cell proliferation and differentiation has to be considered for each cell type

of each species. Generalizations are unwarranted in this field.

Reviews on the control of vertebrate cell proliferation generally consider 2 signal cascades as the main pathways between an extracellular mitogenic signal and mitogenesis itself: the growth factor receptor protein tyrosine kinase pathway and the phosphatidylinositol Ca²⁺-diacylglycerol protein kinase C cascade [2]. In such reviews, the first demonstrated signal cascade, the cyclic AMP system, is either not considered, or merely ascribed the role of an inhibitory pathway [3]. This general picture derives from the fact that growth and proliferation have mostly been studied on 'easy experimental objects', fibroblasts and cell lines of mesenchymal origin. As is often the case in science, the choice of experimental subject has biased the outlook on the whole subject. In fact, in several epithelial cells as in yeast cells, the cyclic AMP cascade is a positive regulatory pathway of cell growth and proliferation [4]. In the thyroid we shall consider at least 3 well defined distinct pathways: the hormone--receptor-adenylate cyclase-cyclic AMP protein kinase system, the hormone receptor-tyrosine protein kinase pathway and the hormone-receptor-phospholipase C-diacylglycerol protein kinase C-calcium calmodulin protein kinase cascade. The receptor tyrosine kinase pathway may be subdivided into 2 branches: some growth factors, such as EGF induce proliferation and repress

differentiation expression, others like FGF or IGF_1 , are mitogenic or are necessary for the proliferation effect of other factors without being mitogenic by themselves, but they do not inhibit differentiation expression.

As we might have assumed by looking at the present complexity of the regulation of a simple organism like the phage, this simple scheme is now expanding daily and becoming more sophisticated. First, new regulatory pathways are being defined, eg, the ANF guanylate cyclase-cyclic GMP, and the hormone-phospholipase A₂-arachidonate cascades. Second, at each level, any of the regulatory pathways may be directly controlled by any step in this or the other pathways. Thus, in the dog thyroid, calcium through calmodulin activates a cyclic nucleotide phosphodiesterase, diacylglycerol inhibits phospho-



Fig 1. Control of thyroid cyclic AMP cascade: inactivating (•) and activating (*) mutations that can lead to constitutive activation. H = hormone; R = receptor; NE = norepinephrine; AC = adenylate cyclase; cA = cyclic AMP; cAPK = cAMP-dependent protein kinase; \rightarrow transformation or transport; --[±] positive control; ---// negative control; G_s, G_i: stimulating and inhibiting G proteins; R_s, R_i: stimulating and inhibiting receptors; H₂O₂.G.S.: H₂O₂ generating system; Y: feedback inhibition on receptor cyclase system; R, C: regulatory, C catalytic unit.

lipase C and the prostaglandin E_2 stimulation of adenylate cyclase and potentiates TSH activation of this enzyme, etc. Third, the level of the proteins involved in each pathway may be controlled at gene transcription or distal to it, by the same or by other pathways. Fourth, the stimulation of a regulatory pathway in one cell may stimulate the production and release of factors that control these same cells (autocrine mechanisms) or neighboring cells (paracrine mechanisms). A mechanism of such interactions has been provided by Gärtner when he demonstrated the secretion by pig thyroid cells of a fibroblast growth factor [5]. Therefore, different mechanisms functioning to the same end will probably coexist in each system, and the demonstration of one such mechanism does not exclude the possibility of another. Moreover, at each level the circuits may differ from one species to another [6]. Therefore, although data may be coherent within one system they may vary from one system to another.

The mitogenic action of TSH

Although there is no doubt that in vivo TSH stimulates the proliferation of thyroid cells, in the 1970s there was no evidence that this effect was direct. Indeed, the ACTH trophic effect on the adrenal appears to be indirect. We first started to study early steps of growth in slices [7], and showed that TSH enhances ornithine decarboxylase activity in dog thyroid cells, which is generally considered as a preliminary to growth. The effect was mimicked by cAMP analogs and inhibited by agents inhibiting cAMP accumulation. As these results were contrary to current opinion, they were generally ignored. We as well as several others showed that TSH increases [3H] uridine incorporation into RNA in sheep thyroid slices. Did this show that TSH increased RNA synthesis? No: increased [3H] uridine incorporation into RNA only reflected an effect on the radioactivity of the precursor pool [8]. We realize from such an experiment why measurements of [3H] thymidine incorporation into DNA do not measure DNA synthesis either [9].

To investigate the problem of proliferation, we used primary cultures and a technique derived from Kerkof [10], Fayet [11] and Rapoport [12], with a serum-free medium supplemented as proposed by Ambesi [13]. Using several methods, we demonstrated that TSH and cAMP analogs or enhancers stimulate proliferation of dog thyroid cells [14, 15]. More recently we confirmed this result in normal human thyroid cells [16]. Other results obtained in various culture systems were sometimes contradictory. While in dog thyroid cells in primary culture, in rat thyroid follicles in suspension [17], in ovine cell lines (OVNI) [18], and in a rat cell line (FRTL) [13], thyrotropin has been

demonstrated to enhance or induce cell proliferation, to our knowledge, no such effect has been obtained in porcine [19], calf [20] or ovine [21] thyroid cells in primary culture. Whether this is due to inaccessibility of the TSH receptor(s), lack of an essential element in the culture medium, alteration of cell program in culture or real unresponsiveness to direct TSH action, is not known. It should be mentioned here that the stimulating effects of ACTH on the proliferation of adrenal cells in vivo have not been convincingly reproduced in vitro. In this case, there are arguments that the stimulating effect may be indirect: ACTH may induce the synthesis and secretion of growth factors by the adrenal cells which would then, acting as extracellular signals, trigger cell proliferation. It is therefore quite possible, and this has been proposed, that in the species for which no direct proliferative action of TSH has been demonstrated, such a mechanism might operate. In porcine thyroid cells, TSH through cyclic AMP induces EGF receptors making these cells more responsive to EGF [22]. In the control of thyroid cell proliferation differences in strategy from one species to another are possible.

It should be noted that in dog thyroid cells, TSH directly stimulates proliferation while maintaining the expression of differentiation. Differentiation expression, as evaluated by iodide transport, or thyroperoxidase and thyroglobulin mRNA content or nuclear transcription, is induced by TSH, forskolin, cholera toxin and cyclic AMP analogs, in dog thyroid cells [23–25]. Similar results, albeit partial, have been obtained in human and calf cells [20, 26]. These effects are obtained in all the cells of a culture, as shown by *in situ* hybridization experiments [27]. They are reversible and can be obtained either after the arrest of proliferation or during the cell division cycle [28].

The question arises as to the role of cyclic AMP in the TSH effects. Within minutes thyrotropin induces a striking morphological change in dog thyroid cells in culture: a rounding up following the disruption of the actin network [23, 28]. All the cells are affected. TSH also enhances the accumulation of cyclic AMP in these cells within < 5 min. Cyclic AMP remains elevated for 48 h in the continuous presence of the hormone. In the dog thyroid cells, analogs of cyclic AMP as well as general cyclase activators (forskolin, cholera toxin) reproduce all the effects of TSH: acute morphological changes, proliferation, expression of differentiation [23, 25]. Moreover, combinations of cyclic AMP analogs which are synergistic on the 2 cyclic AMP dependent kinases isoenzymes are also synergistic on these effects [29]. Cyclic AMP is therefore a general intracellular signal for function, proliferation and differentiation in dog thyroid cells. Similar results have been obtained for proliferation with human [16] and rat thyroid cells in culture [30]

and, despite a first contradictory report [31], in FRTL-5 cells [32]. It is interesting that in cloned, dedifferentiated tumorigenic FRTL-5 derived cells, cyclic AMP, as in fibroblasts, inhibits proliferation [33]. Thus changing the phenotype of these cells may reverse the role of cyclic AMP.

One argument that cAMP may be the mediator of rat thyroid cell proliferation *in vivo* is provided by the fact that methylxanthines, inhibitors of cyclic AMP phosphodiesterases, even at doses which do not further enhance serum TSH levels or decrease serum thyroid hormones greatly potentiate the goitrogenic action of propylthiouracil. By blocking thyroid hormone formation propylthiouracil, chronic increase in TSH levels and thyroid stimulation is induced. The action of methylxanthines is abolished by a high iodine diet or hypophysectomy [34]. Analogues of cyclic AMP injected in rat *in vivo* have also been reported to cause thyroid growth [35].

The kinetics of biochemical effects in the various thyroid mitogenic pathways

The kinetics of the induction of thymidine incorporation into nuclear DNA of dog thyroid cells is very similar for TSH, forskolin, EGF and phorbol esters (TPA) [36]. Whatever the stimulant, there is a similar minimal delay of 16 to 20 h before the onset of labelling, ie of DNA synthesis. This is the minimal time required to prepare the necessary machinery. For the cAMP pathway, the stimulatory agent has to be present during the entire prereplicative period: any interruption of stimulation (eg by forskolin washing) greatly delays the onset of DNA synthesis [37]. What happens between the onset of stimulation and the beginning of DNA synthesis? The time sequence of biochemical phenomena suggests a causal sequence. However, many of such postulated causal relationships remain to be proved.

We have thus studied the phenomenology of EGF. TPA and TSH proliferative action on dog quiescent cells with the aim of identifying steps in this action. Three biochemical aspects of the proliferative response occurring at different times during the prereplicative phase have been considered. The pattern of protein phosphorylation induced within minutes by TSH is reproduced by cyclic AMP analogs [38]. The phosphorylation of at least 11 proteins is increased or induced. NaOH treatment of the gels does not reveal any remaining phosphorylation on these proteins suggestive of tyrosine phosphorylation. In EGF stimulated cells, the phosphorylation of 5 proteins is stimulated, 2 of them phosphorylated on tyrosines (42 kDa). These 2 proteins are similar (isoelectric points, approximate molecular weight, composition in phosphorylated amino acids) [38] to the two 42 kDa proteins described in other systems, which have been implicated in the mitogenic response to diverse agents and recently identified as MAPkinase [39]. This kinase phosphorylates S_6 kinase II which is involved in the control of protein synthesis at ribosome level. Phorbol esters induce the phosphorylation of 19 proteins, including the tyrosine phosphorylated proteins mentioned above. There is no overlap in the patterns of protein phosphorylation induced by TSH and cyclic AMP enhancers on the one hand, and by EGF and phorbol esters on the other hand.

The expression of c-*myc* and c-*fos* has been studied by Northern analysis of RNA extracts [40]. As in other types of cells, EGF and TPA first enhance c-*fos*, then c-*myc* mRNA concentrations. On the other hand, TSH or forskolin enhance strongly but for a limited time c-*myc* mRNA concentration with the same kinetics as for EGF/TPA, c-*fos* mRNA concentration. In fact, cyclic AMP first enhances, then decreases c-*myc* mRNA accumulation. This second phenomenon is akin to what has been observed in fibroblasts in which cyclic AMP negatively regulates growth [41].

The pattern of proteins synthesized in response to the various proliferation stimuli has been studied [42]. Again 2 patterns emerge. TSH and forskolin induce the synthesis of at least 8 proteins and decrease the synthesis of 5 proteins. Epidermal growth factor, phorbol ester and serum induce the synthesis of at least 1 protein and decrease the synthesis of 2 proteins. The only overlap between the 2 patterns concerns the decrease in the synthesis of a protein (18 kDa) which is also reduced by EGF after proliferation has stopped. Only one protein has been shown to be synthesized in response to the 3 pathways: PCNA, the auxiliary protein of DNA polymerase δ ; but the kinetics of this synthesis are very different, with an early synthesis in the cyclic AMP cascade (consistent with a signal role) and a late S phase synthesis in the other cascades [42]. Thus, obviously 2 different phenomenologies are involved in the proliferation response to TSH through cyclic AMP on the one hand, and epidermal growth factor and phorbol ester, presumably through protein tyrosine phosphorylation, on the other hand. Although this conclusion needs to be further substantiated, it certainly suggests that the proliferation of dog thyroid cells is controlled by at least 2 largely independent pathways.

The studies of protein phosphorylation, protooncogene expression and protein synthesis in dog thyrocytes allow discrimination between 2 models of cyclic AMP action on proliferation in this system: a direct effect on the thyrocyte or an indirect effect through the secretion and autocrine action of another growth factor. If the effect of TSH through cyclic AMP involved such an autocrine loop one would expect faster kinetics of action of the growth factor, at least some common parts in the patterns of protein phosphorylation and protein synthesis induced by cyclic AMP and the growth factor. The results do not support such a hypothesis, at least for the growth factors we have tested.

IGF, which is secreted by FRTL-5 cells and is mitogenic for them [43] is not a growth factor for dog thyroid cells by itself. Moreover, our experiments are in general carried out in the presence of high concentrations of insulin that appear to saturate the IGF₁ receptor [37]. For protein kinase C activators and EGF, the kinetics of action of TSH or forskolin are similar for the end point of DNA synthesis for the 3 types of agents. Moreover, the kinetics of protooncogene c-mvc and c-fos expression are not delayed for TSH and cyclic AMP. Finally, the patterns of protein phosphorylation and protein synthesis induced by EGF and phorbol esters show partially common responses, while there was no overlap with the pattern of TSH or cyclic AMP action. Thus there is no evidence to support the involvement of an autocrine loop with a growth factor in the action of TSH and cyclic AMP on dog thyroid cells in primary culture. This does not exclude such a mechanism for the thyroid of other species as suggested by the induction by TSH of EGF receptors in porcine thyroid cells and of IGF₂ in FRTL-5 cells.

The paradox of TSH and cyclic AMP and their apparently opposite effects: stimulation of proliferation, induction of differentiation expression

The incompatibility at the cell level of a proliferation and differentiation program is commonly accepted in biology. In general, cells with a high proliferative capacity are partly differentiated and during development such cells lose this capacity as they progressively differentiate. Some cells even lose all potential to divide when reaching final differentiation; this is called terminal differentiation. Conversely, in tumor cells there is an inverse relationship between proliferation and differentiation expression. It is therefore not surprising that in thyroid cells, the general mitogenic agents and pathways, phorbol esters and the protein kinase C pathway, EGF and the protein tyrosine kinase pathway induce both proliferation and the loss of differentiation expression [44]. The effects of the cyclic AMP cascade are in striking contrast to this general concept. Indeed, TSH and cyclic AMP induce proliferation of dog thyrocytes while maintaining differentiation expression: both proliferation and differentiation programs can be

triggered by TSH in the same cells at the same time [27]. It is tempting to relate this apparent paradox to the role and expression of proto-oncogene in these cells. c-fos expression is enhanced in a great variety of cell stimulation, leading to either proliferation or differentiation expression [45]. On the other hand, if there is one generalization that could be made on proto-oncogenes, it is the dedifferentiating role of cmyc. A rapid and dramatic decrease in c-myc mRNA has been associated with the differentiation of a variety of cell types [46-48]. It is thereaore striking that in the case of the thyrocyte in which the activation of the cAMP cascade leads to both proliferation and differentiation, the kinetics of c-myc gene appear tightly controlled. After a first phase of 1 h of higher level c-myc mRNA, c-myc expression is decreased below control levels [40]. In this second phase, cyclic AMP decreases c-myc mRNA levels, as it does in proliferation inhibited fibroblasts. It even depresses EGF-induced expression [40]. The first phase could be necessary for proliferation, while the second phase could reflect the stimulation of differentiation by TSH. This down-regulation is suppressed by cycloheximide which suggests the involvement of a neosynthesized (by an autoregulatory mechanism) or a labile protein in the inhibition at the transcriptional level or at the stabilization of the mRNA.

Preliminary results indicate that TSH regulates at least at a post-transcriptional level c-myc mRNA expression: as soon as TSH is in the medium, a destabilization of c-myc mRNA is observed when transcription is blocked by actinomycin D. We therefore hypothesized that the first rise in c-myc mRNA expression reflects a very high induction of transcription combined with a destabilization mechanism. Later, the positive transcription effect is repressed and the destabilization mechanism persists, leading to a resulting down-regulation of the c-myc mRNA level. The transcription could be repressed either at the initiation [49–51] or at the elongation level [52]. It would be interesting to test whether cloned tumorigenic FRTL-5 cells, in which cyclic AMP inhibits proliferation [33], have lost the first positive control of c-myc expression. In a feedback mechanism, the neosynthetic protein could even be the c-myc protein itself, specifically modified at the post-translational level by the cyclic AMP pathway. Such an autoregulatory mechanism of blockade of transcriptional initiation requires additional transacting factors and could act as a homeostatic regulator of c-myc expression in vivo [51].

Goitrous growth and mitogenic pathways

The disease in which goiter is easiest to explain is Graves' disease. In this disease autoantibodies

directed against the TSH receptor (TSAb) activate this receptor and consequently the whole cyclic AMP cascade. At the highest concentrations reached in pathology, these TSAb do not activate, as TSH does, the Ca2+ phosphatidylinositol cascade (Laurent E et al, 1991, J Clin Invest (in press)). Thus, hyperthyroidism in Graves' disease appears to result from a chronic hyperstimulation of the cyclic AMP cascade. The effects of this cascade on cultured thyroid cells enhance function and proliferation while maintaining differentiation, ie they represent the in vitro counterparts of what is observed in Graves' diseases thyroids in vivo. It is interesting to note that, as in vitro or in vivo chronically stimulated thyroids, the growth of thyroid in Graves' disease is generally limited. This apparently simple and unicausal disease may lead in time to heterogeneous goiter. Also, in these chronically stimulated thyroids, in which proliferation and the increasing number of mitoses and bound to allow the fixation of more mutations, cancer incidence is increasing [53]. Thus, even though the cyclic AMP cascade itself maintains differentiation while promoting proliferation, the greater number of mitoses will give a higher probability of occurrence to the rare mutagenic events which lead to carcinogenesis.

The goiter resulting from congenital defects in iodine metabolism by the gland is also simply explained by classical concepts of thyroid regulation. Deficiency of thyroid hormone formation resulting from the defect relieves the thyroid hormone feedback on the hypophysis and leads to increased TSH secretion and stimulation of the thyroid. In addition, the lack in iodine metabolism, at the level of trapping or iodination will relieve the negative feedback of iodide and increase the sensitivity of the gland to the TSH growth promoting effect. Impaired iodination due to a congenital defect or to inhibition by antithyroid drugs has been shown to relieve the inhibitory effect of iodide on cyclic AMP accumulation. Defects of iodotyrosine coupling and iodotyrosine deiodination which also lead to iodine depletion will in time have the same effect. It is interesting to note that iodination defects which most severely affect the iodide inhibitory pathway lead to the severest goiters and to a great incidence of thyroid cancers.

The simplest example of a somatic mutation leading to autonomous hyperfunctioning adenomas has been demonstrated by Dr Bourne's group in the hypophysis of acromegalic rats. In the rat somatotrophs, as in dog and human thyroid cells, the activating hormone GRH acts by activating adenylate cyclase and the cyclic AMP cascade, which leads to functional activation and growth. In hyperfunctioning autonomous adenomas of somatotrophs, Landis *et al* demonstrated a mutation in Gs which causes constitutive activation of this transducing protein and consequently of the whole cyclic AMP cascade [54]. A systematic search for similar lesions in other tumors allowed the demonstration of such a mutation in

and the demonstration of seen a mathematical in the demonstratin the demonstration of seen a mathematical in t

Models of constitutive activation of the cyclic AMP mitogenic pathway

Among the mitogenic cascades, the cyclic AMP pathway is the only one that in some cell types induces both proliferation and differentiation and also activates function [4]. As shown above for the thyroid and hypophysis, its permanent stimulation leads to hypertrophied hyperfunctioning lesions. These involve the entire organ in the case of Grave's disease, where the thyroid cascade is constantly stimulated by thyroid stimulating immunoglobins, or an adenoma in cases where a somatic mutation causes constitutive activation of one element of the cascade. It might therefore be warranted at this stage to speculate about other possible somatic mutations that could cause constitutive activation of the cyclic AMP cascade and hyperfunctioning adenomas. Any permanent enhancement of a positive control or suppression of a negative control could give a selective advantage to the cells. This advantage is expressed by a higher mitotic rate and therefore a higher probability of mutations and of further progression of the cells to autonomy and tumorigenesis. However, only constitutive enhancement of a positive control could lead to autonomy from the normal regulatory feedback control (such as the thyroid pituitary feedback). If we take the thyrocytes as an example, the following steps can be considered (fig 1).

Adenylyl cyclase coupled receptors

There are several ways in which receptors could chronically activate adenylate cyclase. Overexpression of the receptor could lead to permanent stimulation if the unoccupied receptor had some basal activity or at least render the thyrocyte more sensitive to TSH and thus give it a selective advantage. Mutation of the receptor could induce or increase its constitutive activity, as has been shown for the EGF receptor in v-erb B oncogene transformation [56]. Heterotypic expression of a non-thyroid receptor (such as the ß adrenergic receptor) could make the cells independent of the normal thyroid pituitary feedback. Heterotypic expression of receptors coupled to the phosphatidylinositol cascade (serotonin 5HT1c [57], angiotensin receptor ie the mas oncogene [58]) leads to fibroblast transformation in vitro. Similarly expression of a receptor to a metabolite (eg adenosine) or a signal (eg a prostaglandin) that is constantly produced by the

cell could also cause apparently constitutive activation. Such physiologically constitutive activation has been conferred in culture on various types of cells by the expression of RDC8 gene, *ie* of the high affinity A2 adenosine receptor [59]. These causes of chronic activation would of course have to override the normal desensitization mechanism.

Cyclase activating GTP binding protein (GS)

Constitutive activation by mutational impairment of the GTPase activity of G_s or its subunit α_s would, as in the inhibition of this activity by cholera toxin, lead to permanent constitutive activation of adenylate cyclase. This type of somatic mutation, discovered by Landis in pituitary adenomas [54], has been described earlier. An inactivating mutation of the inhibitory subunit β of G_s would have the same effect. We do not know if overexpression of Gs could induce a permanent activation of adenylyl cyclase.

Cyclic AMP dependent protein kinase

Inactivating mutations of the inhibitory regulatory subunit of cyclic AMP protein kinase lead to constitutive activation of the enzyme. Such mutants have been produced in cell lines [60].

Mutations on negative controlling elements

Mutations in any negative control element could lead to activation of the affected cell and would confer a selective advantage. Negative controls which could be considered are: i), negative feedbacks in cyclic AMP action (Xi [61], phosphorylated protein inhibitors as postulated in autonomous nodules [62]); ii), the receptor-inhibitory GTP binding protein (Gi) pathway; iii), the iodide inhibitory pathway (at the level of iodide transport, oxidation or inhibitory iodinated derivative Xi synthesis) [61], etc. It is interesting in this regard that adenomas with defects in some steps in the latter pathway have been demonstrated [63]. Similar somatic mutations in cells in which cyclic AMP is a negative signal for growth would of course have the opposite effect, and would select against the affected cells.

Conclusion

The study of the control of thyroid growth has thus led us to define the stimulatory role of the cyclic AMP cascade and now allows us to predict alterations in its pathway that may cause hyperfunctioning and hyperplastic lesions.

References

- Nurse P (1990) Universal control mechanism regulating onset of M-phase. *Nature* 344, 503–508
- 2 Chambard JC, Paris S, L'Allemain G, Pouységur J (1987) Two growth factor signalling pathways in fibroblasts distinguished by pertussis toxin. *Nature* 326, 800–803
- 3 Baserga R (1985) In: The Biology of Cell Reproduction. Harvard University Press, Cambridge, USA, 251
- 4 Dumont JE, Jauniaux JC, Roger PP (1989) The cyclic AMP-mediated stimulation of cell proliferation. *Trends Biochem Sci* 14, 67–71
- 5 Gärtner R, Bechtner G, Stübner D, Greil W (1990) Growth regulation of porcine thyroid follicles *in vitro* by growth factors. *Horm Metab Res* 23 (suppl) 61–67
 6 Raspé E, Reuse S, Maenhaut C, Roger PP, Corvilain B,
- 6 Raspé E, Reuse S, Maenhaut C, Roger PP, Corvilain B, Laurent E, Mockel J, Lamy F, Van Sande J, Dumont JE (1989) Importance and variability of transducing systems in the control of thyroid cell function, proliferation and differentiation. *In: Growth Regulation of Thyroid Gland and Thyroid Tumors, Frontiers of Hormone Research*, Karger, Basel, vol 18, 1–13
- 7 Mockel J, Decaux J, Unger J, Dumont JE (1980) In vitro regulation of ornithine decarboxylase in dog thyroid slices. Endocrinolygy 107, 2069–2075
- 8 Lamy F, Willems C, Lecocq R, Delcroix C, Dumont JE (1971) Stimulation by thyrotropin *in vitro* of uridine incorporation into the RNA of thyroid slices. *Horm Metab Res* 3, 414–422
- 9 Dumont JE, Roger PP, Ludgate M (1987) Assays for the thyroid growth immunoglobulins and their clinical implications: methods, concepts and misconceptions. *Endocrine Rev* 8, 448–452
- 10 Kerkof PR, Long PJ, Chaikoff IL (1964) In vitro effects of thyrotropin hormone. On the pattern of organization of monolayer cultures of isolated sheep thyroid gland. Endocrinology 74, 170–179
- 11 Fayet G, Michel-Bechet M, Lissitzky S (1971) Thyrotropin-induced aggregation and reorganization into follicles of isolated porcine thyroid cells in culture. *Eur J Biochem* 24, 100–111
- 12 Rapoport B (1976) Dog thyroid cells in monolayer tissue culture: adenosine 3'-5'-cyclic monophosphate response to thyrotropic hormone. *Endocrinology* 98, 1189–1197
- 13 Ambesi-Impiombato FS, Parks LAM, Coon HG (1980) Culture of hormone-dependent functional epithelial cells from rat thyroids. *Proc Natl Acad Sci USA* 77, 3455–3459
- Roger PP, Hotimsky A, Moreau C, Dumont JE (1982) Stimulation by thyrotropin, cholera toxin and dibutyryl cyclic AMP of the multiplication of differentiated thyroid cells *in vitro*. *Mol Cell Endocrinol* 26, 165–176
 Roger PP, Servais P, Dumont JE (1983) Stimulation by
- 15 Roger PP, Servais P, Dumont JE (1983) Stimulation by thyrotropin and cyclic AMP of the proliferation of quiescent canine thyroid cells cultured in a defined medium containing insulin. *FEBS Lett* 157, 323–329
- 16 Roger PP, Taton M, Van Sande J, Dumont JE (1988) Mitogenic effects of thyrotropin and cyclic AMP in differentiated human thyroid cells in vitro. J Clin Endocrinol Metab 66, 1158–1165
- 17 Nitsch L, Wollman SH (1980) Thyrotropin preparations are mitogenic for thyroid epithelial cells in follicles in suspension culture. *Proc Natl Acad Sci USA* 77, 2743–2748
- 18 Fayet G, Hovsépian S (1985) Strategy of thyroid cell culture in defined media and the isolation of the Ovnis and Porthos cell strains. *In: Thyroglobulin. The Prothyroid Hormone.* Raven Press, NY, 211–224

- 19 Gärtner R, Greil W, Demharter P, Horn K (1985) Involvement of cyclic AMP, iodine and metabolites of arachidonic acid in the regulation of cell proliferation of isolated porcine thyroid follicles. *Mol Cell Endocrinol* 42, 145–155
- 20 Gerard CM. Roger PP. Dumont JE (1989) Thyroglobulin gene expression as a differentiation marker in primary cultures of calf thyroid cells. *Mol Cell Endocrinol* 61, 23–35
- 21 Eggo MC, Bachrach LR, Fayet G, Errick J, Kudlow JE, Cohen MF, Burrow GN (1984) The effects of growth factors and serum on DNA synthesis and differentiation in thyroid cells in culture. *Mol Cell Endocrinol* 38, 141–150
- 22 Westermark K, Westermark B, Karlsson FA, Ericson LE (1986) Location of epidermal growth factor receptors on portine thyroid follicle cells and receptor regulation by thyrotropin. *Endocrinology* 118, 1040–1046
- 23 Roger PP, Dumont JE (1984) Factors controlling proliferation and differentiation of canine thyroid cells cultured in reduced serum conditions: effects of thyrotropin, cyclic AMP and growth factors. *Mol Cell Endocrinol* 36, 79–93
- 24 Roger PP, Van Heuverswyn B, Lambert C. Reuse S. Vassart G, Dumont JE (1985) Antagonistic effects of thyrotropin and epidermal growth factor on thyroglobulin mRNA level in cultured thyroid cells. *Eur J Biochem* 152, 239–245
- 25 Gerard CM, Lefort A, Christophe D, Libert F, Van Sande J, Dumont JE, Vassart G (1989) Control of thyroperoxidase and thyroglobulin transcription by cAMP: evidence for distinct regulatory mechanisms. *Mol Endocrinol* 3, 2110–2118
- 26 Lamy F, Taton M, Dumont JE, Roger PP (1990) Control of protein synthesis by thyrotropin and epidermal growth factor in human thyrocytes: role of morphological changes. *Mol Cell Endocrinol* 73, 195–209
- 27 Pohl V, Roger PP, Christophe D, Pattyn G, Vassart G, Dumont JE (1990) Differentiation expression during proliferative activity induced through different pathways: *in situ* hybridization study of thyroglobulin gene expression in thyroid epithelial cells. *J Cell Biol* 111, 663–672
- 28 Roger PP, Rickaert F, Lamy F, Authelet M. Dumont JE (1989) Actin stress disruption and tropomyosin isoform switching in normal thyroid epithelial cells stimulated by thyrotropin and phorbol esters. *Exp Cell Res* 182, 1–13
- 29 Van Sande J, Lefort A, Beebe S, Roger P, Perret J, Corbin J, Dumont JE (1989) Pairs of cyclic AMP analogs, that are specifically synergistic for type I and type II cAMP-dependent protein kinases, mimic thyrotropin effects on the function, differentiation expression and mitogenesis of dog thyroid cells. Eur J Biochem 183. 699–708
- 30 Wynford-Thomas D, Smith P, Williams ED (1987) Proliferative response to cyclic AMP elevation of thyroid epithelium in suspension culture. *Mol Cell Endocrinol* 51, 163–166
- 31 Valente WA, Vitti P, Kohn LD, Brandi ML, Rotella CM, Toccafondi R, Tramontano D, Aloj SM, Ambesi-impiombato FS (1983) The relationship of growth and adenylate cyclase activity in cultured thyroid cells. Separate bioeffects of thyrotropin. *Endocrinology* 112, 71–79
- 32 Dere WH, Rapoport B (1986) Control of growth in cultured rat thyroid cells. *Mol Cell Endocrinol* 44, 195–199
- 33 Endo T, Shimura H, Saito T, Onaya T (1990) Cloning of malignantly transformed rat thyroid (FRTL) cells with thyrotropin receptors and their growth inhibition by 3',5'-

cyclic adenosine monophosphate. *Endocrinology* 126, 1492–1497

- 34 Wolf J (1969) Iodide goiter and the pharmacologic effects of excess iodide. Am J Med 47, 101–124
- 35 Pisarev MA, De Groot LJ, Wilber JE (1970) Cyclic-AMP production of goiter. *Endocrinology* 87, 339–342
- 36 Roger PP, Servais P, Dumont JE (1987) Induction of DNA synthesis in dog thyrocytes in primary culture: synergistic effects of thyrotropin and cyclic AMP with epidermal growth factor and insulin. J Cell Physiol 130, 58–67
- 37 Roger PP, Servais P, Dumont JE (1987) Regulation of dog thyroid epithelial cell cycle by forskolin, an adenylate cyclase activator. *Exp Cell Res* 172, 282–292
- 38 Contor L, Lamy F, Lecocq R, Roger PP, Dumont JE (1988) Differential protein phosphorylation in induction of thyroid cell proliferation by thyrotropin, epidermal growth factor, or phorbol ester. *Mol Cell Biol* 8, 2494–2503
- 39 Rossomondo AJ, Weber MJ, Sturgill TW (1989) Evidence that pp42, a major tyrosine kinase target protein, is MAP kinase, a mitogen-activated serine/threonine protein kinase. *Proc Natl Acad Sci USA* 86, 6940–6943
 40 Reuse S, Maenhaut C, Dumont JE (1990) Regulation of
- 40 Reuse S. Maenhaut C. Dumont JE (1990) Regulation of protooncogenes c-fos and c-myc expressions by protein tyrosine kinase, protein kinase C, and cyclic AMP mitogenic pathways in dog primary thyrocytes: a positive and negative control by cyclic AMP on c-myc expression. Exp Cell Res 33-40
- 41 Heldin NE, Paulsson Y, Forsberg K, Heldin CH, Westermark B (1989) Induction of cyclic AMP synthesis by forskolin is followed by a reduction in the expression of *c-myc* messenger RNA and inhibition of ³H-thymidine incorporation in human fibroblasts. *J Cell Physiol* 138, 17–23
- 42 Lamy F. Roger PP. Lecocq R, Dumont JE (1989) Protein synthesis during induction of DNA replication in thyroid epithelial cells: evidence for late markers of distinct mitogenic pathways. J Cell Physiol 138, 568–578
- 43 Maciel RMB, Moses AC, Villone G, Tramontano D, Ingbar SM (1988) Demonstration of the physiological role of insulin-like growth factor II in rat thyroid follicular cells in culture. J Clin Invest 82, 1546–1553
- 44 Roger PP, Reuse S, Servais P. Van Herverswyn B, Dumont JE (1986) Stimulation of cell proliferation and inhibition of differentiation expression by tumorpromoting phorbol esters in dog thyroid cells in primary culture. *Cancer Res* 46, 898–906
- 45 Müller R (1986) Cellular and viral *fos* genes: structure, regulation of expression and biological properties of their encoded products. *Biochim Biophys Acta* 823, 207–225
- 46 Heikkila R, Schwab G, Wickstrom S, Loong Loke S, Pluznik DV, Watt R. Neckers LM (1987) A c-myc antisense oligodeoxynucleotide inhibits entry into S phase but not progress from Go to G1. Nature 328, 445–449
- 47 Prochownik EV, Kukowska J, Rodgers C (1988) C-myc antisense transcripts accelerate differentiation and inhibit G1 progression in murine erythroleukemia cells. *Mol Cell Biol* 8, 3683–3695

- 48 Griep AE, Westphal H (1988) Antisense myc sequences induce differentiation of F9 cells. *Proc Natl Acad Sci USA* 85, 6806–6810
- 49 Takimoto M, Quinn JP, Farina AR, Staudt LM, Levens D (1989) Fos jun and octamer-binding protein interact with a common site in a negative element of the human c-myc gene. J Biol Chem 264, 8992–8999
- 50 Hay N, Takimoto M, Blshop JM (1989) A fos protein is present in a complex that binds a negative regulator of myc. Genes Dev 3, 293–303
- 51 Penn LJZ, Brooks MW, Laufer EM, Land H (1990) Negative autoregulation of c-mvc transcription. *Embo J* 9, 1112–1121
- 52 Miller H, Asselin C, Dufort D, Yang JQ, Gupta K. Marcu KB, Nepveu A (1989) A *cis*-acting element in the promoter region of the murine *c-myc* gene is necessary for transcriptional block. *Mol Cell Biol* 9, 5340–5349
- 53 Mazzaferri EL (1990) Thyroid cancer and Graves' disease. J Clin Endocrinol Metab 70, 826–829
- 54 Landis CA, Masters SB, Spada A, Pace AM, Bourne HR, Vallar L (1989) GTPase inhibiting mutations activate the α chain of G_s and stimulate adenylate cyclase in human pituitary tumours. *Nature* 340, 692–696
- 55 Lyons J, Landis CA, Harsh G, Vallar L, Grünewald K, Feichtinger H, Duh QY, Clark OH, Kawasaki E, Bourne HR, McCormick F (1990) Two G protein oncogenes in human endocrine tumors. *Science* 249, 655–659
- 56 Velu TJ (1990) Structure, function and transforming potential of the epidermal growth factor receptor. *Mol Cell Endocrinol* 70, 205–216
- 57 Julius D, Livelli TJ, Jessell TM, Axel R (1989) Ectopic expression of the serotonin 1c receptor and the triggering of malignant transformation. *Science* 244, 1057–1062`
- 58 Jackson TR, Blair LAC, Marshall J, Goedert M, Hanley MR (1988) The mas oncogene encodes an angiotensin receptor. Nature 335, 437–440
- 59 Maenhaut C, Van Sande J, Libert F, Abramowicz M, Parmentier M, Vanderhaegen JJ, Dumont JE, Vassart G, Schiffmann S (1990) RDC8 codes for an adenosine A₂ receptor with physiological constitutive activity. *Biochem Biophys Res Commun* 173, 1169–1178
- 60 LevinLR, Kuret J, Johnson KE, Powers S, Cameron S, Michaeli T, Wigler M, Zoller MJ (1988) A mutation in the catalytic subunit of cAMP-dependent protein kinase that disrupts regulation. *Science* 240, 68–70
- 61 Cochaux P, Van Sande J, Swillens S, Dumont JE (1987) Iodide-induced inhibition of adenylate cyclase activity in horse and dog thyroid. *Eur J Biochem* 170, 435–442
- 62 Van Sande J, Lamy F, Lecocq R, Mirkine N, Rocmans P, Cochaux P, Mockel J, Dumont JE (1988) Pathogenesis of autonomous thyroid nodules: *in vitro* study of iodine and adenosine 3',5'-monophosphate metabolism. J Clin Endocrinol Metab 66, 570–579
- 63 Demeester-Mirkine N, Van Sande J, Dor P, Heimann R, Cochaux P, Dumont JE (1984) Iodide organification difect in a cold thyroid nodule: absence of iodide effect on cyclic AMP accumulation. *Clin Endocrinol* 20, 473–479

36