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# Update on current and new potential immunotherapies in breast cancer, from bench to bedside

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Impressive advances have been seen in cancer immunotherapy during the last years. Although breast cancer (BC) has been long considered as non-immunogenic, immunotherapy for the treatment of BC is now emerging as a new promising therapeutic approach with considerable potential. This is supported by a plethora of completed and ongoing preclinical and clinical studies in various types of immunotherapies. However, a significant gap between clinical oncology and basic cancer research impairs the understanding of cancer immunology and immunotherapy, hampering cancer therapy research and development. To exploit the accumulating available data in an optimal way, both fundamental mechanisms at play in BC immunotherapy and its clinical pitfalls must be integrated. Then, clinical trials must be critically designed with appropriate combinations of conventional and immunotherapeutic strategies. While there is room for major improvement, this updated review details the immunotherapeutic tools available to date, from bench to bedside, in the hope that this will lead to rethinking and optimizing standards of care for BC patients.

#### KEYWORDS

breast cancer, immunotherapies, immunooncology, cancer treament, immune escape mechanisms

# Introduction

In the last years, there have been many advances and optimization in the treatment of breast cancer (BC). However, despite such progress, resistance to therapy and disease relapse remain important challenges in the management of BC in a considerable proportion of patients. Specifically, impressive advances have been seen in cancer immunotherapy during the last decade. Cancer immunotherapy exploits the host's immune system to eradicate tumor cells. Although BC has long been considered a non-immunogenic process,

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immunotherapy for the treatment of BC is emerging as a new therapeutic approach with considerable potential, supported by a plethora of completed and currently ongoing preclinical and clinical studies in various types of BC immunotherapies. Tumor-infiltrating lymphocytes (TILs) are more commonly found in Human epidermal growth factor receptor 2 (HER-2)-positive BC and triple negative BC (TNBC), where the median percentages are 15% and 20%, respectively (1). However, 10% of TILs are also found in hormone receptor (HR)-positive BC (1). TILs can specifically target tumor cells following activation by antigen presenting cells (APC) via tumor antigen peptide presentation to human leukocyte antigen (HLA) molecules. TILs are associated with a better prognosis in TNBC (2) and node-positive TNBC (1). In HER2-positive BC, the presence of TILs showed contradictory data regarding trastuzumab therapy benefit (3, 4). TILs have also been associated with a higher probability of pathological complete response (pCR) in neoadjuvant settings (5, 6). Likewise, in HRpositive BC, CD8+ T-cell infiltration has been associated with survival (7), although this is currently under debate since contradictory results have been found for this BC subtype in neoadjuvant (8, 9) and adjuvant (10) settings. This led to suggest that the tumor-eradicating properties of TILs are an efficient part of the antitumor immune response and could therefore be exploited as immunotherapy to improve the clinical outcome of BC patients.  $\gamma\delta$ T-cells and natural killer (NK) cells have also been associated with a better prognosis in all BC subtypes (11, 12). Many targets are constantly being discovered on antitumor lymphoid cells, such as immune checkpoints. In addition, other immune cells of the tumor microenvironment (TME) contribute positively or negatively to the antitumor immune response and are currently a topic of intense preclinical and clinical research, such as tumor-infiltrating myeloid cells (13). Herein, we summarize the current and new potential immunotherapeutic strategies showing promising results in the emerging field of BC immunotherapy. We provide a basis for reflection on the available immunotherapeutic tools to date in the hope that this will lead to rethinking and optimizing standards of care for BC patients.

# Directed monoclonal antibodies

HER2 is overexpressed in 15-20% of BC and correlates with higher grade, aggressive phenotype, and poor clinical outcome. Immunotherapies in the form of monoclonal antibodies specifically binding to HER-2 receptor, added to chemotherapy, are the cornerstone for HER-2-overexpressing BC therapy and have led to significant improvements in HER2-positive BC prognosis compared to previous chemotherapy regimens. Trastuzumab has been approved for the treatment of HER2-positive BC patients for approximately the past 20 years and acts through several mechanisms of action. It suppresses the HER2 intracellular signaling pathway by binding to the transmembrane HER2 receptor, which is followed by its internalization, degradation, and downregulation of PI3K pathway. In addition, trastuzumab activates both the innate and adaptive immune systems. Indeed, this monoclonal antibody enhances antibody-dependent cellular cytotoxicity (14), antibody-dependent cellular phagocytosis and macrophage activation (15), Fc-mediated immune priming by dendritic cells (DCs) (16), effector HER-2-specific T cell response (17) and memory T-cell response (16) (Figure 1). These mechanisms seem to be critical for the induction of a pCR after neoadjuvant therapy in HER2-positive BC patients (18). Pertuzumab is a dual HER2/HER3 monoclonal antibody approved in combination with trastuzumab and taxane-based chemotherapy for first-line therapy in HER2-positive metastatic BC and in adjuvant and neoadjuvant settings. It works by blocking HER2 heterodimerization and may act by promoting an antitumor immune response (19), although data regarding its mechanisms of action and its synergism with trastuzumab is still limited. To improve the efficacy of trastuzumab, the immunogenic properties of trastuzumab may be exploited in association with other strategies. For example, margetuximab was approved in combination with chemotherapy for third-line therapy in metastatic HER2-positive BC disease. Margetuximab is a monoclonal antibody similar to trastuzumab, whose modified Fc fragment has a much greater affinity for its activating Fcy receptors and a decreased affinity for its inhibitory Fcy receptors on tumorinfiltrating NK cells and macrophages. This way margetuximab promotes antibody-dependent cellular cytotoxicity and phagocytosis processes against tumor cells. This may explain the influence of Fcy receptor polymorphism on overall survival of BC patients treated with margetuximab compared to trastuzumab (20). In light of the success of anti-HER2 therapies in HER2-positive BC, one might wonder why other monoclonal antibodies targeting tumor antigens other than HER2 have not been developed for other BC subtypes. Such monoclonal antibodies, synergizing with the potential antitumoral properties of the TME, might be particularly efficient in BC tumors wherein myeloid cells are abundant.

# Antibody-drug conjugates

Antibody-drug conjugates (ADC), a new emerging class of antineoplastic agents with a high therapeutic index and impressive clinical efficacy, display both immune mechanisms of action, like those of naked directed monoclonal antibodies, combined with the targeted delivery of chemotherapy directly to antigen-expressing tumor cells (21). They are therefore known as "biological missiles". Recently, other mechanisms of action have been suggested to contribute to both their antitumor activity and adverse events (such as thrombocytopenia). For example, the release of chemotherapy into the TME may lead to the recruitment of particularly immunosuppressive, protumoral and tissue repairing myeloid cells. ADC may be taken up by macrophages through Fcy receptors, leading to myeloid cell depletion or modulation of the activation state (22). Chemotherapy may also deplete regulatory T cells by diffusing into the TME through a bystander effect. Chemotherapy may also further promote NK cellmediated antibody-dependent cellular cytotoxicity (ADCC) or regulate tumor antigen presentation by DCs (Figure 2). Trastuzumab emtansine is an ADC of trastuzumab covalently linked to the cytotoxic agent emtansine (DM1/maytansinoid). It is approved for HER2-positive BC



patients in the metastatic setting and in the adjuvant setting for HER2positive BC patients with residual invasive disease following neoadjuvant trastuzumab and chemotherapy. Trastuzumabderuxtecan is the following ADC of trastuzumab linked to the topoisomerase 1 inhibitor deruxtecan (DXt/camptothecin). While TDM-1 has a non-cleavable linker and an average of 3.5 molecules of payloads per antibody, trastuzumab deruxtecan displays a cleavable linker with 8 molecules of a different payload. These differences could affect their antitumor mechanisms of action, such as the bystander effect and cellular toxicity. The efficacy and safety of trastusumab deruxtecan was compared with trastuzumab emtansine in the DESTINY-BREAST03 phase 3 randomized clinical trial, showing a lower risk of disease progression or death (23). Trastuzumab deruxtecan received accelerated approval in 2019 and has now become the new standard of care for second-line therapy in HER2positive BC patients who have received a prior anti-HER2 based regimen either in the metastatic setting, or in the neoadjuvant or adjuvant setting and have developed disease recurrence during or within 6 months of completing therapy. In addition, it is approved for locally advanced or metastatic HER2-low (IHC 1+ or IHC 2+/FISH-) BC patients who have received a prior chemotherapy in the metastatic setting or developed disease recurrence during or within six months of completing adjuvant chemotherapy. Moreover, trastuzumab deruxtecan is currently compared, when used with or without pertuzumab, to the standard of care which is taxane, pertuzumab and trastuzumab as first-line treatment in the DESTINY-BREAST09 phase 3 clinical trial. Trastuzumab deruxtecan in association with tucatinib is also currently studied in metastatic BC patients, including with active brain metastasis, in the HER2-CLIMB-04 phase 2 clinical trial. Another ADC, sacituzumab govitecan, targets the human trophoblast cell-surface antigen 2 (Trop-2), which is highly expressed in BC, and is coupled with a high drug-to-antibody ratio to SN-38, the active metabolite of irinotecan. This leads to the delivery of high concentrations of the chemotherapy to the tumor cells by intracellular



uptake of SN-38, thereby also allowing the cells of the TME to be eradicated by SN-38 which is released extracellularly from the tumor cells through a bystander effect. This antitumor effect may also be mediated by a significant antibody-dependent cellular cytotoxicity effect against the Trop-2-positive tumor cells (24). Sacituzumab govitecan received accelerated approval in 2020 for the treatment of refractory metastatic TNBC following at least two prior chemotherapies, by showing promising results for this notoriously difficult-to-treat group of patients (25). Recently, it has demonstrated extended progression-free survival (PFS) and overall survival (OS), as well as greater health-related quality of life benefits than chemotherapy, and moved to second-line therapy of TNBC (at least one in the metastatic setting) (25). It may also represent a new option for endocrine-resistant hormone receptor-positive/HER2-negative metastatic BC, since it has recently shown a longer PFS and a statistically significant OS compared to standard chemotherapy (capecitabine, eribulin, vinorelbine or gemcitabine) after CDK4/6 inhibitors and 2 to 4 previous lines of chemotherapy (26). The indication of sacituzumab govitecan is currently investigated in case of residual disease after neoadjuvant chemotherapy.

### Immune checkpoint inhibitors

During the last decade, the emergence of immune checkpoint inhibitors (ICI) has revolutionized the field of cancer therapies, especially in advanced or metastatic cancers where they have shown unprecedented and durable efficacy. They are approved in many different cancer types such as lung cancer, melanoma, renal cell carcinoma, and in any high microsatellite instability or mismatch repair deficiency. Used alone or in combination with other ICI or chemotherapies, they represent a staggering proportion of the ongoing clinical trials in oncology. In all cancer types, the most widely studied immunotherapeutic agents to date are ICI blocking cytotoxic T lymphocyte-associated molecule-4 (CTLA-4), programmed cell death receptor-1 (PD-1) and programmed cell death ligand-1 (PD-L1). While PD-1 is mainly expressed on TILs, PD-L1 is expressed on both cancer cells and tumor-infiltrating immune cells. Although the impact of ICI on the immune response remains to be fully elucidated, the PD-1/PD-L1 immune inhibitory axis is thought to be upregulated in the TME and to impair the effector stage of the antitumor immune response (27).

# PD-1 inhibitors

Nivolumab and pembrolizumab are human monoclonal antibodies that block PD-1 and therefore the interaction of PD-1 with its ligand PD-L1, preventing T-cell suppression.

In TNBC, although pembrolizumab showed antitumor activity in the phase 1b KEYNOTE-012 and the phase 2 KEYNOTE-086 trials, the KEYNOTE-119 phase 3 trial comparing pembrolizumab with

chemotherapy did not show significant improvement in OS in the second or third-line treatment of patients with metastatic TNBC (28). However, in the KEYNOTE-355 phase 3 study, pembrolizumab combined with chemotherapy significantly improved PFS compared with chemotherapy alone in patients with advanced or metastatic PD-L1-positive TNBC (CPS≥10) (29). Moreover, the follow-up of the patients with CPS of 10 or more showed a significantly longer OS with no new safety signals identified when pembrolizumab was added to chemotherapy (30). In addition, among patients with previously untreated stage II or III TNBC, the rate of pCR at definitive surgery was higher in pembrolizumab plus neoadjuvant chemotherapy compared with placebo plus chemotherapy in the KEYNOTE-522 phase 3 trial, along with disease-free survival (DFS) (31). Based on these results, the FDA approved in 2021 pembrolizumab for high-risk, earlystage, TNBC, in combination with chemotherapy as neoadjuvant treatment, and then continued as a single agent as adjuvant treatment. The FDA also granted accelerated approval in 2020 to pembrolizumab in combination with chemotherapy for patients with locally advanced or metastatic TNBC whose tumors express PD-L1. Other treatments with the combination of pembrolizumab and eribulin showed promise for patients with metastatic TNBC, with efficacy that seems greater than reports of either drug alone, according to the KEYNOTE-150 phase 1b/2 study (NCT02513472) (32). A pilot study comparing nivolumab with capecitabine and with combination therapy as adjuvant therapy after residual disease following neoadjuvant chemotherapy is under investigation (NCT03487666).

In HER2-positive BC, the combination of pembrolizumab plus trastuzumab demonstrated a tolerable safety profile, activity and durable clinical benefit, in advanced HER-2-positive, trastuzumab-resistant, PD-L1-positive BC disease (33).

In HR-positive BC patients, fewer clinical trials have been performed so far. In a phase 1b study, pembrolizumab was well tolerated with modest but durable partial response in certain patients with previously treated, advanced, PD-L1-positive HR-positive HER2-negative BC (34). In women with early-stage, high-risk, HR-positive HER2-negative BC, an ongoing phase 2 trial in the neoadjuvant setting with pembrolizumab in association with standard chemotherapy showed improved pCR (35). Since CDK4/6 inhibitors induce, in addition to a tumor cell cycle arrest, an enhanced antitumor immune response (36), they may be used in synergy with ICI to increase tumor immunogenicity (NCT02648477). Other studies currently assessing PD-1 inhibitors in different settings are summarized in Table 1.

## **PD-L1** inhibitors

Atezolizumab, durvalumab and avelumab are human monoclonal antibodies that block PD-L1 and therefore PD-1/PD-L1 interaction, T-cell activation and proliferation.

In TNBC, the association of durvalumab with nab-paclitaxel and doxorubicin-cyclophosphamide neoadjuvant chemotherapy suggested a high rate of pCR in a phase 1/2 trial (43), which was higher in PD-L1-positive and TILs-high than PD-L1-negative patients (44). This has not been observed in the phase 3 trial comparing the addition of atezolizumab to carboplatin and nabpaclitaxel with the chemotherapy regimen alone in a neoadjuvant setting (40). However, the IMpassion130 phase 3 study demonstrated a prolonged PFS with atezolizumab plus nabpaclitaxel in metastatic TNBC patients (37) and a clinically meaningful OS benefit in previously untreated PD-L1-positive patients, compared with placebo plus nab-paclitaxel (38). Following this clinical trial, nab-paclitaxel with atezolizumab received accelerated approval in March 2019 for first-line treatment of locally advanced or metastatic TNBC whose tumors express PD-L1 (>=1%). However, this has been withdrawn after IMpassion131 clinical trial results showing that atezolizumab and paclitaxel under the same settings did not show any improvement of PFS or OS versus paclitaxel alone (39).

The addition of atezolizumab to TDM-1 did not show any improvement and was associated with more adverse events in previously treated HER2-positive locally advanced or metastatic BC patients who received prior trastuzumab and taxane based therapies. However, a benefit in terms of PFS in favor of the combination has been observed in the PD-L1-positive subgroup of patients (45). Further study is required in subpopulations of patients.

Further results from various clinical trials investigating anti-PD-L1 treatments in TNBC, HER2-positive or HR-positive BC patients are summarized in Table 1 (41, 46).

## **CTLA-4** inhibitors

CTLA-4 on TIL surface mediates T-cell suppression by binding to CD80 and CD86 (expressed on the surface of antigen-presenting cells), therefore competing with the co-stimulatory receptor CD28 on the cell surface of T cells (47). Ipilimumab is a human monoclonal antibody that targets CTLA-4, thus preventing its inhibitory effect on T-cell activation. Ipilimumab is under investigation in various settings in BC which are summarized in Table 1 (42).

### Optimizing trial design

Major clinical trials, such Keynote-119 trial assessing pembrolizumab monotherapy, failed to improve OS versus singledrug chemotherapy per investigator's choice after first-line metastatic TNBC. This underscores the inefficacy of PD-1 inhibitors alone and suggests the association with chemotherapies for the next trials. Indeed, various chemotherapies (such as anthracyclines or taxanes) result in tumor cell death and debris which induce immunogenic cell death. Immunogenic cell death is mediated by damage-associated molecular patterns (DAMPs), promoting tumor phagocytosis and antigen presentation, and may facilitate the induction of a robust antitumor immune response by ICI. In contrast, Keynote-355 showed improvement of PFS and OS in PD-L1-positive metastatic TNBC when chemotherapies including platinum or taxanes were associated with PD-1 inhibitors. This suggests the selection of such chemotherapies combined with ICI for further trials. On an early setting, Keynote-522 showed improved pCR when neoadjuvant and adjuvant pembrolizumab was combined with neoadjuvant carboplatin-

#### TABLE 1 Published and ongoing clinical trials related to ICI.

TNBC	Setting	Regimens	Results	Safety data	references
KEYNOTE-119 phase 3	Metastatic	pembro 200mg 1x/3 weeks vs investigator's choice chemo (capecitabine, eribulin, gemcitabine, or vinorelbine)	OS ≈	Anemia and leupopenia ∖ with pembro vs chemo. AE of grade≥3 in 20% of both groups	(28)
KEYNOTE-355 phase 3	Metastatic	Chemo (nab-paclitaxel, paclitaxel, or gemcitabine/ carboplatin)+pembro 200mg 1x/3 weeks vs placebo	<ul> <li>✓ PFS (≥CPS</li> <li>10)</li> <li>✓ OS (≥CPS</li> <li>10): 23 vs</li> <li>16.1 months</li> </ul>	68.1% of AE of grade≥3 in pembro +chemo vs 66.9% in chemo	(29, 30)
KEYNOTE-522 phase 3	Early (neoadjuvant and adjuvant)	Chemo (paclitaxel+carboplatin)+4 cycles of neoadjuvant pembro 200mg 1x/3 weeks vs placebo. Additional four cycles of pembro vs placebo and then doxorubicin +cyclophosphamide or epirubicin+cyclophosphamide for both groups. Adjuvant pembro vs placebo.	<ul> <li>✓ DFS</li> <li>✓ pCR:</li> <li>64.8% vs 51.2%</li> </ul>	78% of AE of grade≥3 in pembro+chemo vs 73% in chemo	(31)
KEYNOTE-150 phase 1b/2	Metastatic	Pembro 200mg d1+eribulin 1.4mg/m <sup>2</sup> d1, d8/21	<ul> <li>✓ ORR in PD- L1+ tumors:</li> <li>28.4% vs 17.3%</li> </ul>	26.3% of neutropenia of grade≥3	(32)
IMpassion130 phase 3	Metastatic	Nab-paclitaxel 100mg/m² d1,8,15/28+atezolizumab 840mg d1, d15 vs placebo		48.7% of AE of grade≥3 in the atezo group vs 42.2% in placebo, 57,3% of potential immune -related AE vs 41.8%, respectively	(37, 38)
IMpassion131 Phase 3	Metastatic	Paclitaxel 90mg/m² d1,8,15/28+atezolizumab 840mg d1 and d15 vs placebo	PFS ≈ OS ≈	53% of AE of grade≥3 in atezo group vs 46% in placebo. Higher incidence of low-grade hypo and hyperthyroidism in atezo group	(39)
Pilot study	Early (residual disease)	adjuvant capecitabine 1250mg/m <sup>2</sup> bid d1-d14/21 or nivolumab 360mg 1x/3 weeks or both	Immunoscore change?	?	NCT03487666
Phase 2	Metastatic	carboplatin+nivolumab 360mg 1x/3 weeks vs placebo	PFS?	?	NCT03414684
Phase 3	Metastatic	Chemotherapies+pembrolizumab 200mg 1x/3 weeks vs placebo	PFS? OS?	?	NCT02819518
Phase 1/2	Metastatic	Niraparib up to 300mg/day d1-21+pembrolizumab 200mg d1/21	DLT? ORR?	Ś	NCT02657889
IMpassion030 Phase 3	Early (adjuvant)	paclitaxel 80mg/m <sup>2</sup> 1x/week for 12 weeks followed by dose-dense doxorubicin 60mg/m <sup>2</sup> or epirubicin 90mg/ m <sup>2</sup> +cyclophosphamide 600mg/m <sup>2</sup> 1x/2weeks for 4 doses +atezolizumab 840 mg 1x/2 weeks for 10 doses and then maintenance 1200 mg 1x/3weeks to complete 1 year vs placebo	iDFS?	?	NCT03498716
NeoTRIP Phase 3	Early (neoadjuvant)	Neoadjuvant carboplatin+nab-paclitaxel 125mg/m <sup>2</sup> d1 and d8+atezolizumab 1200mg 1x/3weeks for 8 cycles vs placebo, followed by adjuvant anthracyclines for 4 cycles	pCR ≈ EFS?	Liver transaminase abnormalities in atezo group	NCT02620280 (40)
IMpasision031 phase 3	Early (neoadjuvant)	Nab-paclitaxel 125mg/m <sup>2</sup> 1x/week+atezolizumab 840mg 1x/2weeks for 12 weeks followed by atezolizumab 840mg +doxorubicin 60mg/m <sup>2</sup> +cyclophosphamide 600mg/m <sup>2</sup> 1x/2weeks for 4 doses followed by adjuvant atezolizumab 1200mg 1x/3weeks for 11 doses vs placebo	✓ pCR in all- randomized population and PD-L1+ status	30% of AE of grade≥3 in atezo group vs 18% in placebo, such as febrile neutropenia, pneumonia and pyrexia	NCT03197935 (41)

(Continued)

#### TABLE 1 Continued

TNBC	Setting	Regimens	Results	Safety data	references
Phase 3	Early relapsing metastatic	Chemotherapy (capecitabine or gemcitabine/carboplatin) and atezolizumab 1200mg 1x/3weeks vs placebo	OS?	?	NCT03371017
Phase 3	Early (adjuvant)	Adjuvant avelumab 10mg/kg 1x/2 weeks vs placebo	DFS? OS?	?	NCT02926196
Phase 2	Stage III (neoadjuvant)	Neoadjuvant ipilimumab 1mg/kg 6 weekly for 2 doses +nivo 240mg every 2 weeks for 6 weeks+weekly paclitaxel 80mg/m <sup>2</sup> for 12 weeks+postoperative nivolumab 480 mg 4-weekly for further 9 months	pCR 37.5% in PD-L1+ and 23% in PD-L1- , 57.6% of ORR	?	ongoing (42)
GeparNuevo Phase 2	Early (neoadjuvant)	Nab-paclitaxel 125mg/m <sup>2</sup> 1x/week+durvalumab 1.5g 1x/4 weeks vs placebo followed by doxorubicin 60mg/ m <sup>2</sup> +cyclophosphamide 600mg/m <sup>2</sup> +durvalumab vs placebo	pCR ≈ /iDFS 85.6% in durva group vs 77.2% in placebo /DDFS 91.7% in durva group vs 78.4% in placebo /OS 95.2% in durva group vs 83.5% in placebo		NCT02685059
HER2+ BC	Setting	Regimens	Results	Safety data	references
PANACEA Phase 1b/2	Metastatic	Trastuzumab 6mg/kg+pembro 200mg 1x/3 weeks vs placebo	OR of 15% in PD-L1+ tumors, 0% in PD-L1- tumors	29% of AE of grade≥3. Immune-related AE in 19% such as thyroid dysfunction, pneumonitis and autoimmune hepatitis.	(33)
KATE2 Phase 2	Metastatic	TDM-1 3.6mg/kg+atezolizumab 1200mg 1x/3 weeks vs placebo	$PFS \approx$ In PD-L1+ tumors: PFS 8.5 months in atezo group vs 4.1 months in placebo	19% of AE of grade≥3 in atezo group vs 3% in placebo	(43)
Phase 2	Early (neoadjuvant)	Neoadjuvant pembro 200mg+trastuzumab 6mg/kg +pertuzumab 420mg/kg 1x/3 weeks	pCR?	?	NCT03988036
Phase 2	Metastatic	5 administrations of pembro 200mg $1x/3$ weeks or VRP- HER2 vaccine $4x10EE8$ IU $1x/2$ weeks for 3 injections or both of them	Anti-HER2 TILs and antibodies?	?	NCT03632941
Phase 3	Metastatic	Paclitaxel 1x/week or docetaxel 1x/3 weeks+ trastuzumab +pertuzumab+atezolizumab 1x/3 weeks vs placebo for 2 years	PFS? OS?	?	NCT03199885
IMpassion050 Phase 3	Early	Neoadjuvant doxorubicin 60mg/m²+cyclophosphamide 600mg/m²+atezolizumab 840 mg 1x/2 weeks vs placebo followed by paclitaxel 80mg/m²1x/week for 12 weeks +trastuzumab 6mg/kg+pertuzumab 420mg 1x/3weeks. Adjuvant to complete up to 1 year HER2-target therapy +atezolizumab 1200mg 1x/3 weeks vs placebo	pCR ≈ DFS? OS?	Neoadjuvant: 47.3% of AE of grade≥3 in atezo group vs 42.2% in placebo Adjuvant: 13.4% of AE of grade≥3 in atezo group vs 9.8% in placebo	NCT03726879 (44)
Phase 2a	Metastatic	Atezolizumab+paclitaxel+trastuzumab+pertuzumab	Antitumor activity (RECIST)?	3	NCT03125928
Phase 1b	Metastatic	Durvalumab+trastuzumab 1x/3 weeks	Recommended phase 2 dose?	?	NCT02649686

(Continued)

#### TABLE 1 Continued

Luminal	Setting	Regimens	Results	Safety data	references
I-SPY2 Phase 2	Early (neoadjuvant)	neoadjuvant paclitaxel 80 mg/m <sup>2</sup> +pembrolizumab 200mg 1x/3 weeks vs placebo followed by doxorubicin 60 mg/ m <sup>2</sup> +cyclophosphamide 600 mg/m <sup>2</sup> . Standard-of-care adjuvant therapy.	pCR 30% vs 13% for HR +HER2- and 60% vs 22% for TNBC	immune-related AE such as thyroid dysfunction $(1,4\% \ge 3)$ and adrenal insufficiency $(7,2\%)$ of $\ge 3$ )	NCT01042379 (35)
Phase 2	Metastatic	Pembrolizumab 1x/3 weeks for 24 months+aromatase inhibitor po QD	ORR? OS?	?	NCT02648477
Phase 2	Early (residual disease)	Adjuvant pembrolizumab 1x/3 weeks up to 24 months +hormone therapy	DFS? OS	?	NCT02971748
KEYNOTE-756 phase 3	Early (neoadjuvant)	Neoadjuvant pembrolizumab 200mg 1x/3 weeks vs placebo+paclitaxel 80mg/m <sup>2</sup> 1x/week followed by doxorubicin 60mg/m <sup>2</sup> or epirubicin 100mg/ m <sup>2</sup> +cyclophosphamide 600mg/m <sup>2</sup> 1x/2 weeks. Adjuvant pembrolizumab 200mg 1x/3 weeks vs placebo +hormone therapy	pCR? EFS?	?	NCT03725059
Phase 3	Metastatic	pembrolizumab 200mg 1x/3 weeks vs placebo +chemotherapy (paclitaxel, nab-paclitaxel, liposomal doxorubicin or capecitabine)	PFS? OS?	Ś	NCT04895358
Phase 2	Early (neoadjuvant)	Tremelimumab 3mg/kg+exemestane 25mg daily followed by durvalumab 20mg/kg 1x/4weeks+exemestane 25mg daily	CD8+ T cells? pCR?	?	NCT02997995
Phase 2	Metastatic	Atezolizumab 840mg 1x/2 weeks+cobimetinib daily	ORR?	Ś	NCT03566485
Phase 2b	Metastatic	Chemotherapy (pegylated liposomal doxorubicin 20mg/ m <sup>2</sup> 1x/2 weeks+cyclophosphamide 50mg daily first 2 weeks in each 4 week cycle)+ibilimumab 1mg 1x/6 weeks and nivolumab 240mg 1x/2 weeks vs placebo	PFS?	?	NCT03409198
Phase 2	Metastatic (hypermutated)	Nivolumab 1x/2 weeks+ibilimumab 1x/6 weeks	ORR?	?	NCT03789110
Phase 2	Metastatic	sacituzumab govitecan 10mg/kg 2x/3 weeks +pembrolizumab 1x/week vs placebo	PFS? ORR? OS?	?	NCT04448886
Phase 3	Early	neoadjuvant chemotherapy (paclitaxel followed by anthracycline+cyclophosphamide) and adjuvant hormonotherapy of investigator's choice+neoadjuvant and adjuvant nivolumab vs placebo	pCR?	?	NCT04109066

paclitaxel followed by anthracycline-based chemotherapy. The IMPassion031 trial combining atezolizumab to nab-paclitaxel followed by atezolizumab with anthracycline-based chemotherapy also significantly increased pCR rate. This further suggests the synergy of ICI with anthracyclines or taxanes. The Impassion130 trial showed increased PFS when atezolizumab was added to nab-paclitaxel chemotherapy in metastatic previously untreated PD-L1-positive TNBC. The trial received accelerated FDA approval which was later withdrawn due to lack of benefit. Accumulating data suggests that immune checkpoint inhibitor antitumor activity is mediated by a non-specific  $Fc\gamma$ R-mediated modulation of the TME, and not only by the blockade of PD-1/PD-L1 axis on TILs. For example, anti-CTLA4 antibodies may activate  $Fc\gamma$ R-mediated elimination of intratumoral

regulatory T cells (48) and anti-PD-L1 antibodies may activate Fc $\gamma$ Rmediated reprogramming of tumor-infiltrating myeloid and NK cells (49). This highlights the role of the Fc domain of therapeutic ICI antibodies in their antitumor activity. The albumin found in nabpaclitaxel (albumin-bound paclitaxel, Abraxane) may prevent the nonspecific binding of the ICI antibodies (through Fc $\gamma$  receptors). Indeed, "blocking solutions", such as bovine serum albumin, are commonly used in laboratories to prevent such non-specific antibody bindings. This might explain the lack of antitumor activity of ICI associated with nab-paclitaxel and suggest replacing nab-paclitaxel by other chemotherapies in the next clinical trials. The involvement of nabpaclitaxel in parallel with atezolizumab in a neoadjuvant setting, such as in the neoTRIP trial, also showed disappointing results up to now, regarding pCR in TNBC patients. In the IMPassion131 trial, adding atezolizumab to paclitaxel did not significantly improve PFS, even in the PD-L1-positive TNBC patients. It should be noted that, according to *post hoc* analyses, most patients were administered corticosteroid premedication throughout paclitaxel therapy, which might alter immunotherapy efficacy. Nevertheless, although the Kaplan-Meier curves remained overlapping for the first 7 months, the trend began to diverge thereafter, favoring the atezolizumab arm and thus, suggesting that the impact of ICI may begin later on, and therefore the need for a longer follow-up.

Regarding HER2-positive BC, HER2-directed monoclonal antibodies, such as trastuzumab, already significantly improved HER2-positive BC prognosis and are probably potent tools for nextgeneration immunotherapies. Indeed, these antibodies have multiple mechanisms of action, as described above. Therefore, the addition of ICI to trastuzumab/pertuzumab seems to be a promising combination that can at the same time target the tumor cells (through antibodydependent cellular cytotoxicity and antibody-dependent cellular phagocytosis by anti-HER2 therapy) and trigger myeloid and lymphoid compartment activation (through anti-HER2 therapy and immune checkpoint inhibitor). Associated chemotherapies may further promote tumor immunogenicity. For example, in a metastatic setting, the phase 1b/2 PANACEA trial showed encouraging results, although associated usual chemotherapies were unfortunately missing and should be investigated. ADC are expected to deliver a toxic payload specifically to antigen-expressing-tumor cells. However, ADC have also been clinically shown to be effective in low antigen expression, whereas trastuzumab is not effective. This highlights other mechanisms of action of ADC which might be antigen-independent, as described above. While these mechanisms of action are not yet clear today, combination therapies including ADC are even more difficult to suggest. In phase 2 KATE2 trial, investigating TDM-1 combined with atezolizumab, there is a trend in increased PFS in favor of atezolizumab only in the PD-L1-positive tumors. Adding chemotherapy to ADC and ICI might facilitate the inflammatory response. On the other hand, chemotherapy might suppress immune cells from the TME and by this way alter this inflammatory response. Clinical trials combining ADC with chemotherapy and/or ICI are ongoing (NCT04538742)(NCT04556773) and will first have to assess the adverse events of such combinations. In early HER2-positive BC disease, phase 3 IMpassion050 trial has investigated the association of atezolizumab to anthracycline-based chemotherapy followed by paclitaxel, trastuzumab and pertuzumab in a neoadjuvant setting. Although the primary endpoint has not been reached (pCR was not improved), the neoadjuvant period was short. We should certainly wait a bit longer to observe the known long-term impact of immunotherapy by checking EFS - which is a secondary end point of this study. Indeed, following the administration of the treatments, the mature dendritic cells migrate to the draining lymphoid organs, activate antitumor tumor T cells, which proliferate and go back to the tumor site to kill specifically the cancer cells. The common concept of pCR, which is associated with an important prognosis factor after chemotherapy, might therefore be very different in an immunotherapy setting and we are not sure if pCR is a proper primary endpoint to choose. To optimize the trial design, we would suggest adding atezolizumab to taxanes (with or without platins) and HER2 blockade (monotherapy or dual therapy) neoadjuvant sequence, and not to the anthracyclinebased neoadjuvant chemotherapy sequence. Indeed, this may allow the combination of chemotherapy, anti-HER2 blockade and ICI in parallel, whose mechanisms of actions might synergize.

In luminal BC, fewer trials involving immunotherapies have been performed so far. While the first trials were disappointing, many results of ongoing trials are awaited. In high-risk early-stage luminal HER2-negative BC, the pCR rate increased from 13% to 30% in the phase 2 adaptively randomized I-SPY2 trial by adding pembrolizumab to neoadjuvant paclitaxel chemotherapy followed by anthracyclinebased chemotherapy. This was associated with a high EFS rate of 93% at 3 years. This chemo-immunotherapy combination is similar to those that are effective in the other BC subtypes, and might succeed in the randomized phase 3 Keynote 756. In metastatic luminal BC, results are encouraging as well (50). While a previous trial adding pembrolizumab to eribulin did not improve PFS or OS in luminal metastatic BC patients pretreated with 2 or more lines of hormonal therapy (51), there are many other combination therapies to investigate. For example, combinations of ICI with aromatase inhibitors and CDK 4/6 inhibitors might synergize. Indeed, CDK 4/ 6 inhibitors induce the presentation of tumor antigens on dendritic cells, stimulate cytotoxic T cells and suppress regulatory T cells activity (52). The impact of hormone therapies on the antitumor immune response is less clear, but aromatase inhibitors might improve CD8 T cell/regulatory T cell ratio (53).

#### New immune checkpoint targets

Immune checkpoint discovery has increased interest in other recently discovered inhibitory and stimulatory immune checkpoint pathways and gave rise to new clinical trials with many other potential immune checkpoint targets (Figure 3). For example, Lymphocyte Activation Gene-3 (LAG-3) signaling pathway is mainly expressed on lymphocytes where it inhibits T-cell activation, proliferation and cytokine production (54). The combination of effilagimod  $\alpha$  (a monoclonal antibody inhibiting LAG-3) with paclitaxel showed clinical benefit in 90% of metastatic BC patients at 6 months, supporting the future development of this agent in combined firstline regimens (55). Ongoing clinical studies investigate its safety and efficacy alone, associated with other immunotherapies, or through a bispecific antibody targeting ICI and LAG-3 simultaneously (NCT02614833)(NCT03742349)(NCT03849469). T-cell Immunoglobulin and Mucin domain-containing protein 3 (TIM-3) contains multiple co-inhibitory receptors expressed on different types of immune cells (56). TIM-3 antagonistic monoclonal antibodies are currently being clinically investigated in advanced tumors including BC, alone or with a co-blockade with other immunotherapies or chemotherapies (NCT02817633)(NCT04370704)(NCT05287113) (NCT03446040). T-cell Immunoreceptor with Ig and ITIM domains (TIGIT) is restricted to lymphocytes where it exerts its immunosuppressive effect by competing with costimulatory signals such as CD226, like the CD28/CTLA-4 pathway (57). Monoclonal antibodies targeting TIGIT undergo early-stage clinical trials as monotherapy or in combination with current ICI in patients with locally advanced or metastatic malignancies including BC



(NCT03628677). V-domain Immunoglobulin Suppressor of T-cell Activation (VISTA) is expressed on myeloid cells, regulatory T cells, and on a lesser extent, on BC tumor cells (58). Preclinical trials of VISTA monoclonal antibody-mediated blockade showed remarkable protective antitumor effects (59). CA-170 is an orally available dual small molecule inhibitor of VISTA and PD-L1 being examined in patients with advanced tumors such as BC (NCT02812875). B7 Homolog 3 protein (B7-H3) displays complex immunomodulatory activity in innate and adaptive immunity with costimulatory as well as inhibitory functions and still requires further elucidation. Its receptor has not yet been identified (60). The blockade of B7-H3 is being investigated in early-stage clinical trials in various cancers including BC (NCT03729596)(NCT04145622)(NCT03406949). OX40 and OX40 ligands are co-stimulatory molecules expressed on different types of

immune cells whose interaction leads to T-cell proliferation and decreased regulatory T cells. Although this axis seems to play a controversial role in the antitumor response, many oncological clinical trials are focusing on agonistic antibodies with potential antitumor activity (61) (NCT03971409)(NCT02410512). Inducible CO-Stimulator of T cells (ICOS) is a costimulatory molecule induced by T-cell activation, leading to secondary stimulatory signals. Besides its expression on antitumor effector TILs, ICOS is also expressed on regulatory T cells from the TME, on which it confers an immunosuppressive activity. Since ICOS does not induce a cytotoxic immune response independently (62), clinical investigation of monoclonal antibodies for potential synergistic effects in association with other immunotherapies in advanced malignancies, including BC, are currently conducted (NCT02904226, NCT03447314,

NCT03829501). Glucocorticoid-Induced TNFR-Related gene (GITR) activation promotes effector T-cell activity and inhibits tumorinfiltrating regulatory T-cell function. GITR agonism alone does not seem to be sufficient to induce a significant clinical response to therapy. However, there is a rationale for reinvigoration TILs exhaustion through combination with PD-1 blockade (63), which is currently under clinical investigation in advanced malignancies including BC (NCT02628574)(NCT03126110). 4-1BB is a powerful costimulatory signal whose agonistic stimulation on CD8+ T-cells enhances survival, function and memory differentiation in preclinical models (64). Anti-4-1BB agonist monoclonal antibodies are under clinical investigation in combination with ICI in patients with locally advanced or metastatic TNBC (NCT02554812)(NCT03971409) and HER2-positive BC (NCT03414658). A bispecific antibody targeting HER2 and 4-1BB has also shown encouraging data of safety and clinical benefit in phase 1 clinical trial (65) (NCT03330561)(NCT03650348). In addition, 4-1BB upregulation through Fcy receptor stimulation on NK cells (66) may suggest a synergy between 4-1BB agonism and anti-HER2 blockade/anti-HER2 ADC, which is also currently investigated (NCT03364348).

CD40 is mainly expressed on antigen-presenting cells, where ligation by CD40 ligand results in antigen-presenting cell activation and therefore priming and activation of effector T cells (67). Agonist monoclonal antibodies are under investigation, as monotherapy or in combination, in patients with advanced malignancies including BC (NCT03329950)(NCT05029999).

#### Adoptive cellular therapy

Despite their antitumor reactivity, TILs, if they are not suppressed within the TME, are often exhausted, contributing to tumor immune escape mechanisms. Autologous TILs can be isolated from tumor specimens, expanded, and activated in vitro, and reinfused into the patient, alone or in combination with interleukins. In TCR-engineered lymphocytes and CAR T cell therapy, the cells are isolated from a patient's peripheral blood through leukapheresis, and genetically modified to express either a T cell receptor (TCR) or a chimeric antigen receptor (CAR) (Figure 4). Unlike its success in haematological malignancies with FDA's approval of tisagenlecleucel and axicabtagene ciloleucel, adoptive cell therapy in solid tumors is associated with major obstacles. This includes the identification of appropriate specific tumor antigen targets and the presence of a strong immunosuppressive TME. However, numerous adoptive cellular strategies have recently been developed to fight solid tumors and are under investigation in preclinical studies and clinical trials, including in BC patients. Once expressed in T cells and accompanied by costimulatory domains (such as 4-1BB or CD28), CAR T cells display antigen-specific recognition, activation, proliferation and cytotoxic function, independent of MHC presentation, which is a major advantage compared to TCR therapy (see hereunder) (68). Several targets have been proposed to date, such as mesothelin (69) (NCT02792114)(NCT02414269), c-Met (70), CEA (NCT04348643), EPCAM (NCT02915445), CD70 (NCT02830724), or MUC1 (NCT04020575)(NCT04025216). Clinical trials have also been started with an infusion of CAR T-cells targeting HER2

(NCT04511871)(NCT04430595)(NCT03740256), including intraventricular administration in patients with brain metastases (NCT03696030). CAR-T cells could also be designed to recognize a universal motif such as an Fc portion of immunoglobulins (71), allowing a potential antitumor activity in synergy with antibody treatments such as trastuzumab. Besides, other molecules demonstrated *in vitro* and *in vivo* antitumor activity against BC, such as Natural killer group 2, member D (NKG2D) (72) or Receptor tyrosine kinase-like orphan receptor 1 (ROR1) (73). CAR-T cells targeting these molecules will probably be clinically investigated soon as well.

Although CAR T-cell therapy has attracted major interest, TCR therapy can target both surface and intracellular proteins whose peptides are presented onto MHC molecules (Figure 4). As a result, TCRs could target more tumor antigens and be more tumor-specific (74). In a metastatic HER2-overexpressing BC patient, an adoptive transfer of autologous HER2-specific cytotoxic T cells has been shown to lead to accumulated T cells in the bone marrow along with a loss of bone marrow-residing disseminated tumor cells (75). The adoptive transfer of allogeneic T cells in metastatic BC patients suggested their contribution to a transient early tumor response (76). TCR T-cell therapy is currently being evaluated in patients with metastatic or locally recurrent and unresectable disease including BC through tumor-associated antigen-specific cytotoxic T lymphocyte infusion (NCT03412877)(NCT03093350).

In addition, several clinical studies investigating TIL therapy in BC have been performed. A phase 2 clinical trial demonstrated a case of adoptive transfer of autologous mutant-protein-specific TILs in association with pembrolizumab and interleukin-2 that led to the complete and durable regression of metastatic disease in an HR-positive BC patient who failed to respond to multiple previous lines of therapy (77). Clinical trials are currently investigating the transfer of autologous TILs in patients with pretreated metastatic BC (NCT01174121)(NCT04111510).

Lastly, a growing interest in CAR-NK cell immunotherapy (78) has demonstrated promising preclinical studies in TNBC. In the future, personalized immunopeptidomic profiling of the tumors may allow us to identify new potential therapeutic target antigens (79). Nextgeneration engineering CAR-T cells may also target several aspects of the TME to treat solid tumors such as the tumor stroma and vasculature as opposed to tumor cells alone, as it has recently been suggested *in vivo* (80). In addition, next-generation CAR-T cells endowed with bispecific CAR dual specificity targeting multiple tumor antigens have been shown to potentiate the antitumor activity in tumor models and to minimize parallel reactivity against normal tissues harboring single antigen, which could present a novel approach focusing CAR-T cells to tumor cells (81).

## **Bispecific antibodies**

With FDA's approval of blinatumomab in the treatment of Bcell malignancies, recent efforts have focused on the extension of multifunctional bispecific antibodies to BC immunotherapy. Most bispecific antibodies for cancer immunotherapy consist of two arms. One arm binds tumor-associated antigens on cancer cells.



The other arm binds effector immune cells, with selective binding to activate Fcy receptors, resulting in the formation of a tri-cell complex and specific elimination of tumor cells (Figure 5). This elimination is independent of TCR specificity, co-stimulatory signals, or peptide antigen presentation, representing a major advantage of bispecific antibodies in cancer immunotherapy. Compared to trastuzumab, it has been shown that these trifunctional antibodies mediate the elimination of tumor cells expressing HER2/neu at low levels, which was associated with a Th1-based cytokine release (82) and a potent antitumor activity (83). Ertumaxomab which targets CD3 and HER2 simultaneously has shown encouraging results in phase 1 clinical trials, with an antitumor response seen in 30-59% of metastatic BC patients, especially in HER2-positive BC patients (84). This provides a strong rationale for further studies involving unlimited combinations of bi- or tri-specific antibodies in the therapeutic strategies against BC. Consecutively, new phase 1 and 2 clinical trials evaluate the efficacy of bi- or tri-specific antibodies in locally advanced or metastatic BC patients, through for example HER2/ CD3 bispecific target (NCT03448042), NK cell/T-cell/HER2 trispecific target (NCT04143711), HER2/PD-1 bi-specific target (NCT04162327), or two non-overlapping domains of HER2 target (NCT04224272). A multitude of other bispecific antibodies are designed to simultaneously target molecules on the tumor cells and/or the TME, such as different immunosuppressive pathways (85) or cell-cell adhesion molecules overexpressed on BC tumor cells (86).

# Therapeutic peptide and proteinbased cancer vaccines

In the age of cancer immunotherapy we see a renewed interest in harnessing cancer-targeting vaccines for therapeutic purposes. Such vaccines can be preventive or therapeutic. Several preventive vaccinations are FDA-approved, such as HPV-related genital/head and neck cancers, and HBV-related hepatocellular carcinoma. Among the FDA-approved therapeutic cancer vaccines today are the Bacillus Calmette-Guérin (BCG) for early-stage bladder cancer and Sipuleucel-T (Provenge) for the treatment of metastatic castrateresistant prostate cancer. No vaccine has been approved for clinical use in BC to date. However, a rising interest for the development of peptide vaccines has been seen in recent years. For example, the most



studied cancer vaccine which has been successful to date is the E75 HER2 peptide vaccine, also known as Nelipepimut-S when combined with Granulocyte-Macrophage Colony-Stimulating Factor (GM-CSF). E75 is a small peptide derived from the HER2 receptor which is expected to bind HLA molecules and therefore activate cytotoxic effector T cells (Figure 6). This vaccine demonstrated in high-risk HLA-A2 BC patients a recurrence rates of 5.6% in vaccinated patients compared to 14.2% in control unvaccinated participants at 20 months follow-up (87). However, the observed difference could not be repeated in later analyses, possibly due to a lack of immune re-boost. Alternatively, one could speculate that the vaccine could be more efficient with trastuzumab or with an associated myeloid-compartment targeting strategy. Consequently, a phase 1/2 clinical trial assessed the administration of nelipepimut-S and GM-CSF in high risk BC patients in the adjuvant setting, with booster inoculations every 6 months until trial completion at 5 years. This trial demonstrated prevention of disease recurrence at 94.6% in optimally vaccinated patients versus 80.2% in the control group, with minimal toxicities (88). Unfortunately, the phase 3 study was discontinued due to futility (89). However, the addition of trastuzumab to nelipepimut-S and GM-CSF has shown a DFS of 92.6% compared with 71.9% for trastuzumab and GM-CSF group in a phase 2b clinical trial in patients with TNBC (90), with no additional overall or cardiac toxicity compared with trastuzumab alone. Together, this suggests a synergy between trastuzumab and the HER2 peptide vaccine and highlights the importance of selecting the most relevant arms when designing a clinical trial. In addition, it highlights the fact that most BC patients are defined as HER2negative tumors following fluorescence in situ hybridization (FISH) amplification methods, though they display some HER2 expression by immunohistochemistry (IHC) and therefore might respond to

anti-HER2 therapies in some combination therapies. In HER2positive BC patients, a possible synergistic immunologic effect of nelipepimut-S and trastuzumab is currently being investigated (NCT02297698). A meta-analysis of randomized clinical trials suggested significant benefits to vaccination over control, though the high heterogeneity of the patients treated in the involved trials led to unclear final results (91). Moreover, the trials generally show low toxicity of therapeutic vaccination. Vaccination thus remains an attractive strategy that seems to promote both effector immune response and protective memory immunity potentially controlling tumor relapse. Preventive BC vaccines are therefore also under investigation in patients in remission to prevent or delay relapse (NCT02780401)(NCT03384914). Additional HER2-derived peptide cancer vaccines have demonstrated beneficial impacts on BC antitumor immunity and clinical outcome, such as GP2 and AE37 (92), folate receptor  $\alpha$  peptide vaccine (93) (NCT03012100), sialyl-Tn (sTn) conjugated to keyhole limpet hemocyanin (KLH) (94), and oxidized mannan-MUC-1 vaccine (95). STn-KLH vaccine (an epitope found among others on MUC1 that activates estrogen receptor- $\alpha$  function), given concurrently with endocrine therapy, offered a robust antibody response to the vaccine and an OS advantage to metastatic BC patients in a retrospective blinded review involving 1028 women (96). This further justifies prospective randomized trials combining anti-MUC1 vaccine with endocrine therapies. Unlike peptide-based cancer vaccines, proteinbased cancer vaccines have not been explored to the same extent up till now. However, clinical studies have suggested safety, immunogenicity, long-term survival and a few high grade adverse events of protein-based cancer vaccines in patients with HER2overexpressing BC refractory to trastuzumab (97). They are under clinical investigation in BC patients undergoing neoadjuvant



endocrine therapy (NCT02204098) and in combination with ICI (NCT03632941). Thanks to the next-generation sequencing of tumor mutations and epitope-prediction strategies, new patient-specific neoantigen-based cancer vaccine will probably allow inducing highly specific antitumor immune responses (NCT02316457).

### Autologous tumor cells

Personalized vaccines targeting patient-specific mutated neoantigens, through autologous tumor cells, alone or pulsed on dendritic cells, are new potential strategies (98). Autologous tumor cell-based cancer vaccines, obtained from tumor cell isolation, may overcome the difficulties of selection of an appropriate tumor-associated antigen. These tumor cells harbor a complete and individualized repertoire of tumor-associated antigens, therefore potentially triggering a polyclonal T-cell response against various tumor cells (99) (Figure 7). Several translational and clinical trials undergone in BC patients have suggested that autologous tumor cell-based vaccines may be effective and safe, even among patients with depressed immunity (100). BC cell lines can also be used and have shown the induction of a humoral and T cell-mediated immune response alone or in combination with low-dose chemotherapy or costimulatory molecules, although the clinical results showed limited success. This is possibly because the cell lines do not express the antigen repertoire of the tumor because of inter- and intra-tumoral heterogeneity among cancer patients (101). This obstacle may be overcome by an autologous tumor cell-based vaccine, which may also minimize tumor immune escape through antigen loss observed in clinical trials (102). In this line, autologous BC tumor cells harvested from stage II-III and metastatic BC patients, irradiated and reinfused, led to encouraging results (103). Vaccination using irradiated, genetically modified GM-CSF-secreting tumor cells has shown to induce an enhanced antitumor immunity in a phase 1/2 clinical trial alone or in association with chemotherapy in metastatic BC patients (104). In an additional clinical trial, an irradiated BC cell line, endowed with antigen-presenting cell activity and in association with interferon- $\alpha$ , showed in heavily pretreated advanced BC patients an objective tumor regression in parallel with a decrease in circulating cancer-associated cells, with no serious adverse events (105). Unfortunately, clinical data about autologous tumor vaccination in BC is still limited to date, despite its promising antitumor immune potential.

# **B** cell-based vaccines

Trastuzumab's success led to enthusiasm for B cell-based vaccines, which induce an endogenous active and specific humoral B cell-immune response (antibodies with antitumor activity) from the patient's own B cells. The antibodies secreted in the patients are similar to those of the drug itself, and may offer a promising alternative to antibody administration. Indeed, trastuzumab antibody yield seems to be a major challenge for large-scale production. Trastuzumab is usually produced in Chinese Hamster Ovary Cells in incubators of 80 up to 12 000 liters, which makes a year of treatment per person very expensive. HER-2 B-cell peptide vaccine already demonstrated a robust production of anti-HER2 antibodies in patients with advanced or metastatic HER2-positive cancer of the stomach or the gastroesophageal junction. Besides, there is a correlation between the levels of anti-HER2 antibodies and the clinical response to therapy. In addition, the early data indicated a benefit in OS (hazard ratio of 0.418) with no added toxicity, in association with standardof-care chemotherapy, compared to chemotherapy alone (106). The combination of two peptide B-cell epitope vaccines, representing trastuzumab and pertuzumab binding sites, showed safety and



antitumor activity performed in patients with advanced solid tumors including BC (107). Other clinical trials are ongoing (NCT01376505). The next generation of clinical studies will have to involve the combination of vaccines with ICI, since preclinical data suggest synergistic mechanisms of action. Indeed, IFN $\gamma$  secreted by cytotoxic antitumor T cells upregulates PD-1/PD-L1 axis-mediated immune suppression. The addition of ICI to vaccine administration may block the induced immunosuppressive feedback and allow the induction of robust antitumor immunity (108).

# Dendritic cell-based vaccines and TLR agonists

Whole-cell dendritic cell (DC) vaccines are tumor-specific DC infusion, mainly isolated from patients' peripheral blood monocytic cells, exogenously matured and expanded using various cytokines,

and loaded with tumor lysate or antigens (Figure 8). This approach has shown therapeutic potential with limited toxicities and is extensively investigated in clinical trials. For example, DC loaded with tumor antigens, in association with chemotherapy, have been shown to significantly increase PFS and OS in metastatic BC patients compared to chemotherapy alone over a 10-year followup (109). Another clinical study showed a prolonged PFS from 31% to 76.9% with DC vaccine in ER/PR double-negative stage II/III BC patients (110). In addition, the DC vaccine added to neoadjuvant chemotherapy increased the pCR rate to 28.9% compared to 9.09% in the control group with neoadjuvant chemotherapy alone (111). Remarkably, these results were observed in the PD-L1-negative population, and may probably be explained by a poorly immunosuppressed TME where the vaccine can trigger an appropriate antitumor activity. This supports again the hypothesis that the combination of ICI to cancer vaccine might provide a synergistic antitumor activity. Lapuleucel-T, consisting of an adoptive transfer of autologous antigen-presenting cells activated in vitro with a recombinant fusion protein comprising HER-2



sequences, demonstrated safety and significant antitumor immune response in patients with HER-2-overexpressing metastatic BC, associated with clinical response or disease stabilization in some patients (112). Another example is the vaccination with p53 peptide-pulsed DCs, which showed disease stabilization of advanced BC patients, and a correlation between p53 expression of tumor cells and the induction of a p53-specific T cell response (113). Other clinical trials are currently investigating DC vaccine in BC patients, such as HER2-pulsed DC vaccine (NCT02063724) an multiepitope DC vaccine (NCT00266110), and are also investigated in a neoadjuvant setting (NCT02018458)(NCT02061423) (NCT03387553).

Human DC cells express Toll-Like receptors (TLRs) which play a key role in recognizing signals from the microenvironment and adapting accordingly. The DCs can be targeted and activated through TLR agonists which have shown promise in preclinical and clinical cancer studies, enhancing antitumor T-cell response. Several clinical trials are currently evaluating TLR agonists. For example, topical TLR-7 agonist imiquimod, in association with nabpaclitaxel, has shown an immune-mediated disease regression in treatment-refractory BC chest wall metastases in phase 2 clinical trial (114). Systemic TLR7 agonists are also under development to activate the antitumor immune response in patients with advanced cancer (115). Therapeutic cancer vaccination using TLR3 agonists, such as PolyICLC, are also currently under investigation in patients with advanced cancer including BC (NCT02643303). Systemic DC expansion is also being tested in patients with cancers including metastatic BC, through vaccination with polyICLC in association with Flt3L (NCT03789097).

# DNA/mRNA-based cancer vaccines

The favorable clinical experience observed with COVID-19 vaccine may help facilitate research and development in the field of DNA/mRNA-based cancer vaccines. These vaccines are mainly viral replicon particles, or completely synthetic lipid nanoparticles, and contain the genetic information coding for tumor-specific

antigens or tumor-associated antigens. Upon administration, the DNA is transcribed into mRNA, which is then translated to synthesize peptides and proteins within the ribosomes. This leads to the presentation of the peptides onto HLA, ultimately activating highly specific cytotoxic and memory T cells against tumor, possibly for a longer period of time compared to peptide or protein-based cancer vaccines. A phase 1 clinical trial investigating a naked plasmid DNA vaccine in metastatic BC patients is currently ongoing (NCT02204098) and preliminary evidence suggested an increased specific CD8+ T-cell proliferation and cytotoxicity along with an improved PFS rate (116). Another phase 1 clinical trial of 66 BC patients with advanced HER2-positive disease investigated the administration of a plasmid DNA coding for HER2 molecule, associated with GM-CSF as an adjuvant, for 3 immunizations. This study demonstrated the induction of an anti-HER2 immunity in most patients, persisting after the end of the vaccinations, and safety with 10-year postvaccine toxicity assessments (117). Importantly, the authors underscore the fact that high anti-HER2 immunity is associated with favorable clinical outcomes after trastuzumab therapy. In line with these results, a randomized phase 2 trial is currently in progress (117). Moreover, in TNBC patients with residual disease after neoadjuvant chemotherapy, another phase 1 clinical trial has indicated that DNA vaccine induced neoantigen-specific immune response in 88.8% of the patients along with a PFD of 87.5% in the vaccinated patients, compared to 49% in historical controls (118). Adverse events were mainly injection site reactions and other limited toxicities. The administration of a mRNA-based vaccine encoding a portion of HER2 (VRP HER2), with or without an additional anti-HER2 targeted therapy, also demonstrated stable diseases and partial response in a phase 1 clinical trial with advanced HER2-positive BC patients (119). In this study, PFS correlated with perforin expression by memory T cells. Other clinical trials investigating DNA/mRNA-based vaccine immunotherapies are ongoing, for example in advanced and metastatic BC patients (NCT02157051), with concurrent ICI (NCT03632941), or in non-metastatic, node-positive BC patients who are in remission (NCT02780401).

# $\delta T$ cells

In contrast to conventional  $\alpha\beta$  T cells,  $\delta$  T cells are a distinct little subset of T cells, which typically recognize and kill malignant cells rapidly and independently of HLA restriction. This highlights the antitumor impact that  $\delta \delta T$  cells may play in the next generations of cancer immunotherapies, for example through adoptive transfers of allogeneic cells from healthy donors rather than autologous ones. In addition,  $\delta T$  cells contribute also indirectly to the antitumor immune response by communicating with other tumor-infiltrating immune cells (120). Vγ9Vδ2 T-cell therapy has demonstrated safety and higher survival of late-stage lung or liver cancer patients (121). In BC, although  $\delta$  T cells have been suggested to display a degree of functional plasticity with possible opposing effects on the growth of breast tumors (122), the antitumor potential of these cells makes them an encouraging immunotherapeutic tool to exploit as well. Indeed, γδ T cells were correlated with an improved pCR rate following neoadjuvant therapy, and an improved DFS and OS (11). In addition, the antitumor activity demonstrated by the use of bisphosphonates in some BC clinical trials may be at least partly due to  $\delta T$  cell activation (123). Furthermore, BC-infiltrating  $\delta T$ cells have been suggested to be involved in the efficacy of trastuzumab, through a mechanism of ADCC (124). Xo T cells have also been associated with remission in TNBC patients (125), although conflicting observations have been reported regarding a potential suppressive function of  $\delta T$  cells on  $\alpha\beta$  T cells (126).

For example, a phase 1 clinical trial was conducted in which the bisphosphonate zoledronate, a Vy9V82 T-cell agonist, plus lowdose interleukin-2 (IL-2), were administered to terminal advanced metastatic BC patients. The treatment was well tolerated by all patients. In addition, the patients who showed a robust peripheral population of V<sub>3</sub>9Vδ2 T cells had declining CA15.3 levels and displayed partial remission and stable disease, compared to patients who failed to sustain Vγ9Vδ2 T cells (127). Infusions and other agonists of V<sub>N</sub>9Vδ2 T cells are investigated in clinical trials (128) (NCT03183206)(NCT02781805) as monotherapy or in combination with an ICI (NCT04243499). Furthermore, we expect that adoptive transfers of engineered T cells expressing antitumor  $\delta$  TCR will also be investigated in BC patients in the upcoming years (129). Although most clinical results have not shown encouraging results thus far, these attractive cells must be better understood and will then certainly show us their hidden secrets, hopefully to the advantage of BC immunotherapy.

### Tumor-associated macrophages

Macrophages are among the most abundant immune cells within the TME, where they play a key role in the antitumor immune response through pro-inflammatory macrophage activation, tumor cell phagocytosis and antigen presentation. Clinically approved cancer therapies such as trastuzumab exert at least partially their effects through these mechanisms (130), while pre-clinical studies attempt to optimize these mechanisms (131). Experimental mouse models and clinical studies demonstrated that macrophages are educated by the TME and generally adopt protumoral and immunosuppressive functions, in contrast to their tumoricidal role following *in vitro* activation (132). A metaanalysis of BC patients demonstrated that Tumor-Associated Macrophages (TAMs) were significantly correlated with an aggressive phenotype, metastasis and poor clinical prognosis (133), lack of pCR, resistance to chemotherapy (134) and tamoxifen (135). Furthermore, the protumoral effect of TAMs is often further reinforced after cancer treatments, through macrophage recruitment and polarization, limiting the efficacy of chemotherapy and radiotherapy and promoting early tumor recurrence (136). Clinical trials investigating TAMs-targeting strategies (Figure 9) suggest potential impact in the treatment of cancer.

Blocking SIRPa on the surface of TAMs and DCs, which blocks its interaction with CD47 on the surface of tumor cells and their subsequent "don't eat me" signal, restores the phagocytosis of tumor cells. Clinical trials suggested safety and therapeutic potential (137) in several cancer patients. Furthermore, the concomitant blockade of the CD47/SIRPa axis with tumortargeting monoclonal antibodies such as trastuzumab may provide a synergistic phagocytic antitumor activity. In this way, the preliminary antitumor activity of this association has been demonstrated in rituximab-refractory non-Hodgkin lymphoma patients and in solid tumors (138, 139). In BC, no clinical trial has assessed such a combination up to now. However, CD47 gene expression has been found to limit the therapeutic activity of trastuzumab in HER2-positive BC patients (15). Other "don't eat me" anti-phagocytic signals include PD-L1, or recently highlighted CD24, which has been recently suggested as a new therapeutic target for BC immunotherapy (140). Other potential myeloid immunotherapeutic strategies may be the blockade of myeloid cell recruitment to the TME, differentiation into TAMs or proliferation within the TME. For example, CCR2-CCL2 axis contributes to metastatic BC progression and early relapse through mechanisms such as myeloid cell recruitment or TAM polarization (141). Another example is the colony-stimulating factor 1 (CSF1)/ colony-stimulating factor 1 receptor (CSF-1R) axis, which is a key regulator of myeloid cell differentiation and chemotaxis, and which has been associated with BC progression and mortality (142). However, several treatments targeting these signaling pathways have been investigated in early phase clinical trials and have had disappointing results in the clinic to date, in part due to compensatory feedback mechanisms with no long-term benefit in solid tumors or metastatic cancers. Depletion of TAM could also be a valuable approach to facilitate the antitumor immune response in BC. For example, trabectidin (Yondelis) is effective at killing TAMs in addition to cancer cells (143). Therefore, its antitumor activity should be investigated in some BC subtypes with a high proportion of TAM, such as HR-positive BC (11), where TAM have been associated with worse survival (144). Remarkably, trabectidin has already shown a manageable safety profile and up to 56% of stable disease in several clinical trials with advanced or metastatic BC patients (145). In heavily pretreated metastatic HER2-positive or TNBC patients, partial response occurred in 12% of HER2-positive BC patients (146). In addition, zoledronic acid, which is commonly



used in bone metastatic patients, showed extra-squeletal beneficial effects such as the prevention of distant relapse in BC patients after menopause (147), likely through TAMs depletion (148). One of the limitations of such myeloid immunotherapeutic strategies is the targeting of macrophages as a whole population, without taking into consideration the functional properties of subpopulations of macrophages. Indeed, some macrophage subtypes have for example been correlated with less advanced stages, less aggressive tumors (149) and better survival (150) in BC patients, while other subtypes have been suggested to be an independent prognostic marker for longer DFS and OS (151). In this context, another strategy aims at targeting a specific so-called "M2-like" subpopulation of macrophages or switching TAM polarization toward an antitumor phenotype, rather than depleting macrophages indiscriminately (152). Modulators of macrophage phenotype are tested in clinical trials in patients with solid tumors including BC patients, in combination with ICI (NCT02637531).

# Radiotherapeutics

Radiotherapeutics is based on a theranostic strategy that combines a whole-body non-invasive mapping of the cancer disease (through a targeted radioactive drug) and the delivery of a targeted therapy to the cancer cells (through a second radioactive drug). Although evidence is still limited, radiotherapeutics is emerging as a superior approach when compared to usual 18F-FDG PET-CT approach in detecting primary and metastatic BC disease. In addition, radiotherapeutics may help select patients who will benefit from therapy, and can become a new efficient targeted therapeutic option in metastatic setting. For example, the radiolabelling of trastuzumab with zirconium-89 or lutetium-177 has been suggested as specific radioimmunotherapy for HER2positive BC patients (153, 154). Clinical trials assessing Her2 expression detection and anti-HER2 radionuclide therapy are currently ongoing in BC patients (NCT04674722) (NCT04467515). Prostate-specific membrane antigen (PSMA) is specifically expressed in tumor-associated vasculature of solid tumors such as BC, suggesting that it may be targeted as a new anti-angiogenic therapy. Progressive TNBC patients are currently being recruited to assess the concordance between lesions observed on Ga-PSMA PET-CT and 18F-FDG PET-CT and evaluate the feasibility of lutetium-177 PSMA therapy (NCT06059469). Another example is the Fibroblast Activation Protein (FAP)-targeted radionuclide therapy such as lutetium-177-FAPi which targets FAP-expressing Cancer-Associated Fibroblasts (CAFs), stroma cells from the TME endowed mainly with protumoral and immunosuppressive properties. Several studies suggested feasibility, safety, detection of metastases in specific areas (such as in the brain), and reduction of pain in metastatic BC (155, 156). Many other therapeutic isotopes and immunosuppressive targets within the TME might provide new radiotherapeutic and palliative tools for metastatic BC patients.

# **Oncolytic viruses**

Oncolytic viruses have been produced within the last decade. They are engineered to (or they preferentially) target tumor cells. Then, viruses replicate specifically in tumor cells, and stimulate

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antitumor immunity, ultimately killing tumor cells. This is mediated by the release of damage-associated molecular patterns and pathogen-associated molecular patterns, along with tumor-specific or tumor-associated antigens presentation. Furthermore, the oncolytic viruses induce immunogenic tumor cell death. Talimogene laherparepvec, a genetically modified herpes simplex oncolytic virus, has been approved by the FDA for the local treatment of metastatic melanoma with unresectable cutaneous lesions. In BC, many preclinical studies suggest antitumor effects of oncolytic virotherapy and its synergistic effects with chemotherapy.

The oncolvtic herpes virus HF10 has been injected in BC patients with recurrent BC and suggested higher tumor-infiltrating CD8+ T cells, although the number of patients involved in the study was limited (157). Given the limited efficacy observed in single virotherapy to date, combination drug approaches including virotherapy are being further evaluated in BC patients. For example, a phase 2 randomized study of paclitaxel alone or in combination with oncolytic reovirus demonstrated, in 74 previously treated metastatic BC patients, a significantly longer OS when the combination treatment was administered (158). In another study, SD were observed in metastatic TNBC patients treated by virotherapy associated with lowdose cyclophosphamide (159). Oncolytic virotherapy is currently being investigated in advanced and metastatic BC patients in association with chemotherapy (NCT01656538)(NCT02630368), chemotherapy and ICI (NCT02630368)(NCT02977156) (NCT04215146). The sequence of the therapies over time might also be important in the efficacy of combinatorial therapeutic strategies. For example, preliminary oncolytic virotherapy has been suggested to sensitize BC to following chemo- or immunotherapy, since it induces a preexisting nonexhausted antitumor immunity (160). In this way, clinical trials are now recruiting BC patients for oncolytic virotherapy in association with neoadjuvant chemotherapy (NCT02779855)(NCT03564782), or with radiation therapy followed by pembrolizumab (NCT03004183). In addition, new immunotherapeutic strategies, such as oncolytic virotherapy coding for localized trastuzumab monoclonal antibody production (161), may provide synergistic antitumor effects. Nonreplicating virus strategies have also been investigated. One strategy consists of the delivery of a gene that converts a drug into a cytotoxic drug. For example, a phase 1 trial used a local injection of a retrovirus encoding the human cytochrome P450 gene in BC metastatic cutaneous nodules in association with oral administration of the prodrug cyclophosphamide. The trial suggested safety and antitumor efficacy (162).

## Cytokine-based immunotherapy

Cytokines are major and pleiotropic regulators of the immune response. In the era of cancer immunotherapy, a renewed interest in the properties of cytokines has led to an increased number of clinical trials assessing their safety and efficacy. In BC, since cytokine-based immunotherapy has shown limited efficacy up to now, cytokines in association with other immunotherapies are currently being investigated to increase their efficacy. For example, it has been hypothesized that interferon- $\alpha$  (IFN- $\alpha$ ), which upregulates tumor antigen presentation on tumor cells, might synergize with a cancer vaccine. Therefore, IFN- $\alpha$  has been administered in association with the CEA vaccine in thirty-three CEA-expressing cancer patients including BC. The administration of IFN-a induced a significantly increased OS compared to the vaccine alone (163). Interferon- $\gamma$  (IFN- $\gamma$ ) is another cytokine which plays a crucial role in tumor cell cytotoxicity, and has also been suggested to be used as an adjuvant for immunotherapy. Hence, clinical trials are for example currently studying the association of IFN-y with paclitaxel, trastuzumab and pertuzumab in HER-2positive BC patients (NCT03112590), or with ICI in TNBC (NCT02614456). TGF- $\beta$  is a cytokine which contributes to the immune suppression of the TME and has been associated with resistance to cancer immunotherapy. Its blockade during radiotherapy in metastatic BC patients has shown a higher median OS (164). Other major pro-inflammatory cytokines, such as IL-12 or IL-15, stimulate among others the production of IFN- $\gamma$ from CD8+ T cells and induce the differentiation of CD4+ T cells into Th1. IL-12 or IFNy, in combination with ICI and chemotherapy, may synergize efficiently. Indeed, these cytokines play key roles in the crosstalk between myeloid and lymphoid cells and in cellular cytotoxicity. This is currently under clinical investigation (NCT03567720)(NCT03112590). However, the severe IL-12-mediated toxicity restricted its use in clinical trials. IL-15 is currently investigated in association with ICI in patients with refractory cancers such as BC (NCT03388632). Finally, IL-2 plays a crucial role by stimulating, among others, the proliferation and the cytotoxicity of CD8+ T cells, but also the proliferation of regulatory T cells, which are major suppressors of the antitumor immune response. To overcome these limitations, engineered IL-2 and new inhibitors of regulatory T cell activity are under investigation. ICI have been suggested to inhibit regulatory T cell activity, and are therefore investigated in association with IL-2 in advanced cancers including BC (NCT05086692). The properties of cytokines make them a promising additional tool in cancer immunotherapy and will probably help facilitate the antitumor immune response, once they will be better understood and exploited.

### Immunometabolic targets

Accumulating evidence indicate a metabolic competition for consumption of glucose, amino acids and fatty acids between the tumor cells and the tumor-infiltrating immune cells. These molecules are essential for immune cell survival and activity, modulating in this way the antitumor immune response. This led to the concept of metabolic reprogramming. For example, enzymes such as arginase-1 (Arg-1) or indoleamine 2,3-dioxygenase (IDO-1), which catabolize arginine and tryptophan respectively, seem to be mainly involved in immunosuppressive pathways. Therefore, they have been targeted in BC preclinical models, demonstrating synergistic antitumor effects in association with chemotherapy (165, 166) and ICI (167). There are still a few immunometabolic targets investigated in BC clinical trials to date. A randomized clinical trial with HER2-negative metastatic BC patients failed to show improved PFS when IDO-1 inhibitor was added to chemotherapy (168). In

contrast, IDO-1 inhibitor in association with p53-DC vaccine has been shown to increase the IFN x-producing CD8 and the IL-2producing CD4 T cell response in phase 1/2 study of metastatic BC patients, although it did not increase the objective response rate (169). Arginase inhibition, alone or in combination with ICI, is currently being evaluated in phase 1 study in advanced or metastatic cancer patients (NCT02903914). Accumulating evidence suggests that metabolic changes in adipose tissue are also associated with immunological dysregulations in BC (170). Fasting, or antihyperglycemic agents such as metformin, may modulate the antitumor immune response, improve the response to chemoand immunotherapies, and reduce side effects (171). While fasting has been suggested to reduce the risk of BC recurrence (172), fasting (NCT05023967) and other metabolic health patterns (NCT05432856) are currently being clinically investigated. In addition, in most preclinical studies, hypoxia within the TME, and its associated VEGF induction, also contributes to tumor immune escape mechanisms and tumor progression. Moreover, hypoxia generally increases along with tumor progression, further promoting its deleterious effects (173). Furthermore, hypoxia increases extracellular levels of adenosine in the TME. Adenosine binds to its receptors on immune cells and further contributes to the establishment of an immunosuppressive TME (174). Rapidly proliferating malignant cells generate also high amount of lactate, a by-product of tumoral aerobic glycolysis. Lactate contributes to acidosis, stimulates angiogenesis, acts as cancer cell metabolic fuel, exerts deleterious effects on tumor-infiltrating immune cells and has been suggested to predict response to immunotherapy (175). Unfortunately, hypoxia-inducible factor  $1-\alpha$  (HIF- $1\alpha$ ) and lactate metabolism inhibitors have mainly been investigated at a basic research level up to now or have only occasionally been clinically assessed. Adenosine has emerged as a key negative regulator of antitumor immunity through the CD39-CD73-A2AR pathway (176). Several inhibitors of this pathway are currently being evaluated in early phase clinical trials alone or in combination with ICI or chemotherapy in patients with advanced solid tumors including BC patients (NCT02740985)(NCT02503774) (NCT02754141), with trastuzumab (NCT05143970) or in combination with radiotherapy (NCT03875573). Nevertheless, metabolic reprogramming is emerging and require further preclinical and clinical investigations before leading to safe and efficient immunotherapeutic adjuvants for BC immunotherapies.

## Discussion

It is well established that the TME plays a crucial role in cancer outcomes and response to therapy. Indeed, it provides a supportive but also active protumoral and immunosuppressive framework for tumor progression and dissemination. On the other hand, evidence demonstrating how to harness the immune system in favor of strong antitumor immunity is overflowing. To improve research and development in BC immunotherapy, clinical trials must be critically designed, considering both molecular and cellular mechanisms at play and its clinical pitfalls.

The next generation of cancer immunotherapy will probably involve combination immunotherapies. Indeed, the current approach of cancer immunotherapy mainly focuses on the T-cell compartment, allowing one to speculate that additional complementary immunotherapeutic strategies targeting different immune compartments could lead to synergistic therapeutic approaches. More specifically, numerous clinical trials assess immunotherapeutic combinations that mechanistically regulate redundant pathways (such as the combination of two different ICI for example). Instead, targeting different complementary pathways could trigger a stronger antitumor immune response. Specifically, the combination of myeloid and lymphoid "immune checkpoints" should be investigated further. Lymphoid immune checkpoints on T cells are widely studied. In contrast, myeloid cells (such as macrophages and dendritic cells) and their potential associated therapeutic targets are clinically less well-known. However, the myeloid cells are still key modulators of the adaptive immune system. Indeed, they can cooperate with TILs to develop a strong and long-lasting antitumor immune response together. Unfortunately, the tumor modulates its TME. Instead of promoting an inflammatory response, many preclinical studies demonstrated that most tumor-associated myeloid cells strongly suppress TILs. Therefore, there has been an explosive growth of clinical trials targeting tumor-infiltrating myeloid cells worldwide, most of them still at an early phase (177, 178). However, targeting only the lymphoid or the myeloid compartment of the TME may not adequately restore the antitumor immune response. We believe that restoring a positive crosstalk between tumorinfiltrating myeloid and lymphoid immune cells may optimize BC immunotherapy.

Distinct macrophages and dendritic cells have been suggested to predict the response to ICI immunotherapy in human BC (179, 180) and other solid tumors (181, 182). An antitumor vaccine has demonstrated long-term tumor control in a HER2-positive BC model but only when combined with anti-PD-1 treatment. This has led to the investigation of this therapeutic combination in a phase 2 clinical trial (NCT03632941). Furthermore, a first-inhuman phase 1 trial supports further clinical investigation of evorpacept, a protein that promotes tumor phagocytosis by dendritic cells and macrophages, combined with pembrolizumab, in patients with solid tumors (139). Lastly, eganelisib, a potential first-in-class tumor macrophage-targeting agent (NCT02637531), is already showing PFS benefit in metastatic TNBC patients in addition to atezolizumab and nab-paclitaxel in an ongoing phase 2 MARIO-3 trial (NCT03961698).

Moreover, compensatory mechanisms are very often at play in the TME, further suggesting the need for combination immunotherapies. For example, several studies suggested an increased immunosuppressive microenvironment after CAR T cell therapy (102). CAR T-cells infusion without targeting the TME in parallel, which suppresses TILs, might seem senseless. New complementary strategies aiming at improving T-cell trafficking into/proliferation within the TME, such as myeloid cell depletion, may improve CAR T-cell efficacy. In addition, tumor PD-L1 upregulation occurs in response to IFN $\gamma$  release by effector immune cells, leading to subsequent immune suppression, a process known as

adaptive immune resistance, where tumor cells protect themselves from immune attack (108). Therefore, the addition of ICI to other types of immunotherapies, such as cancer vaccines for example, may restrain induced immunosuppressive feedback. Moreover, combinations of conventional therapies and immunotherapies should be investigated. Indeed, although radiotherapy and chemotherapy can result in immunogenic cell death (183), they can also limit their own therapeutic effects. For example, conventional cytotoxic drugs and vascular-targeting agents induce tumor cells to produce macrophage recruitment factors (136), while macrophages can also be recruited and polarized during radiotherapy treatment (184). This promotes tumor tissue repair and early tumor recurrence, which might be thwarted by myeloid-targeting immunotherapeutic strategies. Other myeloid and lymphoid-based treatment combinations involving, in addition to an ICI, a CD40 agonist (NCT03424005) or an antitumor vaccine (NCT03632941), may synergize by activating dendritic cells. An additional rational combination might be the concomitant blockade of the CD47/ SIRP $\alpha$  axis (or other anti-phagocytic signal blockade) with trastuzumab, which might provide a synergistic phagocytic antitumor activity. In accordance with this hypothesis, CD47 gene expression has been found to limit the therapeutic activity of trastuzumab in HER2-positive BC patients (15). In addition, some treatments such as corticosteroids, mainly used for symptomatic purposes in oncology, should be administered carefully, since their impact on the antitumor immune response is still not well understood (185). The sequence of immunotherapies over time might also impact their efficacy. Indeed, the TME involves a complex dynamic network of immune cells interacting with each other, displaying changes in their activation state over time, and in this way affecting tumor progression and response to therapy. This concept is underestimated in trials evaluating heavily pretreated metastatic BC patients and excluding earlier settings. Furthermore, the dose and schedule of therapies play an important role in their activities. For example, Gonadotropin-Releasing Hormone (GnRH) agonists achieve castration because of continuous pituitary stimulation in contrast to the physiologic pulsatile fashion. Another example is cyclophosphamide, which has cytotoxic and immunosuppressive effects at high dosage, but displays immunostimulatory and antiangiogenic effects at a daily lower dose. New trial designs involving metronomic chemotherapies in association with ICI should be done in search of the optimal antitumor activity (NCT03971045). In a near future, new technologies such as machine learning will probably be a tool of an inestimable value to help us better understand immune cell subpopulation activities and interactions under therapies, and suggest efficient combinations of therapies.

Predictive biomarkers of response to immunotherapy are sorely lacking today and must be developed. Indeed, PD-1 and PD-L1 expression as biomarkers are not reliable enough, probably because the response to therapy is much more complex than that of those sole molecules. While TILs are considered in clinical practice as a promise of good prognosis (186, 187) and response to therapy, they consist of many subpopulations of lymphocytes exerting various and sometimes opposite immune functions (188). For example, cytotoxic CD8+ T cells and CD4+ Th1 T cells are associated with BC survival. On the other hand, potent immunosuppressive TILs such as regulatory T cells are associated with poor BC clinical outcome. As seen previously, different tumor-infiltrating myeloid cells can also exert critical opposite functions around the tumor cells and in metastatic niches. Based on the characterization of various tumor-infiltrating immune cell subpopulations and their correlations with clinical outcomes, a cytotoxic/regulatory immunogenic ratio might be conceived and used as a new predictive biomarker of response to therapy in future biopsies.

To conclude, immunotherapy using ICI has shown unprecedented and long-term efficacy in the treatment of cancer patients with various advanced solid tumors in the last decade, and is currently used in many oncology treatments and clinical trials (189). It has become one of the pillars of cancer treatment, but its success is currently still the visible tip of the iceberg. Indeed, countless efforts have been made to develop cancer immunotherapies and multiple new promising therapies are likely to emerge. Meanwhile, many challenges are still to be overcome, and the next decade will see the fruitfulness of BC immunotherapy investigations, including appropriate synergistic combinations with standards of care.

#### Author contributions

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# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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# References

1. Loi S, Sirtaine N, Piette F, Salgado R, Viale G, Van Eenoo F, et al. Prognostic and predictive value of tumor-infiltrating lymphocytes in a phase III randomized adjuvant breast cancer trial in node-positive breast cancer comparing the addition of docetaxel to doxorubicin with doxorubicin-based chemotherapy: BIG 02-98. J Clin Oncol Off J Am Soc Clin Oncol (2013) 31(7):860–7. doi: 10.1200/JCC.2011.41.0902

2. Adams S, Gray RJ, Demaria S, Goldstein L, Perez EA, Shulman LN, et al. Prognostic value of tumor-infiltrating lymphocytes in triple-negative breast cancers from two phase III randomized adjuvant breast cancer trials: ECOG 2197 and ECOG 1199. J Clin Oncol Off J Am Soc Clin Oncol (2014) 32(27):2959–66. doi: 10.1200/JCO.2013.55.0491

3. Loi S, Michiels S, Salgado R, Sirtaine N, Jose V, Fumagalli D, et al. Tumor infiltrating lymphocytes are prognostic in triple negative breast cancer and predictive for trastuzumab benefit in early breast cancer: results from the FinHER trial. *Ann Oncol Off J Eur Soc Med Oncol* (2014) 25(8):1544–50. doi: 10.1093/annonc/mdu112

4. Perez EA, Ballman KV, Tenner KS, Thompson EA, Badve SS, Bailey H, et al. Association of stromal tumor-infiltrating lymphocytes with recurrence-free survival in the N9831 adjuvant trial in patients with early-stage HER2-positive breast cancer. *JAMA Oncol* (2016) 2(1):56–64. doi: 10.1001/jamaoncol.2015.3239

5. Loi S, Michiels S, Salgado R, Sirtaine N, Jose V, Fumagalli D, et al. Abstract S1-05: Tumor infiltrating lymphocytes (TILs) indicate trastuzumab benefit in early-stage HER2-positive breast cancer (HER2+ BC). *Cancer Res* (2013) 73(24 Supplement):S1– S1-05. doi: 10.1158/0008-5472.SABCS13-S1-05

6. Denkert C, von Minckwitz G, Brase JC, Sinn BV, Gade S, Kronenwett R, et al. Tumor-infiltrating lymphocytes and response to neoadjuvant chemotherapy with or without carboplatin in human epidermal growth factor receptor 2-positive and triplenegative primary breast cancers. J Clin Oncol Off J Am Soc Clin Oncol (2015) 33(9):983– 91. doi: 10.1200/JCO.2014.58.1967

7. Ali HR, Provenzano E, Dawson SJ, Blows FM, Liu B, Shah M, et al. Association between CD8+ T-cell infiltration and breast cancer survival in 12,439 patients. *Ann Oncol Off J Eur Soc Med Oncol* (2014) 25(8):1536–43. doi: 10.1093/annonc/mdu191

8. Skriver SK, Jensen M-B, Knoop AS, Ejlertsen B, Laenkholm A-V. Tumourinfiltrating lymphocytes and response to neoadjuvant letrozole in patients with early oestrogen receptor-positive breast cancer: analysis from a nationwide phase II DBCG trial. *Breast Cancer Res BCR* (2020) 22(1):46. doi: 10.1186/s13058-020-01285-8

9. Dunbier AK, Ghazoui Z, Anderson H, Salter J, Nerurkar A, Osin P, et al. Molecular profiling of aromatase inhibitor-treated postmenopausal breast tumors identifies immune-related correlates of resistance. *Clin Cancer Res Off J Am Assoc Cancer Res* (2013) 19(10):2775–86. doi: 10.1158/1078-0432.CCR-12-1000

10. Criscitiello C, Vingiani A, Maisonneuve P, Viale G, Viale G, Curigliano G. Tumor-infiltrating lymphocytes (TILs) in ER+/HER2- breast cancer. *Breast Cancer Res Treat* (2020) 183(2):347-54. doi: 10.1007/s10549-020-05771-7

11. Bense RD, Sotiriou C, Piccart-Gebhart MJ, Haanen JBAG, van Vugt MATM, de Vries EGE, et al. Relevance of tumor-infiltrating immune cell composition and functionality for disease outcome in breast cancer. *J Natl Cancer Inst* (2017) 109(1). doi: 10.1093/jnci/djw192

12. Nersesian S, Schwartz SL, Grantham SR, MacLean LK, Lee SN, Pugh-Toole M, et al. NK cell infiltration is associated with improved overall survival in solid cancers: A systematic review and meta-analysis. *Transl Oncol* (2020) 14(1):100930. doi: 10.1016/j.tranon.2020.100930

13. Awad RM, De Vlaeminck Y, Maebe J, Goyvaerts C, Breckpot K. Turn back the TIMe: targeting tumor infiltrating myeloid cells to revert cancer progression. *Front Immunol* (2018) 9:1977. doi: 10.3389/fimmu.2018.01977

14. Collins DM, O'Donovan N, McGowan PM, O'Sullivan F, Duffy MJ, Crown J. Trastuzumab induces antibody-dependent cell-mediated cytotoxicity (ADCC) in HER-2-non-amplified breast cancer cell lines. *Ann Oncol Off J Eur Soc Med Oncol* (2012) 23 (7):1788–95. doi: 10.1093/annonc/mdr484

15. Tsao L-C, Crosby EJ, Trotter TN, Agarwal P, Hwang BJ, Acharya C, et al. CD47 blockade augmentation of trastuzumab antitumor efficacy dependent on antibody-dependent cellular phagocytosis. *JCI Insight* (2019) 4(24). doi: 10.1172/jci.insight.131882

16. Kim PS, Armstrong TD, Song H, Wolpoe ME, Weiss V, Manning EA, et al. Antibody association with HER-2/neu-targeted vaccine enhances CD8 T cell responses in mice through Fc-mediated activation of DCs. *J Clin Invest* (2008) 118(5):1700–11. doi: 10.1172/JCI34333

17. Chen G, Gupta R, Petrik S, Laiko M, Leatherman JM, Asquith JM, et al. A feasibility study of cyclophosphamide, trastuzumab, and an allogeneic GM-CSF-secreting breast tumor vaccine for HER2+ metastatic breast cancer. *Cancer Immunol Res* (2014) 2:949–61. doi: 10.1158/2326-6066.CIR-14-0058

18. Muraro E, Comaro E, Talamini R, Turchet E, Miolo G, Scalone S, et al. Improved Natural Killer cell activity and retained anti-tumor CD8(+) T cell responses contribute to the induction of a pathological complete response in HER2-positive breast cancer patients undergoing neoadjuvant chemotherapy. *J Transl Med* (2015) 13:204. doi: 10.1186/s12967-015-0567-0

19. Nami B, Maadi H, Wang Z. Mechanisms underlying the action and synergism of trastuzumab and pertuzumab in targeting HER2-positive breast cancer. *Cancers* (2018) 10:342. doi: 10.3390/cancers10100342

20. Rugo HS, Im SA, Cardoso F, Cortes J, Curigliano G, Musolino A, et al. Margetuximab versus trastuzumab in patients with previously treated HER2-positive advanced breast cancer (SOPHIA): final overall survival results from a randomized phase 3 trial. *J Clin Oncol JCO* (2022) 21:2937. doi: 10.1200/JCO.21.02937

21. Drago JZ, Modi S, Chandarlapaty S. Unlocking the potential of antibody-drug conjugates for cancer therapy. *Nat Rev Clin Oncol* (2021) 18:327–44. doi: 10.1038/ s41571-021-00470-8

 Li F, Ulrich M, Jonas M, Stone IJ, Linares G, Zhang X, et al. Tumor-associated macrophages can contribute to antitumor activity through fcγR-mediated processing of antibody–drug conjugates. *Mol Cancer Ther* (2017) 16:1347–54. doi: 10.1158/1535-7163.MCT-17-0019

23. Cortés J, Kim SB, Chung WP, Im SA, Park YH, Hegg R, et al. Trastuzumab deruxtecan versus trastuzumab emtansine for breast cancer. *N Engl J Med* (2022) 386:1143–54. doi: 10.1056/NEJMoa2115022

24. Perrone E, Lopez S, Zeybek B, Bellone S, Bonazzoli E, Pelligra S, et al. Preclinical activity of sacituzumab govitecan, an antibody-drug conjugate targeting trophoblast cell-surface antigen 2 (Trop-2) linked to the active metabolite of irinotecan (SN-38), in ovarian cancer. *Front Oncol* (2020) 10:118. doi: 10.3389/fonc.2020.00118

25. Bardia A, Hurvitz SA, Tolaney SM, Loirat D, Punie K, Oliveira M, et al. Sacituzumab govitecan in metastatic triple-negative breast cancer. *N Engl J Med* (2021) 384:1529–41. doi: 10.1056/NEJMoa2028485

26. Rugo HS, Bardia A, Marmé F, Cortes J, Schmid P, Loirat D, et al. Sacituzumab govitecan in hormone receptor-positive/human epidermal growth factor receptor 2-negative metastatic breast cancer. *J Clin Oncol Off J Am Soc Clin Oncol* (2022) 40:3365–76. doi: 10.1200/JCO.22.01002

27. Seidel JA, Otsuka A, Kabashima K. Anti-PD-1 and anti-CTLA-4 therapies in cancer: mechanisms of action, efficacy, and limitations. *Front Oncol* (2018) 8:86. doi: 10.3389/fonc.2018.00086

28. Winer EP, Lipatov O, Im SA, Goncalves A, Muñoz-Couselo E, Lee KS, et al. Pembrolizumab versus investigator-choice chemotherapy for metastatic triple-negative breast cancer (KEYNOTE-119): a randomised, open-label, phase 3 trial. *Lancet Oncol* (2021) 22:499–511. doi: 10.1016/S1470-2045(20)30754-3

29. Cortes J, Cescon DW, Rugo HS, Nowecki Z, Im SA, Yusof MM, et al. KEYNOTE-355: Randomized, double-blind, phase III study of pembrolizumab + chemotherapy versus placebo + chemotherapy for previously untreated locally recurrent inoperable or metastatic triple-negative breast cancer. J Clin Oncol (2020) 38:1000–0. doi: 10.1200/JCO.2020.38.15\_suppl.1000

30. Cortes J, Rugo HS, Cescon DW, Im SA, Yusof MM, Gallardo C, et al. Pembrolizumab plus chemotherapy in advanced triple-negative breast cancer. *N Engl J Med* (2022) 387:217–26. doi: 10.1056/NEJMoa2202809

31. Schmid P, Cortes J, Pusztai L, McArthur H, Kümmel S, Bergh J, et al. Pembrolizumab for early triple-negative breast cancer. *N Engl J Med* (2020) 382:810–21. doi: 10.1056/NEJMoa1910549

32. Tolaney SM, Kalinsky K, Kaklamani VG, D'Adamo DR, Aktan G, Tsai ML, et al. Eribulin plus pembrolizumab in patients with metastatic triple-negative breast cancer (ENHANCE 1): A phase ib/II study. *Clin Cancer Res* (2021) 27(11):3061–8. doi: 10.1158/1078-0432.CCR-20-4726

33. Loi S, Giobbie-Hurder A, Gombos A, Bachelot T, Hui R, Curigliano G, et al. Pembrolizumab plus trastuzumab in trastuzumab-resistant, advanced, HER2-positive breast cancer (PANACEA): a single-arm, multicentre, phase 1b-2 trial. *Lancet Oncol* (2019) 20:371–82. doi: 10.1016/S1470-2045(18)30812-X

34. Rugo HS, Delord JP, Im SA, Ott PA, Piha-Paul SA, Bedard PL, et al. Safety and antitumor activity of pembrolizumab in patients with estrogen receptor-positive/ human epidermal growth factor receptor 2-negative advanced breast cancer. *Clin Cancer Res Off J Am Assoc Cancer Res* (2018) 24:2804–11. doi: 10.1158/1078-0432.CCR-17-3452

35. Nanda R, Liu MC, Yau C, Shatsky R, Pusztai L, Wallace A, et al. Effect of pembrolizumab plus neoadjuvant chemotherapy on pathologic complete response in women with early-stage breast cancer: an analysis of the ongoing phase 2 adaptively randomized I-SPY2 trial. *JAMA Oncol* (2020) 6:676–84. doi: 10.1001/jamaoncol.2019.6650

36. Goel S, DeCristo MJ, Watt AC, BrinJones H, Sceneay J, Li BB, et al. CDK4/6 inhibition triggers anti-tumour immunity. Nature (2017) 548:471–5. doi: 10.1038/ nature23465

37. Schmid P, Adams S, Rugo HS, Schneeweiss A, Barrios CH, Iwata H, et al. Atezolizumab and nab-paclitaxel in advanced triple-negative breast cancer. *N Engl J Med* (2018) 379:2108–21. doi: 10.1056/NEJMoa1809615

38. Schmid P, Adams S, Rugo HS, Schneeweiss A, Barrios CH, Iwata H, et al. IMpassion130: updated overall survival (OS) from a global, randomized, double-blind, placebo-controlled, Phase III study of atezolizumab (atezo) + nab-paclitaxel (nP) in previously untreated locally advanced or metastatic triple-negative breast cancer (mTNBC). J Clin Oncol (2019) 37:1003–3. doi: 10.1200/JCO.2019.37.15\_suppl.1003

39. Miles D, Gligorov J, André F, Cameron D, Schneeweiss A, Barrios C, et al. Primary results from IMpassion131, a double-blind, placebo-controlled, randomised phase III trial of first-line paclitaxel with or without atezolizumab for unresectable locally advanced/metastatic triple-negative breast cancer. Ann Oncol Off J Eur Soc Med Oncol (2021) 32:994–1004. doi: 10.1016/j.annonc.2021.05.801

40. Gianni L, Huang CS, Egle D, Bermejo B, Zamagni C, Thill M, et al. Pathologic complete response (pCR) to neoadjuvant treatment with or without atezolizumab in triple-negative, early high-risk and locally advanced breast cancer: NeoTRIP Michelangelo randomized study. *Ann Oncol Off J Eur Soc Med Oncol* (2022) 33:534–43. doi: 10.1016/j.annonc.2022.02.004

41. Mittendorf EA, Zhang H, Barrios CH, Saji S, Jung KH, Hegg R, et al. Neoadjuvant atezolizumab in combination with sequential nab-paclitaxel and anthracycline-based chemotherapy versus placebo and chemotherapy in patients with early-stage triple-negative breast cancer (IMpassion031): a randomised, double-blind, phase 3 trial. *Lancet* (2020) 396:1090–100. doi: 10.1016/S0140-6736 (20)31953-X

42. Loi S, Francis PA, Zdenkowski N, Gebski V, Fox SB, White M, et al. Neoadjuvant ipilimumab and nivolumab in combination with paclitaxel following anthracyclinebased chemotherapy in patients with treatment resistant early-stage triple-negative breast cancer (TNBC): A single-arm phase 2 trial. *J Clin Oncol* (2022) 40:602–2. doi: 10.1200/JCO.2022.40.16\_suppl.602

43. Pusztai L, et al. Durvalumab (MEDI4736) concurrent with nab-paclitaxel and dose dense doxorubicin cyclophosphamide (ddAC) as neoadjuvant therapy for triple negative breast cancer (TNBC). *J Clin Oncol* (2018) 36:586–6. doi: 10.1200/JCO.2018.36.15\_suppl.586

44. Foldi J, Silber A, Reisenbichler E, Singh K, Fischbach N, Persico J, et al. Neoadjuvant durvalumab plus weekly nab-paclitaxel and dose-dense doxorubicin/ cyclophosphamide in triple-negative breast cancer. *NPJ Breast Cancer* (2021) 7:9. doi: 10.1038/s41523-021-00219-7

45. Emens LA, Esteva FJ, Beresford M, Saura C, De Laurentiis M, Kim SB, et al. Trastuzumab emtansine plus atezolizumab versus trastuzumab emtansine plus placebo in previously treated, HER2-positive advanced breast cancer (KATE2): a phase 2, multicentre, randomised, double-blind trial. *Lancet Oncol* (2020) 21:1283–95. doi: 10.1016/S1470-2045(20)30465-4

46. Huober J, Barrios CH, Niikura N, Jarząb M, Chang YC, Huggins-Puhalla SL, et al. Atezolizumab with neoadjuvant anti-human epidermal growth factor receptor 2 therapy and chemotherapy in human epidermal growth factor receptor 2-positive early breast cancer: primary results of the randomized phase III IMpassion050 trial. J Clin Oncol Off J Am Soc Clin Oncol (2022) 40:2946–56. doi: 10.1200/JCO.21.02772

47. Rudd CE, Taylor A, Schneider H. CD28 and CTLA-4 coreceptor expression and signal transduction. *Immunol Rev* (2009) 229:12–26. doi: 10.1111/j.1600-065X.2009.00770.x

48. Arce Vargas F, Furness AJS, Litchfield K, Joshi K, Rosenthal R, Ghorani E, et al. Fc effector function contributes to the activity of human anti-CTLA-4 antibodies. *Cancer Cell* (2018) 33:649–663.e4. doi: 10.1016/j.ccell.2018.02.010

49. Dahan R, Sega E, Engelhardt J, Selby M, Korman AJ, Ravetch JV. FcyRs modulate the anti-tumor activity of antibodies targeting the PD-1/PD-L1 axis. *Cancer Cell* (2015) 28:285–95. doi: 10.1016/j.ccell.2015.08.004

50. Yuan Y, Lee JS, Yost SE, Frankel PH, Ruel C, Egelston CA, et al. Phase I/II trial of palbociclib, pembrolizumab and letrozole in patients with hormone receptor-positive metastatic breast cancer. *Eur J Cancer Oxf Engl 1990* (2021) 154:11–20. doi: 10.1016/ j.ejca.2021.05.035

51. Tolaney SM, Barroso-Sousa R, Keenan T, Li T, Trippa L, Vaz-Luis I, et al. Effect of eribulin with or without pembrolizumab on progression-free survival for patients with hormone receptor-positive, ERBB2-negative metastatic breast cancer: A randomized clinical trial. *JAMA Oncol* (2020) 6:1598–605. doi: 10.1001/jamaoncol.2020.3524

52. Pandey P, Khan F, Upadhyay TK, Sharangi AB. Deciphering the immunomodulatory role of cyclin-dependent kinase 4/6 inhibitors in the tumor microenvironment. *Int J Mol Sci* (2023) 24:2236. doi: 10.3390/ijms24032236

53. Huang H, Zhou J, Chen H, Li J, Zhang C, Jiang X, et al. The immunomodulatory effects of endocrine therapy in breast cancer. *J Exp Clin Cancer Res* (2021) 40:19. doi: 10.1186/s13046-020-01788-4

54. Maçon-Lemaître L, Triebel F. The negative regulatory function of the lymphocyte-activation gene-3 co-receptor (CD223) on human T cells. Immunology (2005) 115:170–8. doi: 10.1111/j.1365-2567.2005.02145.x

55. Brignone C, Gutierrez M, Mefti F, Brain E, Jarcau R, Cvitkovic F, et al. First-line chemoimmunotherapy in metastatic breast carcinoma: combination of paclitaxel and IMP321 (LAG-3Ig) enhances immune responses and antitumor activity. *J Transl Med* (2010) 8:71. doi: 10.1186/1479-5876-8-71

56. Wolf Y, Anderson AC, Kuchroo VK. TIM3 comes of age as an inhibitory receptor. *Nat Rev Immunol* (2020) 20:173–85. doi: 10.1038/s41577-019-0224-6

57. Dougall WC, Kurtulus S, Smyth MJ, Anderson AC. TIGIT and CD96: new checkpoint receptor targets for cancer immunotherapy. *Immunol Rev* (2017) 276:112–20. doi: 10.1111/imr.12518

58. Zong L, Mo S, Yu S, Zhou Y, Zhang M, Chen J, et al. Correction to: Expression of the immune checkpoint VISTA in breast cancer. *Cancer Immunol Immunother*. (2020) 69:1447–7. doi: 10.1007/s00262-020-02602-y

59. Le Mercier I, Chen W, Lines JL, Day M, Li J, Sergent P, et al. VISTA regulates the development of protective antitumor immunity. *Cancer Res* (2014) 74:1933–44. doi: 10.1158/0008-5472.CAN-13-1506

60. Janakiram M, Shah UA, Liu W, Zhao A, Schoenberg MP, Zang X. The third group of the B7-CD28 immune checkpoint family: HHLA2, TMIGD2, B7x, and B7-H3. *Immunol Rev* (2017) 276:26–39. doi: 10.1111/imr.12521

61. Deng J, Zhao S, Zhang X, Jia K, Wang H, Zhou C, et al. OX40 (CD134) and OX40 ligand, important immune checkpoints in cancer. *OncoTargets Ther* (2019) 12:7347–53. doi: 10.2147/OTT.S214211

62. Solinas C, Gu-Trantien C, Willard-Gallo K. The rationale behind targeting the ICOS-ICOS ligand costimulatory pathway in cancer immunotherapy. *ESMO Open* (2020) 5:e000544. doi: 10.1136/esmoopen-2019-000544

63. Zappasodi R, Sirard C, Li Y, Budhu S, Abu-Akeel M, Liu C, et al. Rational design of anti-GITR-based combination immunotherapy. *Nat Med* (2019) 25:759–66. doi: 10.1038/s41591-019-0420-8

64. Etxeberria I, Glez-Vaz J, Teijeira Á, Melero I. New emerging targets in cancer immunotherapy: CD137/4-1BB costimulatory axis. *ESMO Open* (2020) 4. doi: 10.1136/ esmoopen-2020-000733

65. Ku G, Bendell JC, Tolcher AW, Hurvitz SA, Krishnamurthy A, El-Khoueiry AB, et al. 525O A phase I dose escalation study of PRS-343, a HER2/4-1BB bispecific molecule, in patients with HER2-positive Malignancies. *Ann Oncol* (2020) 31:S462–3. doi: 10.1016/j.annonc.2020.08.639

66. Masu T, Atsukawa M, Nakatsuka K, Shimizu M, Miura D, Arai T, et al. Anti-CD137 monoclonal antibody enhances trastuzumab-induced, natural killer cellmediated cytotoxicity against pancreatic cancer cell lines with low human epidermal growth factor-like receptor 2 expression. *PloS One* (2018) 13:e0200664. doi: 10.1371/ journal.pone.0200664

67. Vonderheide RH. CD40 agonist antibodies in cancer immunotherapy. *Annu Rev Med* (2020) 71:47–58. doi: 10.1146/annurev-med-062518-045435

68. Harris DT, Kranz DM. Adoptive T cell therapies: A comparison of T cell receptors and chimeric antigen receptors. *Trends Pharmacol Sci* (2016) 37:220–30. doi: 10.1016/j.tips.2015.11.004

69. Hu W, Zi Z, Jin Y, Li G, Shao K, Cai Q, et al. CRISPR/Cas9-mediated PD-1 disruption enhances human mesothelin-targeted CAR T cell effector functions. *Cancer Immunol Immunother. CII* (2019) 68:365–77. doi: 10.1007/s00262-018-2281-2

70. Shah PD, Huang ACC, Xu X, Zhang PJ, Orlowski R, Matlawski T, et al. Phase I trial of autologous cMET-directed CAR-t cells administered intravenously in patients with melanoma & breast carcinoma. *J Clin Oncol* (2020) 38:10035–5. doi: 10.1200/ JCC.2020.38.15\_suppl.10035

71. Kudo K, Imai C, Lorenzini P, Kamiya T, Kono K, Davidoff AM, et al. T lymphocytes expressing a CD16 signaling receptor exert antibody-dependent cancer cell killing. *Cancer Res* (2014) 74:93–103. doi: 10.1158/0008-5472.CAN-13-1365

72. Han Y, Xie W, Song D-G, Powell DJ. Control of triple-negative breast cancer using ex vivo self-enriched, costimulated NKG2D CAR T cells. J Hematol Oncol.J Hematol Oncol (2018) 11:92. doi: 10.1186/s13045-018-0635-z

73. Wallstabe L, Göttlich C, Nelke LC, Kühnemundt J, Schwarz T, Nerreter T, et al. ROR1-CAR T cells are effective against lung and breast cancer in advanced microphysiologic 3D tumor models. *JCI Insight* (2019) 4(18). doi: 10.1172/jci.insight.126345

74. Venetis K, Invernizzi M, Sajjadi E, Curigliano G, Fusco N. Cellular immunotherapy in breast cancer: The quest for consistent biomarkers. *Cancer Treat Rev* (2020) 90:102089. doi: 10.1016/j.ctrv.2020.102089

75. Bernhard H, Neudorfer J, Gebhard K, Conrad H, Hermann C, Nährig J, et al. Adoptive transfer of autologous, HER2-specific, cytotoxic T lymphocytes for the treatment of HER2-overexpressing breast cancer. *Cancer Immunol Immunother CII* (2008) 57:271–80. doi: 10.1007/s00262-007-0355-7

76. Hardy NM, Mossoba ME, Steinberg SM, Fellowes V, Yan XY, Hakim FT, et al. Phase I trial of adoptive cell transfer with mixed-profile type-I/type-II allogeneic T cells for metastatic breast cancer. *Clin Cancer Res Off J Am Assoc Cancer Res* (2011) 17:6878– 87. doi: 10.1158/1078-0432.CCR-11-1579

77. Zacharakis N, Chinnasamy H, Black M, Xu H, Lu YC, Zheng Z, et al. Immune recognition of somatic mutations leading to complete durable regression in metastatic breast cancer. *Nat Med* (2018) 24:724–30. doi: 10.1038/s41591-018-0040-8

78. Hu Z. Tissue factor as a new target for CAR-NK cell immunotherapy of triplenegative breast cancer. *Sci Rep* (2020) 10:2815. doi: 10.1038/s41598-020-59736-3

79. Ternette N, Olde Nordkamp MJM, Müller J, Anderson AP, Nicastri A, Hill AVS, et al. Immunopeptidomic profiling of HLA-A2-positive triple negative breast cancer identifies potential immunotherapy target antigens. *Proteomics* (2018) 18(12): e1700465. doi: 10.1002/pmic.201700465

80. Xie YJ, Dougan M, Jailkhani N, Ingram J, Fang T, Kummer L, et al. Nanobody-based CAR T cells that target the tumor microenvironment inhibit the growth of solid tumors in immunocompetent mice. *Proc Natl Acad Sci U. S. A.* (2019) 116(16):7624–31. doi: 10.1073/pnas.1817147116

 Lanitis E, Poussin M, Klattenhoff AW, Song D, Sandaltzopoulos R, June CH, et al. Chimeric antigen receptor T Cells with dissociated signaling domains exhibit focused antitumor activity with reduced potential for toxicity. *vivo. Cancer Immunol Res* (2013) 1:43–53. doi: 10.1158/2326-6066.CIR-13-0008

82. Jäger M, Schoberth A, Ruf P, Hess J, Lindhofer H. The trifunctional antibody ertumaxomab destroys tumor cells that express low levels of human epidermal growth factor receptor 2. *Cancer Res* (2009) 69:4270–6. doi: 10.1158/0008-5472.CAN-08-2861

83. Liu J, Wu X, Lin L, Pan H, Wang Y, Li Y, et al. Bp-bs, a novel T-cell engaging bispecific antibody with biparatopic her2 binding, has potent anti-tumor activities. *Mol Ther Oncolytics* (2019) 14:66–73. doi: 10.1016/j.omto.2019.03.009

84. Lum LG, Thakur A, Al-Kadhimi Z, Colvin GA, Cummings FJ, Legare RD, et al. Targeted T-cell therapy in stage IV breast cancer: A phase I clinical trial. *Clin Cancer Res* (2015) 21:2305–14. doi: 10.1158/1078-0432.CCR-14-2280

85. Yi M, Wu Y, Niu M, Zhu S, Zhang J, Yan Y, et al. Anti-TGF-β/PD-L1 bispecific antibody promotes T cell infiltration and exhibits enhanced antitumor activity in triplenegative breast cancer. *J Immunother. Cancer* (2022) 10(12):e005543. doi: 10.1136/jitc-2022-005543

86. Harding JJ, Garrido-Laguna I, Chen X, Basu C, Dowlati A, Forgie A, et al. A phase 1 dose-escalation study of PF-06671008, a bispecific T-cell-engaging therapy targeting P-cadherin in patients with advanced solid tumors. *Front Immunol* (2022) 13:845417. doi: 10.3389/fimmu.2022.845417

87. Peoples GE, Holmes JP, Hueman MT, Mittendorf EA, Amin A, Khoo S, et al. Combined clinical trial results of a HER2/neu (E75) vaccine for the prevention of recurrence in high-risk breast cancer patients: U.S. Military Cancer Institute Clinical Trials Group Study I-01 and I-02. *Clin Cancer Res Off J Am Assoc Cancer Res* (2008) 14:797-803. doi: 10.1158/1078-0432.CCR-07-1448

88. Mittendorf EA, Clifton GT, Holmes JP, Schneble E, van Echo D, Ponniah S, et al. Final report of the phase I/II clinical trial of the E75 (nelipepimut-S) vaccine with booster inoculations to prevent disease recurrence in high-risk breast cancer patients. *Ann Oncol Off J Eur Soc Med Oncol* (2014) 25:1735–42. doi: 10.1093/annonc/mdu211

89. Mittendorf EA, Lu B, Melisko M, Hiller JP, Bondarenko I, Brunt AM, et al. Efficacy and safety analysis of nelipepimut-S vaccine to prevent breast cancer recurrence: A randomized, multicenter, phase III clinical trial. *Clin Cancer Res* (2019) 25:4248-54. doi: 10.1158/1078-0432.CCR-18-2867

90. Clifton GT, Hale D, Vreeland TJ, Hickerson AT, Litton JK, Alatrash G, et al. Results of a randomized phase IIb trial of nelipepimut-S + Trastuzumab versus trastuzumab to prevent recurrences in patients with high-risk HER2 low-expressing breast cancer. *Clin Cancer Res Off J Am Assoc Cancer Res* (2020) 26:2515-23. doi: 10.1158/1078-0432.CCR-19-2741

91. Dafni U, Martín-Lluesma S, Balint K, Tsourti Z, Vervita K, Chenal J, et al. Efficacy of cancer vaccines in selected gynaecological breast and ovarian cancers: A 20year systematic review and meta-analysis. *Eur J Cancer Oxf. Engl 1990* (2021) 142:63–82. doi: 10.1016/j.ejca.2020.10.014

92. Brown TA, Mittendorf EA, Hale DF, Myers JW, Peace KM, Jackson DO, et al. Prospective, randomized, single-blinded, multi-center phase II trial of two HER2 peptide vaccines, GP2 and AE37, in breast cancer patients to prevent recurrence. *Breast Cancer Res Treat* (2020) 181:391–401. doi: 10.1007/s10549-020-05638-x

93. Kalli KR, Block MS, Kasi PM, Erskine CL, Hobday TJ, Dietz A, et al. Folate receptor alpha peptide vaccine generates immunity in breast and ovarian cancer patients. *Clin Cancer Res* (2018) 24:3014–25. doi: 10.1158/1078-0432.CCR-17-2499

94. Gilewski TA, Ragupathi G, Dickler M, Powell S, Bhuta S, Panageas K, et al. Immunization of high-risk breast cancer patients with clustered sTn-KLH conjugate plus the immunologic adjuvant QS-21. *Clin Cancer Res Off J Am Assoc Cancer Res* (2007) 13:2977–85. doi: 10.1158/1078-0432.CCR-06-2189

95. Apostolopoulos V, Pietersz GA, Tsibanis A, Tsikkinis A, Drakaki H, Loveland BE, et al. Pilot phase III immunotherapy study in early-stage breast cancer patients using oxidized mannan-MUC1 [ISRCTN71711835]. *Breast Cancer Res BCR* (2006) 8: R27. doi: 10.1186/bcr1505

96. Ibrahim NK, Murray JL, Zhou D, Mittendorf EA, Sample D, Tautchin M, et al. Survival advantage in patients with metastatic breast cancer receiving endocrine therapy plus sialyl tn-KLH vaccine: *post hoc* analysis of a large randomized trial. *J Cancer* (2013) 4:577–84. doi: 10.7150/jca.7028

97. Hamilton E, Blackwell K, Hobeika AC, Clay TM, Broadwater G, Ren XR, et al. Phase 1 clinical trial of HER2-specific immunotherapy with concomitant HER2 kinase inhibition [corrected]. *J Transl Med* (2012) 10:28. doi: 10.1186/1479-5876-10-28

98. Ophir E, Bobisse S, Coukos G, Harari A, Kandalaft LE. Personalized approaches to active immunotherapy in cancer. *Biochim Biophys Acta* (2016) 1865:72–82. doi: 10.1016/j.bbcan.2015.07.004

99. Kurtz SL, Ravindranathan S, Zaharoff DA. Current status of autologous breast tumor cell-based vaccines. *Expert Rev Vaccines* (2014) 13:1439–45. doi: 10.1586/14760584.2014.969714

100. Niavarani S-R, Lawson C, Boudaud M, Simard C, Tai L-H. Oncolytic vesicular stomatitis virus–based cellular vaccine improves triple-negative breast cancer outcome by enhancing natural killer and CD8+ T-cell functionality. *J Immunother. Cancer* (2020) 8:e000465. doi: 10.1136/jitc-2019-000465

101. Srivatsan S, Patel JM, Bozeman EN, Imasuen IE, He S, Daniels D, et al. Allogeneic tumor cell vaccines: the promise and limitations in clinical trials. *Hum Vaccines Immunother*. (2014) 10:52–63. doi: 10.4161/hv.26568

102. O'Rourke DM, Nasrallah MP, Desai A, Melenhorst JJ, Mansfield K, Morrissette JJD, et al. A single dose of peripherally infused EGFRvIII-directed CAR T cells mediates antigen loss and induces adaptive resistance in patients with recurrent glioblastoma. *Sci Transl Med* (2017) 9(399). doi: 10.1126/scitranslmed.aaa0984

103. Anderson KS, Erick TK, Chen M, Daley H, Campbell M, Colson Y, et al. The feasibility of using an autologous GM-CSF-secreting breast cancer vaccine to induce immunity in patients with stage II-III and metastatic breast cancers. *Breast Cancer Res Treat* (2022) 194:65–78. doi: 10.1007/s10549-022-06562-y

104. Emens LA, Asquith JM, Leatherman JM, Kobrin BJ, Petrik S, Laiko M, et al. Timed sequential treatment with cyclophosphamide, doxorubicin, and an allogeneic granulocyte-macrophage colony-stimulating factor-secreting breast tumor vaccine: a chemotherapy dose-ranging factorial study of safety and immune activation. J Clin Oncol Off J Am Soc Clin Oncol (2009) 27:5911–8. doi: 10.1200/JCO.2009.23.3494

105. Williams W, Holmes JP, Bhattacharya S, Calfa C, Dakhil SR, Lukas JJ, et al. Safety and efficacy of a phase I/IIa trial (NCT03066947) of a modified whole tumor cell targeted immunotherapy in patients with advanced breast cancer. *J Clin Oncol* (2019) 37:e14026–6. doi: 10.1200/JCO.2019.37.15\_suppl.e14026

106. Laeufle R, Maglakelidze M, Bulat I, Ryspayeva D, Zoran A, Chawla T, et al. HERIZON: A phase 1B/2 open-label study of imu-131 HER2/neu peptide vaccine PLUS standard of care chemotherapy with randomization in phase 2 in patients with HER2/neu overexpressing metastatic or advanced adenocarcinoma of the stomach or gastroesophageal junction. *J Clin Oncol* (2021) 39:e16065–5. doi: 10.1200/JCO.2021.39.15\_suppl.e16065

107. Bekaii-Saab T, Wesolowski R, Ahn DH, Wu C, Mortazavi A, Lustberg M, et al. Phase I immunotherapy trial with two chimeric HER-2 B-cell peptide vaccines emulsified in montanide ISA 720VG and nor-MDP adjuvant in patients with advanced solid tumors. *Clin Cancer Res Off J Am Assoc Cancer Res* (2019) 25:3495– 507. doi: 10.1158/1078-0432.CCR-18-3997

108. Ribas A. Adaptive immune resistance: How cancer protects from immune attack. *Cancer Discovery* (2015) 5:915–9. doi: 10.1158/2159-8290.CD-15-0563

109. Lin M, Liang S, Jiang F, Xu J, Zhu W, Qian W, et al. 2003-2013, a valuable study: Autologous tumor lysate-pulsed dendritic cell immunotherapy with cytokine-induced killer cells improves survival in stage IV breast cancer. *Immunol Lett* (2017) 183:37–43. doi: 10.1016/j.imlet.2017.01.014

110. Qi C-J, Ning YL, Han YS, Min HY, Ye H, Zhu YL, et al. Autologous dendritic cell vaccine for estrogen receptor (ER)/progestin receptor (PR) double-negative breast cancer. *Cancer Immunol Immunother. CII* (2012) 61:1415–24. doi: 10.1007/s00262-011-1192-2

111. Santisteban M, Solans BP, Hato L, Urrizola A, Mejías LD, Salgado E, et al. Final results regarding the addition of dendritic cell vaccines to neoadjuvant chemotherapy in early HER2-negative breast cancer patients: clinical and translational analysis. *Ther Adv Med Oncol* (2021) 13:17588359211064653. doi: 10.1177/17588359211064653

112. Park JW, Melisko ME, Esserman LJ, Jones LA, Wollan JB, Sims R. Treatment with autologous antigen-presenting cells activated with the HER-2 based antigen Lapuleucel-T: results of a phase I study in immunologic and clinical activity in HER-2 overexpressing breast cancer. *J Clin Oncol Off J Am Soc Clin Oncol* (2007) 25:3680–7. doi: 10.1200/JCO.2006.10.5718

113. Svane IM, Pedersen AE, Johansen JS, Johnsen HE, Nielsen D, Kamby C, et al. Vaccination with p53 peptide-pulsed dendritic cells is associated with disease stabilization in patients with p53 expressing advanced breast cancer; monitoring of serum YKL-40 and IL-6 as response biomarkers. *Cancer Immunol Immunother. CII* (2007) 56:1485–99. doi: 10.1007/s00262-007-0293-4

114. Salazar LG, Lu H, Reichow JL, Childs JS, Coveler AL, Higgins DM, et al. Topical imiquimod plus nab-paclitaxel for breast cancer cutaneous metastases. *JAMA Oncol* (2017) 3:969–73. doi: 10.1001/jamaoncol.2016.6007

115. Dudek AZ, Yunis C, Harrison LI, Kumar S, Hawkinson R, Cooley S, et al. First in human phase I trial of 852A, a novel systemic toll-like receptor 7 agonist, to activate innate immune responses in patients with advanced cancer. *Clin Cancer Res Off J Am Assoc Cancer Res* (2007) 13:7119–25. doi: 10.1158/1078-0432.CCR-07-1443

116. Tiriveedhi V, Tucker N, Herndon J, Li L, Sturmoski M, Ellis M, et al. Safety and preliminary evidence of biologic efficacy of a mammaglobin-a DNA vaccine in patients with stable metastatic breast cancer. *Clin Cancer Res Off J Am Assoc Cancer Res* (2014) 20:5964–75. doi: 10.1158/1078-0432.CCR-14-0059

117. Disis ML, Guthrie KA, Liu Y, Coveler AL, Higgins DM, Childs JS, et al. Safety and outcomes of a plasmid DNA vaccine encoding the ERBB2 intracellular domain in patients with advanced-stage ERBB2-positive breast cancer: A phase 1 nonrandomized clinical trial. *JAMA Oncol* (2023) 9:71–8. (Nora). doi: 10.1001/jamaoncol.2022.5143

118. Zhang X, Goedegebuure SP, Myers NB, Vickery T, McLellan MD, Gao F, et al. Neoantigen DNA vaccines are safe, feasible, and capable of inducing neoantigenspecific immune responses in patients with triple negative breast cancer. *medRxiv* (2021), p. 2021.11.19.21266466. Available at: https://www.medrxiv.org/content/10. 1101/2021.11.19.21266466v1.

119. Crosby EJ, Gwin W, Blackwell K, Marcom PK, Chang S, Maecker HT, et al. Vaccine-Induced Memory CD8+ T Cells Provide Clinical Benefit in HER2 Expressing Breast Cancer: A Mouse to Human Translational Study. *Clin Cancer Res Off J Am Assoc Cancer Res* (2019) 25:2725–36. doi: 10.1158/1078-0432.CCR-18-3102

120. Park JH, Lee HK. Function of  $\gamma\delta$  T cells in tumor immunology and their application to cancer therapy. *Exp Mol Med* (2021) 53:318–27. doi: 10.1038/s12276-021-00576-0

121. Xu Y, Xiang Z, Alnaggar M, Kouakanou L, Li J, He J, et al. Allogeneic Vγ9Vδ2 T-cell immunotherapy exhibits promising clinical safety and prolongs the survival of patients with late-stage lung or liver cancer. *Cell Mol Immunol* (2021) 18:427–39. doi: 10.1038/s41423-020-0515-7

122. Morrow ES, Roseweir A, Edwards J. The role of gamma delta T lymphocytes in breast cancer: a review. *Transl Res J Lab Clin Med* (2019) 203:88–96. doi: 10.1016/j.trsl.2018.08.005

123. Hamilton E, Clay TM, Blackwell KL. New perspectives on zoledronic acid in breast cancer: potential augmentation of anticancer immune response. *Cancer Invest.* (2011) 29:533–41. doi: 10.3109/07357907.2011.605413

124. Capietto A-H, Martinet L, Fournié J-J. Stimulated γδ T cells increase the in vivo efficacy of trastuzumab in HER-2+ breast cancer. J Immunol Baltim Md 1950 (2011) 187:1031–8. doi: 10.4049/jimmunol.1100681

125. Wu Y, Kyle-Cezar F, Woolf RT, Naceur-Lombardelli C, Owen J, Biswas D, et al. An innate-like V $\delta$ 1+  $\gamma\delta$  T cell compartment in the human breast is associated with remission in triple-negative breast cancer. *Sci Transl Med* (2019) 11(513). doi: 10.1126/scitranslmed.aax9364

126. Peng G, Wang HY, Peng W, Kiniwa Y, Seo KH, Wang RF. Tumor-infiltrating gammadelta T cells suppress T and dendritic cell function *via* mechanisms controlled by a unique toll-like receptor signaling pathway. *Immunity* (2007) 27:334–48. doi: 10.1016/j.immuni.2007.05.020

127. Meraviglia S, Eberl M, Vermijlen D, Todaro M, Buccheri S, Cicero G, et al. *In vivo* manipulation of Vgamma9Vdelta2 T cells with zoledronate and low-dose interleukin-2 for immunotherapy of advanced breast cancer patients. *Clin Exp Immunol* (2010) 161:290–7. doi: 10.1111/j.1365-2249.2010.04167.x

128. Bennouna J, Levy V, Sicard H, Senellart H, Audrain M, Hiret S, et al. Phase I study of bromohydrin pyrophosphate (BrHPP, IPH 1101), a Vgamma9Vdelta2 T lymphocyte agonist in patients with solid tumors. *Cancer Immunol Immunother CII* (2010) 59:1521–30. doi: 10.1007/s00262-010-0879-0

129. Janssen A, Villacorta Hidalgo J, Beringer DX, van Dooremalen S, Fernando F, van Diest E, et al.  $\gamma\delta$  T-cell receptors derived from breast cancer-infiltrating T lymphocytes mediate antitumor reactivity. *Cancer Immunol Res* (2020) 8:530–43. doi: 10.1158/2326-6066.CIR-19-0513

130. Shi Y, Fan X, Deng H, Brezski RJ, Rycyzyn M, Jordan RE, et al. Trastuzumab triggers phagocytic killing of high HER2 cancer cells in *vitro* and in *vivo* by interaction with Fcγ receptors on macrophages. *J Immunol Baltim. Md* 1950 (2015) 194:4379–86. doi: 10.4049/jimmunol.1402891

131. Brandsma AM, Ten Broeke T, Nederend M, Meulenbroek LAPM, van Tetering G, Meyer S, et al. Simultaneous targeting of fc $\gamma$ Rs and fc $\alpha$ RI enhances tumor cell killing. *Cancer Immunol Res* (2015) 3:1316–24. doi: 10.1158/2326-6066.CIR-15-0099-T

132. Biswas SK, Allavena P, Mantovani A. Tumor-associated macrophages: functional diversity, clinical significance, and open questions. *Semin Immunopathol* (2013) 35:585–600. doi: 10.1007/s00281-013-0367-7

133. Ni C, Yang L, Xu Q, Yuan H, Wang W, Xia W, et al. CD68- and CD163-positive tumor infiltrating macrophages in non-metastatic breast cancer: a retrospective study and meta-analysis. *J Cancer* (2019) 10:4463–72. doi: 10.7150/jca.33914

134. Ali HR, Chlon L, Pharoah PDP, Markowetz F, Caldas C. Patterns of immune infiltration in breast cancer and their clinical implications: A gene-expression-based retrospective study. *PloS Med* (2016) 13:e1002194. doi: 10.1371/journal.pmed.1002194

135. Xuan Q, Wang JX, Nanding A, Wang ZP, Liu H, Lian X, et al. Tumorassociated macrophages are correlated with tamoxifen resistance in the postmenopausal breast cancer patients. *Pathol Oncol Res POR* (2014) 20:619–24. doi: 10.1007/s12253-013-9740-z

136. De Palma M, Lewis CE. Macrophage regulation of tumor responses to anticancer therapies. *Cancer Cell* (2013) 23:277–86. doi: 10.1016/j.ccr.2013.02.013

137. Sikic BI, Lakhani N, Patnaik A, Shah SA, Chandana SR, Rasco D, et al. First-inhuman, first-in-class phase I trial of the anti-CD47 antibody hu5F9-G4 in patients with advanced cancers. J Clin Oncol Off J Am Soc Clin Oncol (2019) 37:946–53. doi: 10.1200/ JCO.18.02018

138. Advani R, Flinn I, Popplewell L, Forero A, Bartlett NL, Ghosh N, et al. CD47 blockade by hu5F9-G4 and rituximab in non-hodgkin's lymphoma. *N Engl J Med* (2018) 379:1711–21. doi: 10.1056/NEJMoa1807315

139. Lakhani NJ, Chow LQM, Gainor JF, LoRusso P, Lee KW, Chung HC, et al. Evorpacept alone and in combination with pembrolizumab or trastuzumab in patients with advanced solid tumours (ASPEN-01): a first-in-human, open-label, multicentre, phase 1 dose-escalation and dose-expansion study. *Lancet Oncol* (2021) 22:1740–51. doi: 10.1016/S1470-2045(21)00584-2

140. Barkal AA, Brewer RE, Markovic M, Kowarsky M, Barkal SA, Zaro BW, et al. CD24 signalling through macrophage Siglec-10 is a target for cancer immunotherapy. *Nature* (2019) 572:392–6. doi: 10.1038/s41586-019-1456-0

141. Kitamura T, Qian BZ, Soong D, Cassetta L, Noy R, Sugano G, et al. CCL2induced chemokine cascade promotes breast cancer metastasis by enhancing retention of metastasis-associated macrophages. *J Exp Med* (2015) 212:1043–59. doi: 10.1084/ jem.20141836

142. Richardsen E, Uglehus RD, Johnsen SH, Busund L-T. Macrophage-colony stimulating factor (CSF1) predicts breast cancer progression and mortality. *Anticancer Res* (2015) 35:865–74.

143. Germano G, Frapolli R, Belgiovine C, Anselmo A, Pesce S, Liguori M, et al. Role of macrophage targeting in the antitumor activity of trabectedin. *Cancer Cell* (2013) 23:249–62. doi: 10.1016/j.ccr.2013.01.008

144. Pelekanou V, Villarroel-Espindola F, Schalper KA, Pusztai L, Rimm DL. CD68, CD163, and matrix metalloproteinase 9 (MMP-9) co-localization in breast tumor microenvironment predicts survival differently in ER-positive and -negative cancers. *Breast Cancer Res* (2018) 20:154. doi: 10.1186/s13058-018-1076-x

145. Goldstein LJ, Gurtler J, Prete SAD, Tjulandin S, Semiglazov VF, Bayever E, et al. Trabectedin as a single-agent treatment of advanced breast cancer after anthracycline and taxane treatment: A multicenter, randomized, phase II study comparing 2 administration regimens. *Clin Breast Cancer* (2014) 14:396–404. doi: 10.1016/j.clbc.2014.06.006

146. Blum JL, Gonçalves A, Efrat N, Debled M, Conte P, Richards PD, et al. A phase II trial of trabectedin in triple-negative and HER2-overexpressing metastatic breast cancer. *Breast Cancer Res Treat* (2016) 155:295–302. doi: 10.1007/s10549-015-3675-x

147. Costa L. Bisphosphonates in adjuvant setting for breast cancer: a review of the meta-analysis of bisphosphonates' effects on breast cancer recurrence presented in December 2013 at San Antonio Breast Conference. *Curr Opin Support Palliat Care* (2014) 8:414–9. doi: 10.1097/SPC.00000000000101

148. Panagiotakou A, Yavropoulou M, Nasiri-Ansari N, Makras P, Basdra EK, Papavassiliou AG, et al. Extra-skeletal effects of bisphosphonates. *Metabolism* (2020) 110:154264. doi: 10.1016/j.metabol.2020.154264

149. Shiota T, Miyasato Y, Ohnishi K, Yamamoto-Ibusuki M, Yamamoto Y, Iwase H, et al. The clinical significance of CD169-positive lymph node macrophage in patients with breast cancer. *PloS One* (2016) 11:e0166680. doi: 10.1371/journal.pone.0166680

150. Honkanen TJ, Tikkanen A, Karihtala P, Mäkinen M, Väyrynen JP, Koivunen JP. Prognostic and predictive role of tumour-associated macrophages in HER2 positive breast cancer. *Sci Rep* (2019) 9:10961. doi: 10.1038/s41598-019-47375-2

151. Jeong H, Hwang I, Kang SH, Shin HC, Kwon SY. Tumor-associated macrophages as potential prognostic biomarkers of invasive breast cancer. J Breast Cancer (2019) 22:38–51. doi: 10.4048/jbc.2019.22.e5

152. Shiraishi D, Fujiwara Y, Horlad H, Saito Y, Iriki T, Tsuboki J, et al. CD163 is required for protumoral activation of macrophages in human and murine sarcoma. *Cancer Res* (2018) 78:3255–66. doi: 10.1158/0008-5472.CAN-17-2011

153. Bhusari P, Bhatt J, Sood A, Kaur R, Vatsa R, Rastogi A, et al. Evaluating the potential of kit-based 68Ga-ubiquicidin formulation in diagnosis of infection: a pilot study68Ga. *Nucl Med Commun* (2019) 40:228–34. doi: 10.1097/MNM.00000000000943

154. Gebhart G, Lamberts LE, Wimana Z, Garcia C, Emonts P, Ameye L, et al. Molecular imaging as a tool to investigate heterogeneity of advanced HER2-positive breast cancer and to predict patient outcome under trastuzumab emtansine (T-DM1): the ZEPHIR trial. *Ann Oncol Off J Eur Soc Med Oncol* (2016) 27:619–24. doi: 10.1093/ annonc/mdv577

155. Taralli S, Lorusso M, Perrone E, Perotti G, Zagaria L, Calcagni ML. PET/CT with fibroblast activation protein inhibitors in breast cancer: diagnostic and theranostic application-A literature review. *Cancers* (2023) 15:908. doi: 10.3390/cancers15030908

156. Ballal S, Yadav MP, Kramer V, Moon ES, Roesch F, Tripathi M, et al. A theranostic approach of [68Ga]Ga-DOTA.SA.FAPi PET/CT-guided [177Lu]Lu-DOTA.SA.FAPi radionuclide therapy in an end-stage breast cancer patient: new frontier in targeted radionuclide therapy. *Eur J Nucl Med Mol Imaging* (2021) 48:942–4. doi: 10.1007/s00259-020-04990-w

157. Sahin TT, Kasuya H, Nomura N, Shikano T, Yamamura K, Gewen T, et al. Impact of novel oncolytic virus HF10 on cellular components of the tumor microenviroment in patients with recurrent breast cancer. *Cancer Gene Ther* (2012) 19:229–37. doi: 10.1038/cgt.2011.80

158. Bernstein V, Ellard SL, Dent SF, Tu D, Mates M, Dhesy-Thind SK, et al. A randomized phase II study of weekly paclitaxel with or without pelareorep in patients with metastatic breast cancer: final analysis of Canadian Cancer Trials Group IND.213. *Breast Cancer Res Treat* (2018) 167:485–93. doi: 10.1007/s10549-017-4538-4

159. Bramante S, Koski A, Liikanen I, Vassilev L, Oksanen M, Siurala M, et al. Oncolytic virotherapy for treatment of breast cancer, including triple-negative breast cancer. *Oncoimmunology* (2016) 5:e1078057. doi: 10.1080/2162402X.2015.1078057

160. Bourgeois-Daigneault M-C, Roy DG, Aitken AS, El Sayes N, Martin NT, Varette O, et al. Neoadjuvant oncolytic virotherapy before surgery sensitizes triplenegative breast cancer to immune checkpoint therapy. *Sci Transl Med* (2018) 10(422). doi: 10.1126/scitranslmed.aao1641

161. Liikanen I, Tähtinen S, Guse K, Gutmann T, Savola P, Oksanen M, et al. Oncolytic adenovirus expressing monoclonal antibody trastuzumab for treatment of HER2-positive cancer. *Mol Cancer Ther* (2016) 15:2259–69. doi: 10.1158/1535-7163.MCT-15-0819

162. Braybrooke JP, Slade A, Deplanque G, Harrop R, Madhusudan S, Forster MD, et al. Phase I study of MetXia-P450 gene therapy and oral cyclophosphamide for patients with advanced breast cancer or melanoma. *Clin Cancer Res Off J Am Assoc Cancer Res* (2005) 11:1512–20. doi: 10.1158/1078-0432.CCR-04-0155

163. Duggan MC, Jochems C, Donahue RN, Richards J, Karpa V, Foust E, et al. A phase I study of recombinant (r) vaccinia-CEA(6D)-TRICOM and rFowlpox-CEA (6D)-TRICOM vaccines with GM-CSF and IFN-α-2b in patients with CEA-expressing carcinomas. *Cancer Immunol Immunother CII* (2016) 65:1353–64. doi: 10.1007/s00262-016-1893-7

164. Formenti SC, Lee P, Adams S, Goldberg JD, Li X, Xie MW, et al. Focal irradiation and systemic TGF $\beta$  Blockade in metastatic breast cancer. *Clin Cancer Res Off J Am Assoc Cancer Res* (2018) 24:2493–504. doi: 10.1158/1078-0432.CCR-17-3322

165. Secondini C, Coquoz O, Spagnuolo I, Spinetti T, Peyvandi S, Ciarloni L, et al. Arginase inhibition suppresses lung metastasis in the 4T1 breast cancer model independently of the immunomodulatory and anti-metastatic effects of VEGFR-2 blockade. *Oncoimmunology* (2017) 6:e1316437. doi: 10.1080/2162402X.2017.1316437

166. Gao J, Deng F, Jia W. Inhibition of indoleamine 2,3-dioxygenase enhances the therapeutic efficacy of immunogenic chemotherapeutics in breast cancer. J Breast Cancer (2019) 22:196–209. doi: 10.4048/jbc.2019.22.e23

167. Su X, Xu Y, Fox GC, Xiang J, Kwakwa KA, Davis JL, et al. Breast cancer-derived GM-CSF regulates arginase 1 in myeloid cells to promote an immunosuppressive microenvironment. *J Clin Invest* (2021) 131:e145296. doi: 10.1172/JCI145296

168. Mariotti V, Han H, Ismail-Khan R, Tang SC, Dillon P, Montero AJ, et al. Effect of taxane chemotherapy with or without indoximod in metastatic breast cancer: A randomized clinical trial. *JAMA Oncol* (2021) 7:61–9. doi: 10.1001/jamaoncol.2020.5572

169. Soliman H, Khambati F, Han HS, Ismail-Khan R, Bui MM, Sullivan DM, et al. A phase-1/2 study of adenovirus-p53 transduced dendritic cell vaccine in combination with indoximod in metastatic solid tumors and invasive breast cancer. *Oncotarget* (2018) 9:10110–7. doi: 10.18632/oncotarget.24118

170. Savva C, Copson E, Johnson PWM, Cutress RI, Beers SA. Obesity is associated with immunometabolic changes in adipose tissue that may drive treatment resistance in breast cancer: immune-metabolic reprogramming and novel therapeutic strategies. *Cancers* (2023) 15:2440. doi: 10.3390/cancers15092440

171. Cortellino S, Raveane A, Chiodoni C, Delfanti G, Pisati F, Spagnolo V, et al. Fasting renders immunotherapy effective against low-immunogenic breast cancer while reducing side effects. *Cell Rep* (2022) 40:111256. doi: 10.1016/j.celrep.2022.111256

172. Marinac CR, Nelson SH, Breen CI, Hartman SJ, Natarajan L, Pierce JP, et al. Prolonged nightly fasting and breast cancer prognosis. *JAMA Oncol* (2016) 2:1049–55. doi: 10.1001/jamaoncol.2016.0164

173. Fu Z, Mowday AM, Smaill JB, Hermans IF, Patterson AV. Tumour hypoxiamediated immunosuppression: mechanisms and therapeutic approaches to improve cancer immunotherapy. *Cells* (2021) 10:1006. doi: 10.3390/cells10051006

174. Ohta A. A metabolic immune checkpoint: adenosine in tumor microenvironment. Front Immunol (2016) 7:109. doi: 10.3389/fimmu.2016.00109

175. Yin T-T, Huang MX, Wang F, Jiang YH, Long J, Li L, et al. Lactate score predicts survival, immune cell infiltration and response to immunotherapy in breast cancer. *Front Genet* (2022) 13. doi: 10.3389/fgene.2022.943849

176. Leone RD, Emens LA. Targeting adenosine for cancer immunotherapy. J Immunother. Cancer (2018) 6:57. doi: 10.1186/s40425-018-0360-8

177. Fendl B, Berghoff AS, Preusser M, Maier B. Macrophage and monocyte subsets as new therapeutic targets in cancer immunotherapy. *ESMO Open* (2023) 8:100776. doi: 10.1016/j.esmoop.2022.100776

178. Wang S, Yang Y, Ma P, Huang H, Tang Q, Miao H, et al. Landscape and perspectives of macrophage -targeted cancer therapy in clinical trials. *Mol Ther - Oncolytics* (2022) 24:799–813. doi: 10.1016/j.omto.2022.02.019

179. Wu S-Y, Zhang SW, Ma D, Xiao Y, Liu Y, Chen L, et al. CCL19+ dendritic cells potentiate clinical benefit of anti-PD-(L)1 immunotherapy in triple-negative breast cancer. *Med N Y. N* (2023) 4:373–393.e8. doi: 10.1016/j.medj.2023.04.008

180. House IG, Savas P, Lai J, Chen AXY, Oliver AJ, Teo ZL, et al. Macrophagederived CXCL9 and CXCL10 are required for antitumor immune responses following immune checkpoint blockade. *Clin Cancer Res* (2020) 26:487–504. doi: 10.1158/1078-0432.CCR-19-1868

181. Gavrielatou N, Liu Y, Vathiotis I, Zugazagoitia J, Aung TN, Shafi S, et al. Association of PD-1/PD-L1 co-location with immunotherapy outcomes in non-small cell lung cancer. *Clin Cancer Res Off J Am Assoc Cancer Res* (2022) 28:360–7. doi: 10.1158/1078-0432.CCR-21-2649

182. Zeng D, Ye Z, Wu J, Zhou R, Fan X, Wang G, et al. Macrophage correlates with immunophenotype and predicts anti-PD-L1 response of urothelial cancer. *Theranostics* (2020) 10:7002–14. doi: 10.7150/thno.46176

183. Fucikova J, Kepp O, Kasikova L, Petroni G, Yamazaki T, Liu P, et al. Detection of immunogenic cell death and its relevance for cancer therapy. *Cell Death Dis* (2020) 11:1–13. doi: 10.1038/s41419-020-03221-2

184. Barker HE, Paget JTE, Khan AA, Harrington KJ. The tumour microenvironment after radiotherapy: mechanisms of resistance and recurrence. *Nat Rev Cancer* (2015) 15:409-25. doi: 10.1038/nrc3958

185. Aldea M, Orillard E, Mansi L, Marabelle A, Scotte F, Lambotte O, et al. How to manage patients with corticosteroids in oncology in the era of immunotherapy? *Eur J Cancer Oxf Engl 1990* (2020) 141:239–51. doi: 10.1016/j.ejca.2020.09.032

186. Wein L, Savas P, Luen SJ, Virassamy B, Salgado R, Loi S. Clinical validity and utility of tumor-infiltrating lymphocytes in routine clinical practice for breast cancer patients: current and future directions. *Front Oncol* (2017) 7:156. doi: 10.3389/ fonc.2017.00156

187. Ibrahim EM, Al-Foheidi ME, Al-Mansour MM, Kazkaz GA. The prognostic value of tumor-infiltrating lymphocytes in triple-negative breast cancer: a meta-analysis. *Breast Cancer Res Treat* (2014) 148:467–76. doi: 10.1007/s10549-014-3185-2

188. Chraa D, Naim A, Olive D, Badou A. T lymphocyte subsets in cancer immunity: Friends or foes. J Leukoc. Biol (2019) 105:243-55. doi: 10.1002/JLB.MR0318-097R

189. Robert C. A decade of immune-checkpoint inhibitors in cancer therapy. *Nat Commun* (2020) 11:3801. doi: 10.1038/s41467-020-17670-y