

Dear Author,

Here are the proofs of your article.

- You can submit your corrections **online**, via **e-mail** or by **fax**.
- For **online** submission please insert your corrections in the online correction form. Always indicate the line number to which the correction refers.
- You can also insert your corrections in the proof PDF and **email** the annotated PDF.
- For fax submission, please ensure that your corrections are clearly legible. Use a fine black pen and write the correction in the margin, not too close to the edge of the page.
- Remember to note the **journal title**, **article number**, and **your name** when sending your response via e-mail or fax.
- **Check** the metadata sheet to make sure that the header information, especially author names and the corresponding affiliations are correctly shown.
- **Check** the questions that may have arisen during copy editing and insert your answers/ corrections.
- **Check** that the text is complete and that all figures, tables and their legends are included. Also check the accuracy of special characters, equations, and electronic supplementary material if applicable. If necessary refer to the *Edited manuscript*.
- The publication of inaccurate data such as dosages and units can have serious consequences. Please take particular care that all such details are correct.
- Please **do not** make changes that involve only matters of style. We have generally introduced forms that follow the journal's style. Substantial changes in content, e.g., new results, corrected values, title and authorship are not allowed without the approval of the responsible editor. In such a case, please contact the Editorial Office and return his/her consent together with the proof.
- If we do not receive your corrections **within 48 hours**, we will send you a reminder.
- Your article will be published **Online First** approximately one week after receipt of your corrected proofs. This is the **official first publication** citable with the DOI. **Further changes are, therefore, not possible.**
- The **printed version** will follow in a forthcoming issue.

#### **Please note**

After online publication, subscribers (personal/institutional) to this journal will have access to the complete article via the DOI using the URL: [http://dx.doi.org/\[DOI\]](http://dx.doi.org/[DOI]).

If you would like to know when your article has been published online, take advantage of our free alert service. For registration and further information go to: <http://www.springerlink.com>.

Due to the electronic nature of the procedure, the manuscript and the original figures will only be returned to you on special request. When you return your corrections, please inform us if you would like to have these documents returned.

# Metadata of the article that will be visualized in OnlineFirst

ArticleTitle	Persistence of critical flicker fusion frequency impairment after a 33 mfw SCUBA dive: evidence of prolonged nitrogen narcosis?	
Article Sub-Title		
Article CopyRight	Springer-Verlag (This will be the copyright line in the final PDF)	
Journal Name	European Journal of Applied Physiology	
Corresponding Author	Family Name	<b>Lafère</b>
	Particle	
	Given Name	<b>P.</b>
	Suffix	
	Division	
	Organization	DAN Europe Research Division
	Address	Brussels, Belgium
	Division	
	Organization	Environmental and Occupational Physiology Laboratory, Haute Ecole "Paul Henri Spaak"
	Address	Brussels, Belgium
	Division	Centre For Hyperbaric Oxygen Therapy
	Organization	Military Hospital "Queen Astrid"
	Address	Bruynstreet 1, Brussels, 1120, Belgium
	Email	doc.lafere@skynet.be
Author	Family Name	<b>Balestra</b>
	Particle	
	Given Name	<b>C.</b>
	Suffix	
	Division	
	Organization	DAN Europe Research Division
	Address	Brussels, Belgium
	Division	
	Organization	Environmental and Occupational Physiology Laboratory, Haute Ecole "Paul Henri Spaak"
	Address	Brussels, Belgium
	Email	daneuben@skynet.be
Author	Family Name	<b>Germonpré</b>
	Particle	
	Given Name	<b>P.</b>
	Suffix	
	Division	
	Organization	DAN Europe Research Division
	Address	Brussels, Belgium
	Division	
	Organization	Environmental and Occupational Physiology Laboratory, Haute Ecole "Paul Henri Spaak"

Address	Brussels, Belgium
Division	Centre For Hyperbaric Oxygen Therapy
Organization	Military Hospital "Queen Astrid"
Address	Bruynstreet 1, Brussels, 1120, Belgium
Email	peter.germonpre@mil.be

---

Schedule	Received	29 June 2011
	Revised	
	Accepted	19 March 2012

---

**Abstract** One of the possible risks incurred while diving is inert gas narcosis (IGN), yet its mechanism of action remains a matter of controversy. Although providing insights in the basic mechanisms of IGN, research has been primarily limited to animal studies. A human study, in real diving conditions, was needed. Twenty volunteers within strict biometrical criteria (male, age 30–40 years, BMI 20–23, non smoker) were selected. They performed a no-decompression dive to a depth of 33 mfw for 20 min and were assessed by the means of critical flicker fusion frequency (CFFF) measurement before the dive, during the dive upon arriving at the bottom, 5 min before the ascent, and 30 min after surfacing. After this late measurement, divers breathed oxygen for 15 min and were assessed a final time. Compared to the pre-dive value the mean value of each measurement was significantly different ( $p < 0.001$ ). An increase of CFFF to  $104 \pm 5.1$  % upon arriving to the bottom is followed by a decrease to  $93.5 \pm 4.3$  %. This impairment of CFFF persisted 30 min after surfacing, still decreased to  $96.3 \pm 8.2$  % compared to pre-dive CFFF. Post-dive measures made after 15 min of oxygen were not different from control (without nitrogen supersaturation),  $124.4 \pm 10.8$  versus  $124.2 \pm 3.9$  %. This simple study suggests that IGN (at least partially) depends on gas-protein interactions and that the cerebral impairment persists for at least 30 min after surfacing. This could be an important consideration in situations where precise and accurate judgment or actions are essential.

---

**Keywords (separated by '-')** Diving - Inert gas narcosis - Critical flicker fusion frequency

---

**Footnote Information** Communicated by Dag Linnarsson.

---

Journal: 421  
Article: 2391

Author Query Form  Springer  
the language of science

**Please ensure you fill out your response to  
the queries raised below  
and return this form along with your  
corrections**

Dear Author

During the process of typesetting your article, the following queries have arisen. Please check your typeset proof carefully against the queries listed below and mark the necessary changes either directly on the proof/online grid or in the ‘Author’s response’ area provided below

<b>Query</b>	<b>Details required</b>	<b>Author's response</b>
1.	Please check and confirm the author names and initials are correct. Also, kindly confirm the details in the metadata are correct.	
2.	Please check and confirm that the authors and their respective affiliations have been correctly identified and amend if necessary.	
3.	Please confirm the inserted city name is correct and amend if	

	necessary.	
4.	Please confirm the corresponding author is correctly identified and amend if necessary.	
5.	Please confirm the section headings are correctly identified.	
6.	Kindly update the reference Lauridsen MM (2011) with volume id and page range if possible	

2 **Persistence of critical flicker fusion frequency impairment**  
3 **after a 33 mfw SCUBA dive: evidence of prolonged nitrogen**  
4 **narcosis?**

5 C. Balestra · P. Lafère · P. Germonpré

6 Received: 29 June 2011 / Accepted: 19 March 2012  
7 © Springer-Verlag 2012

8 **Abstract** One of the possible risks incurred while diving  
9 is inert gas narcosis (IGN), yet its mechanism of action  
10 remains a matter of controversy. Although providing  
11 insights in the basic mechanisms of IGN, research has been  
12 primarily limited to animal studies. A human study, in real  
13 diving conditions, was needed. Twenty volunteers within  
14 strict biometrical criteria (male, age 30–40 years, BMI  
15 20–23, non smoker) were selected. They performed a no-  
16 decompression dive to a depth of 33 mfw for 20 min and  
17 were assessed by the means of critical flicker fusion fre-  
18 quency (CFFF) measurement before the dive, during the  
19 dive upon arriving at the bottom, 5 min before the ascent,  
20 and 30 min after surfacing. After this late measurement,  
21 divers breathed oxygen for 15 min and were assessed a  
22 final time. Compared to the pre-dive value the mean value  
23 of each measurement was significantly different  
24 ( $p < 0.001$ ). An increase of CFFF to  $104 \pm 5.1 \%$  upon  
25 arriving to the bottom is followed by a decrease to  
26  $93.5 \pm 4.3 \%$ . This impairment of CFFF persisted 30 min  
27 after surfacing, still decreased to  $96.3 \pm 8.2 \%$  compared

to pre-dive CFFF. Post-dive measures made after 15 min of  
oxygen were not different from control (without nitrogen  
supersaturation),  $124.4 \pm 10.8$  versus  $124.2 \pm 3.9 \%$ . This  
simple study suggests that IGN (at least partially) depends  
on gas-protein interactions and that the cerebral impair-  
ment persists for at least 30 min after surfacing. This could  
be an important consideration in situations where precise  
and accurate judgment or actions are essential.

**Keywords** Diving · Inert gas narcosis · Critical flicker  
fusion frequency

**Introduction**

Although SCUBA (self-contained underwater breathing  
apparatus) diving is relatively safe, one of the possible risks  
incurred is inert gas narcosis (IGN), also called “nitrogen  
narcosis” or rapture of the depths.

IGN can provoke several troubles (Lowry 2005; Richardson et al. 2005) such as temporal and spatial dis-orientation, physical coordination alteration, mood disorders, loss of long term memory. Symptoms of IGN resemble alcohol intoxication or the early stage of anesthesia or hypoxia (Dean et al. 2003). As depth and pressure increase, the symptoms worsen and eventually lead to unconsciousness (Bennett 2004; Pastena et al. 2005).

Although in 1935 Behnke et al. (1935) correctly associated these phenomena to a raised partial pressure of nitrogen, its precise mechanism of action remains a matter of controversy. For long, inert gas narcosis was regarded as a pure biophysical phenomenon and it was assumed that breathed nitrogen did not interact biochemically with the cellular metabolism (Bennett 2004; Lowry 2005). The traditional view was that narcosis or anesthesia occurred

A1 Communicated by Dag Linnarsson.

A2 C. Balestra · P. Lafère · P. Germonpré  
A3 DAN Europe Research Division, Brussels, Belgium  
A4 e-mail: daneuben@skynet.be

A5 P. Germonpré  
A6 e-mail: peter.germonpre@mil.be

A7 C. Balestra · P. Lafère · P. Germonpré  
A8 Environmental and Occupational Physiology Laboratory, Haute  
A9 Ecole “Paul Henri Spaak”, Brussels, Belgium

A10 P. Lafère (✉) · P. Germonpré  
A11 Centre For Hyperbaric Oxygen Therapy, Military Hospital  
A12 “Queen Astrid”, Bruynstreet 1, 1120 Brussels, Belgium  
A13 e-mail: doc.lafere@skynet.be

60 when the volume of a hydrophobic membrane site was  
 61 caused to expand beyond a critical level by the absorption  
 62 of molecules of a narcotic gas. The observation of the pres-  
 63 sure reversal effect during general anesthesia has long sup-  
 64 ported this lipid theory (Jibu 2001; Włodarczyk et al. 2006).  
 65 However, results of the most recent animal studies have  
 66 revealed that nitrogen narcosis could interact with the pro-  
 67 duction, release and uptake of several brain neurotransmitters  
 68 supporting a protein binding theory (Rostain et al. 2011).

69 In rats, neurochemical studies in the striatum have  
 70 demonstrated that a rise in nitrogen partial pressure induced  
 71 a decrease in dopamine release (Dedieu et al. 2004), a  
 72 decrease of glutamate concentration (Vallee et al. 2009,  
 73 2010), and also enhanced gamma-aminobutyric acid  
 74 (GABA-A) receptors activity (Balon et al. 2002; David  
 75 et al. 2001; Lavoute et al. 2008).

76 Because of the paucity of the literature a human study in  
 77 real diving conditions was needed to confirm that changes  
 78 in human brains parallel the observations made in vivo in  
 79 the rodent brain. However reliable indices to quantify the  
 80 effects of inert gas narcosis are not yet available. Ideally,  
 81 these indices should be reproducible, less subject- or  
 82 investigator-dependent than a psychometric behavioral  
 83 approach, based on observing a change in neurological  
 84 parameters like electroencephalographic recordings (Pas-  
 85 tena et al. 2005) but easy to implement underwater. The  
 86 critical flicker fusion frequency (CFFF) seems to answer  
 87 these needs. It is a tool that has already been used in the  
 88 field of diving medicine research (Seki and Hugon 1976).  
 89 The CFFF variations occur parallel to EEG modifications  
 90 and may reveal neuropsychological troubles that are not  
 91 apparent from subjective reports (Seki and Hugon 1976).  
 92 The use of such measure is advocated by the particular  
 93 characteristics of the CFFF: non invasive and of good  
 94 reliability in cortical arousal (Hou et al. 2007; Rota-Baterlink  
 95 1999) as well as a good marker of cortical alteration to  
 96 physical workload (Davranche and Pichon 2005; Luczak et al.  
 97 1995; Luczak and Sobolewski 2005), drug administration  
 98 (Hindmarch 1982; Hunter et al. 1994), alcohol intoxication  
 99 (Leigh 1982; Liu and Ho 2010; Schillaci and Fazio 1967),  
 100 anesthesia (Salib et al. 1992; Sharma et al. 2011; Wernberg  
 101 et al. 1980), hypoxia (Truszczynski et al. 2009) or in case of  
 102 encephalopathy (Ali et al. 1994; Chang et al. 2007; Kircheis  
 103 et al. 2002; Lauridsen et al. 2011). Using the CFFF, we  
 104 performed an objective measurement of the effects of IGN in  
 105 divers.

## 106 Materials and methods

107 After written informed consent and Ethics Committee  
 108 approval (CE2008/66), 20 male experienced divers (Min-  
 109 imum certification “Autonomous Divers” according to

European norm EN 14153-2 or ISO 24801-2 with at least 110  
 50 logged dives) volunteered for this study. They were 111  
 selected from a large sports diver population in order to obtain 112  
 a group of comparable age [30–40 years,  $35.38 \pm 3.59$  113  
 (mean  $\pm$  SD)], body composition (BMI between 20 and 114  
 25,  $23.6 \pm 1.15$ ) and comparable health status: non 115  
 smokers with regular but not excessive physical activity 116  
 (aerobic exercise one to three times a week). Prior to entry 117  
 into the study, they were assessed fit to dive. Divers 118  
 needing visual correction underwater and divers taking any 119  
 medications such as steroids, benzodiazepine, barbiturates, 120  
 or psychoactive drugs were excluded. Participants were 121  
 instructed not to dive 72 h prior to the experimental dive 122  
 and not to drink any alcoholic or caffeine-containing bev- 123  
 erages 4 h before the dive. 124

Each diver performed a dive to a depth of 33 mfw for 125  
 20 min in a pool environment (Nemo33, Brussels, Belgium) 126  
 with a water temperature of 33 °C, thus needing no thermal 127  
 protection suit. This depth-time profile falls within accepted 128  
 “no-decompression limits” (NAVSEA 2008). Descent 129  
 speed was at 15 m per minute and ascent speed was at 10 130  
 meters per minute to the surface, with no safety stop (none 131  
 required according to the dive table used). 132

Divers were assessed with the CFFF using a specific 133  
 watertight device built for the occasion by Human 134  
 Breathing Technology (HBT, Trieste, Italy). The device 135  
 consists of a rotating ring, surrounding a short cylindrical 136  
 waterproof housing of 8 cm diameter containing the 137  
 numeric (digital) frequency indicator. Attached to this 138  
 housing is a flexible cable, on the end of which a single 139  
 blue LED (Light Emitting Diode) (color temperature 140  
 8,000 K) is enclosed in a smaller cylindrical container (to 141  
 shield it from stray light and reflections). While the subject 142  
 to be tested is looking straight at the LED light at a distance 143  
 individually adapted to his personal vision (generally 144  
 around 50 cm), the investigator turns the dial slowly 145  
 clockwise or anticlockwise in order to increase or decrease 146  
 the flickering frequency of the LED. As there are no 147  
 markings on the dial, nor a visible “starting position”, the 148  
 test subject has no indication whatsoever of the actual 149  
 flicker frequency. When the subject sees a change from 150  
 fusion to flicker (or flicker to fusion), he signals this to the 151  
 investigator, who notes the actual frequency—which is the 152  
 definition of CFFF (Rota-Baterlink 1999; Tytla et al. 153  
 1990). This test is carried out systematically three times in 154  
 order to check its reproducibility. The average of the three 155  
 measurements was noted as the actual individual CFFF. 156  
 Divers were assessed immediately before the dive (base- 157  
 line), upon arriving at the bottom, 5 min before the ascent 158  
 (after 15 min at 33 mfw), and 30 min after surfacing. Once 159  
 the late measurement was made, the diver breathed oxygen 160  
 for 15 min (using a non-rebreather mask at a flow of 15 L 161  
 per minute) and then CFFF was assessed a final time. 162



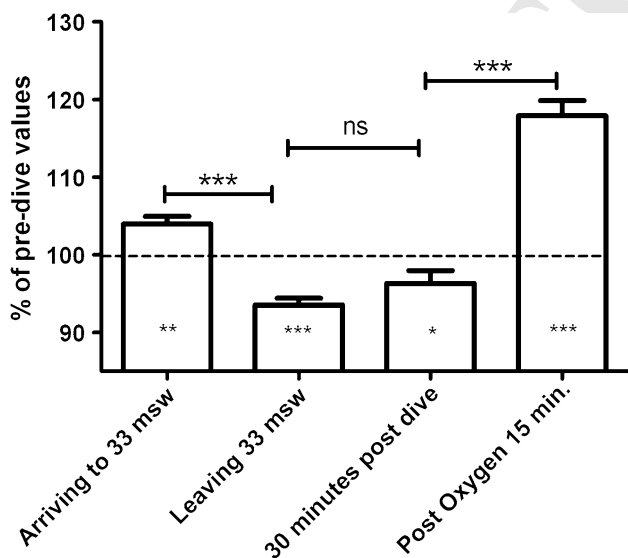
163 We furthermore performed a control experiment, where  
 164 the same individuals were assessed with CFFF before and  
 165 after 15 min of oxygen breathing without any dive sched-  
 166 uled or performed within a 3 days period in order to assess  
 167 any oxygen effect in absence of nitrogen supersaturation.

168 Taking the initial value as 100 %, percentage variations  
 169 were calculated allowing an appreciation of the magnitude  
 170 of the change rather than the absolute values. Standard  
 171 statistical analysis was performed after testing for normal-  
 172 ity, using GraphPad Prism version 5.00 for Windows  
 173 (GraphPad Software, San Diego, CA, USA) on a personal  
 174 computer.

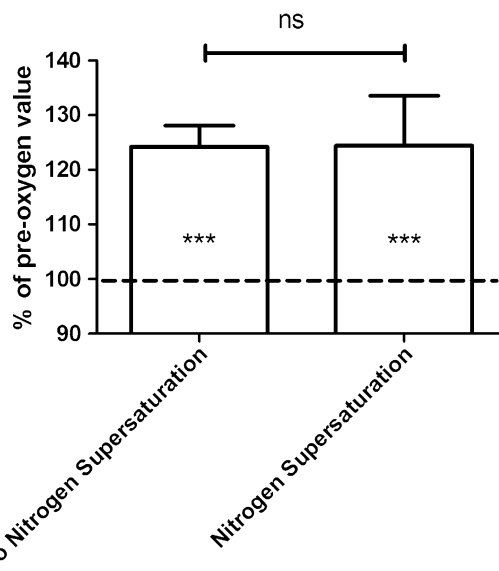
175 **Results**

176 All sets of data passed both Kolmogorov–Smirnov and  
 177 Shapiro–Wilk normality tests, allowing us to assume a  
 178 Gaussian distribution.

179 The evolution of CFFF during and after the dive is  
 180 illustrated in Fig. 1. Compared to the pre-dive value  
 181 (100 %) the mean value of each measurement is signifi-  
 182 cantly different. An increase of CFFF to  $104.0 \pm 5.1$  %  
 183 when arriving to the bottom is followed 15 min later by a  
 184 decrease to  $93.5 \pm 4.3$  %. This impairment of CFFF  
 185 persists 30 min after surfacing, being still decreased to  
 186  $96.3 \pm 8.2$  % compared to the pre-dive CFFF (100 %).  
 187 Each single measurement is statistically different from the  
 188 baseline (one sample *t* test  $p < 0.05$  or lower). Paired *t* test  
 189 demonstrated a statistical difference between the first and  
 190 second underwater measurement ( $p < 0.001$ ), but no



**Fig. 1** Percentage variation of CFFF during and after a 20 min dive to 33 mfw/110 ffw. Pre-dive CFFF value is taken as 100 %. Each subject is compared to his own pre-dive value. (\*\* $p < 0.001$ ; \*\* $p < 0.01$ ; \* $p < 0.05$ ; ns not significant) ( $n = 20$ )



**Fig. 2** Variation of CFFF after 15 min of oxygen breathing with and without diving. Pre-oxygen breathing CFFF value is taken as 100 % (when diving, pre-oxygen value is the post-dive value). Each subject is compared to his own pre-oxygen value. (\*\* $p < 0.0001$ ; ns not significant) ( $n = 20$ )

191 statistical difference between the second underwater mea-  
 192 surement and the post-dive measurement ( $p = 0.099$ ).  
 193 After 15 min of oxygen breathing, CFFF increases signifi-  
 194 cantly ( $p < 0.0001$ ) and is  $117.9 \pm 9.8$  % higher than the  
 195 pre-dive CFFF (Paired *t* test,  $p < 0.001$ ).

196 When non nitrogen supersaturated (Fig. 2), compared to  
 197 the pre-oxygen value (100 %), an increase of CFFF up to  
 198  $124.2 \pm 3.9$  % was noted which was statistically signifi-  
 199 cant (one sample *t* test,  $p < 0.001$ ). When diving (with  
 200 nitrogen supersaturation) we took the post-dive pre-oxygen  
 201 value as a new baseline to compare the oxygen effect with  
 202 the control experiment (without nitrogen supersaturation).  
 203 With this new baseline, the increase ( $124.4 \pm 10.8$  %  
 204 observed after oxygen breathing in the post-dive period) is  
 205 statistically not different from the non nitrogen saturated  
 206 increase (paired *t* test,  $p = 0.72$ ). This suggests that the  
 207 post-dive, post-oxygen increase of CFFF is due to a direct  
 208 effect of oxygen rather than to a supplemental nitrogen  
 209 washout effect by oxygen.

210 **Discussion**

211 Indices to quantify the effects of IGN can be roughly  
 212 divided into two approaches.

213 The first is a behavioral approach, measuring task per-  
 214 formance such as mental arithmetic, memory, reaction time  
 215 and manual dexterity. Although these behavioral studies  
 216 have confirmed a progressive deterioration with increasing  
 217 pressure, many of these tests have been criticized because

Author Proof

218 of the influences of motivation, experience and learning on  
219 the test results (Lowry 2005).

220 The second approach relies on observing a change in  
221 objective, measurable neurological parameters. In this  
222 matter, even if there are some limitations (Rota-Baterlink  
223 1999; Tytla et al. 1990), some authors have emphasized the  
224 advantages of CFFF assessment (Davranche and Pichon  
225 2005; Luczak and Sobolewski 2005; Truszczynski et al.  
226 2009) as an objective, quantitative, and important method  
227 for measuring alertness and arousal (Feshchenko et al.  
228 1994; Ginsburg et al. 1982; Luczak and Sobolewski 2000;  
229 Railton et al. 2009). Moreover, CFFF seems to be a better  
230 way of testing cerebral arousal than the classical behavioral  
231 approach as in anesthesia, CFFF has been demonstrated to  
232 parallel brain impairment earlier than subjective symptoms  
233 (Salib et al. 1992; Wernberg et al. 1980) or behavioral tests  
234 (number connection test A and B, digit symbol test, serial  
235 dotting test, and line tracing test) (Sharma et al. 2011).  
236 When executed in standard conditions, the CFFF test  
237 makes it possible to measure in a longitudinal way the  
238 evolution of the state of cortical arousal in test subjects  
239 (Luczak and Sobolewski 2005). The construction of a  
240 waterproof housing for the CFFF test device, designed to  
241 keep the test subject fully blinded to the frequency read-out  
242 of the flickering LED, has allowed for the first time “real-  
243 life” measurements of CFFF while under water.

244 The results of this study are also unique because to our  
245 knowledge, it is the first time that effect of inert gas nar-  
246 cosis is measured for a period of time after surfacing. One  
247 of the most remarkable observations was undoubtedly that  
248 the CFFF results at the 30 min post-dive time point dem-  
249 onstrated impairment of cerebral arousal persisting long  
250 after surfacing. Indeed, based on the lipid theory (Jibu  
251 2001; Włodarczyk et al. 2006), diver’s training programs  
252 advise that in the event of nitrogen narcosis, divers should  
253 ascend a few meters in order for the narcotic effects to  
254 dissipate rapidly. However, it is shown here that, even if  
255 subjective feelings of narcosis may rapidly abate, the  
256 cerebral impairment persists for at least 30 min after sur-  
257 facing. This may be an important consideration in situa-  
258 tions where precise and accurate judgment or actions are  
259 essential, such as in the hazardous situations in recreational  
260 diving or in professional (industrial, military) diving.

261 Recent observations suggest that there is a correlation  
262 between CFFF and post-dive perceived fatigue. In a previous  
263 study (Lafere et al. 2010) we have shown that in a large group  
264 of divers ( $n = 219$ ), the change in perceived fatigue level  
265 after a single dive is significantly lower when enriched air  
266 Nitrox (EAN<sub>x</sub>) was breathed rather than air which was  
267 demonstrated with a post-dive decrease of CFFF while  
268 breathing air and a slight post-dive increase while breathing  
269 EAN<sub>x</sub>. The only difference between these two groups resi-  
270 ded in the different proportion of oxygen/nitrogen in the

271 breathing mixture, emphasizing the importance of the effect  
272 of these two gases on brain function. Indeed, electroen-  
273 cephalographic recordings of subjects exposed to com-  
274 pressed atmosphere in a pressure chamber in which the  
275 partial pressure of both oxygen and nitrogen were controlled,  
276 showed that any changes observed were related to the oxygen  
277 partial pressure and that the depressant effect of nitrogen is  
278 only revealed when a mixture containing a partial pressure of  
279 0.2 ATA of oxygen is breathed (Pastena et al. 2005).

280 As oxygen seems to be the most important gas, it has to  
281 be remembered that hyperoxia has been shown to facilitate  
282 nerve conduction, possibly as a consequence of oxidative  
283 stress (Brerrow-Saby et al. 2010). An enhanced production of  
284 reactive oxygen species (ROS) alters the conductance of  
285 potassium channels in excitable cells (Kovachich et al.  
286 1981; Matalon et al. 2003). Oxygen is also known to  
287 interact with GABA neurotransmission by influencing the  
288 synthesis, secretion, and recapture of this neurotransmitter.  
289 Indeed, when rat hippocampal slices are deprived of oxy-  
290 gen and glucose, GABA levels increase rapidly and then  
291 normalize within 15 min of reoxygenation (Radomski and  
292 Watson 1973; Schwartz-Bloom and Sah 2001). Finally,  
293 oxygen acts on the production of ammonia (NH<sub>3</sub>) by des-  
294 amination of catecholamines, tending to decrease the  
295 cerebral concentration of GABA (Banister and Singh  
296 1981). The consequence of all these mechanisms could be  
297 among others an increased inhibition of the inhibitory  
298 cerebral pathways.

299 These mechanisms have been studied in hyperbaric  
300 hyperoxia, and are able to provoke “hyperoxic” seizures  
301 as a result of imbalance between glutaminergic and  
302 GABAergic synaptic function (Demchenko and Piantad-  
303 osi 2006). However, even in “normobaric hyperoxia”  
304 (PpO<sub>2</sub> ≤ 1 ATA) this effect can be measured (Zhang  
305 et al. 1993). Abraini et al. (2003) have also emphasize  
306 the possible significant role of GABA (A) receptor as their  
307 results support a selective antagonism of the narcotic  
308 action of nitrogen.

309 The CFFF measurements, before and after oxygen  
310 breathing in non-divers, seem to confirm the effect of  
311 oxygen on cerebral arousal. CFFF increased by almost  
312 25 % compared to baseline measurements. This same  
313 effect could be responsible for the increased CFFF  
314 observed in the beginning of the dive. While at 33 mfw  
315 depth, divers breathing air actually breathe a gas with a  
316 PpO<sub>2</sub> of 0.9 ATA (Dalton’s Law: 21 % × 4.3 ATA),  
317 which is almost equivalent to breathing pure oxygen at  
318 surface. It could also be a good explanation for the effect of  
319 post-dive oxygen breathing, as the increase from the CFFF  
320 at 30 min post-dive is also 24.4 %. Although an acceler-  
321 ated nitrogen washout (denitrogenation) effect cannot for-  
322 mally be excluded, the similarity in CFFF increase is  
323 striking.

324 Moreover, the progressive reduction of the CFFF in the  
325 course of the dive seems to suggest a competition between  
326 the effect of oxygen and the effect of nitrogen. With time at  
327 depth, brain nitrogen concentrations increase up to a suf-  
328 ficient level within the effect-site and narcosis sets in, as  
329 measured by the reduction of CFFF after 15 min into the  
330 dive. Upon return to surface, blood nitrogen concentrations  
331 return to baseline, but the persistent reduction of CFFF  
332 shows that that the narcotic effects dissipate only slowly.  
333 Breathing oxygen after surfacing again decreases the  
334 inhibitory pathways, restoring CFFF to a supra-normal  
335 level.

336 Although these phenomena are quite complex, this  
337 study, carried out in real diving condition, provides an  
338 objective and reproducible measurement and makes it  
339 possible to suggest some conclusions, namely that nitrogen  
340 narcosis seems indeed to depend partly on a gas-protein  
341 interaction and that the system seems to be adaptive. Fur-  
342 ther studies may shed more light on the complex phe-  
343 nomena involved in the functional changes of the nervous  
344 system in the diving environment.  
345

## 346 References

347 Abraïni JH, Kriem B, Balon N, Rostain JC, Risso JJ (2003) Gamma-  
348 aminobutyric acid neuropharmacological investigations on nar-  
349 cosis produced by nitrogen, argon, or nitrous oxide. *Anesth*  
350 *Analg* 96:746–749 (Table of contents)  
351 Ali MR, Khaleque A, Khanam M, al-Shatti A, Ahmed RU et al (1994)  
352 Critical flicker frequency of mentally retarded and normal  
353 persons. *Percept Mot Skills* 79:1235–1238  
354 Balon N, Kriem B, Weiss M, Rostain JC (2002) GABA(A) receptors  
355 in the pars compacta and GABA(B) receptors in the pars  
356 reticulata of rat substantia nigra modulate the striatal dopamine  
357 release. *Neurochem Res* 27:373–379  
358 Banister EW, Singh AK (1981) The central role of Acmonia in OHD-  
359 induced Convulsions. In: Bachrach AJ, Matzen MM (eds)  
360 Underwater physiology VII. Undersea Med Soc Inc, Bethesda,  
361 pp 37–44  
362 Behnke AR, Thomson RM, Motley EP (1935) The physiologic effects  
363 from breathing air at 4 atmospheres pressure. *Am J Physiol*  
364 112:554–558  
365 Bennett PB (2004) Inert gas narcosis and high-pressure nervous  
366 syndrome In: Bove AA (ed) Bove and Davis' diving medicine  
367 (4th edn). Saunders, Philadelphia, pp 225–240  
368 Brerro-Saby C, Delliaux S, Steinberg JG, Jammes Y (2010) The  
369 changes in neuromuscular excitability with normobaric hyper-  
370 oxia in humans. *Exp Physiol* 95:153–159  
371 Chang TT, Ciuffreda KJ, Kapoor N (2007) Critical flicker frequency  
372 and related symptoms in mild traumatic brain injury. *Brain Inj*  
373 21:1055–1062  
374 David HN, Balon N, Rostain JC, Abraïni JH (2001) Nitrogen at raised  
375 pressure interacts with the GABA(A) receptor to produce its  
376 narcotic pharmacological effect in the rat. *Anesthesiology*  
377 95:921–927  
378 Davranche K, Pichon A (2005) Critical flicker frequency threshold  
379 increment after an exhausting exercise. *J Sport Exerc Psychol*  
380 27:515–520

Dean JB, Mulkey DK, Garcia AJ 3rd, Putnam RW, Henderson RA 381  
3rd (2003) Neuronal sensitivity to hyperoxia, hypercapnia, and 382  
inert gases at hyperbaric pressures. *J Appl Physiol* 95:883–909 383  
Dedieu D, Balon N, Weiss M, Risso JJ, Kinkead R, Rostain JC (2004) 384  
Microdialysis study of striatal dopaminergic dysfunctions 385  
induced by 3 MPa of nitrogen- and helium-oxygen breathing 386  
mixtures in freely moving rats. *Brain Res* 998:202–207 387  
Demchenko IT, Piantadosi CA (2006) Nitric oxide amplifies the 388  
excitatory to inhibitory neurotransmitter imbalance accelerating 389  
oxygen seizures. *Undersea Hyperb Med* 33:169–174 390  
Feshchenko VA, Reinsel RA, Veselis RA (1994) Optimized method 391  
of estimation of critical flicker frequency (CFF). *Proc Annu*  
392 *Symp Comput Appl Med Care* 1006 393  
Ginsburg N, Jurenovskis M, Jamieson J (1982) Sex differences in 394  
critical flicker frequency. *Percept Mot Skills* 54:1079–1082 395  
Hindmarch I (1982) Critical flicker fusion frequency (CFF): the 396  
effects of psychotropic compounds. *Pharmacopsychiatry* 15 397  
(44):48 398  
Hou RH, Langley RW, Szabadi E, Bradshaw CM (2007) Comparison 399  
of diphenhydramine and modafinil on arousal and autonomic 400  
functions in healthy volunteers. *J Psychopharmacol* 21:567–578 401  
Hunter KM, Zacharias M, Parkinson R, Luyk NH (1994) Effect of 402  
flumazenil on the recovery from intravenous midazolam. *N Z*  
403 *Dent J* 90:9–12 404  
Jibu M (2001) Theory of cell membrane organizers and pressure 405  
reversal of anesthesia. *Med Hypotheses* 56:26–32 406  
Kircheis G, Wettstein M, Timmermann L, Schnitzler A, Haussinger D 407  
(2002) Critical flicker frequency for quantification of low-grade 408  
hepatic encephalopathy. *Hepatology* 35:357–366 409  
Kovachich GB, Mishra OP, Clark JM (1981) Depression of cortical 410  
Na<sup>+</sup>, K<sup>+</sup>-ATPase activity in rats exposed to hyperbaric oxygen. 411  
*Brain Res* 206:229–232 412  
Lafere P, Balestra C, Hemelryck W, Donda N, Sakr A, Taher A, 413  
Marroni S, Germonpre P (2010) Evaluation of critical flicker 414  
fusion frequency and perceived fatigue in divers after air and 415  
enriched air nitrox diving. *Diving Hyperbaric Med* 40:114–118 416  
Lauridsen MM, Jepsen P, Vilstrup H (2011) Critical flicker frequency 417  
and continuous reaction times for the diagnosis of minimal 418  
hepatic encephalopathy. A comparative study of 154 patients 419  
with liver disease. *Metab Brain Dis* 26:135–139 420  
Lavoute C, Weiss M, Rostain JC (2008) Alterations in nigral NMDA 421  
and GABAA receptor control of the striatal dopamine level after 422  
repetitive exposures to nitrogen narcosis. *Exp Neurol* 212:63–70 423  
Leigh G (1982) The combined effects of alcohol consumption and 424  
cigarette smoking on critical flicker frequency. *Addict Behav*  
425 7:251–259 426  
Liu YC, Ho CH (2010) Effects of different blood alcohol concen- 427  
trations and post-alcohol impairment on driving behavior and 428  
task performance. *Traffic Inj Prev* 11:334–341 429  
Lowry C (2005) Inert gas narcosis. In: Edmons C, Lowry C, 430  
Pennefather J, Walker R (eds) *Diving and Subaquatic Medicine*, 431  
4th edn. Hodder Arnold, London, pp 183–193 432  
Luczak A, Sobolewski A (2000) The relationship between critical 433  
flicker fusion frequency (CFFF) and temperamental character- 434  
istics. *Int J Occup Saf Ergon* 6:493–505 435  
Luczak A, Sobolewski A (2005) Longitudinal changes in critical 436  
flicker fusion frequency: an indicator of human workload. 437  
*Ergonomics* 48:1770–1792 438  
Luczak A, Kurkus-Rozowska B, Sobolewski A (1995) Flicker test as 439  
a load measurement during the combined effect of heat and 440  
noise. *Int J Occup Saf Ergon* 1:160–166 441  
Matalon S, Hardiman KM, Jain L, Eaton DC, Kotlikoff M, Eu JP, Sun 442  
J, Meissner G, Stampler JS (2003) Regulation of ion channel 443  
structure and function by reactive oxygen-nitrogen species. *Am J*  
444 *Physiol Lung Cell Mol Physiol* 285:L1184–L1189 445

- 446 NAVSEA (2008) The air decompression table. In: NAVSEA (ed) US  
447 Navy Diving Manual (Revision 6): SS521-AG-PRO-010/0910-  
448 LP-106-0957. US Navy
- 449 Pastena L, Faralli F, Mainardi G, Gagliardi R (2005) EEG patterns  
450 associated with nitrogen narcosis (breathing air at 9 ATA). *Aviat*  
451 *Space Environ Med* 76:1031–1036
- 452 Radomski MW, Watson WJ (1973) Effect of lithium on acute oxygen  
453 toxicity and associated changes in brain gamma-aminobutyric  
454 acid. *Aerosp Med* 44:387–392
- 455 Railton RC, Foster TM, Temple W (2009) A comparison of two  
456 methods for assessing critical flicker fusion frequency in hens.  
457 *Behav Processes* 80:196–200
- 458 Richardson D, Kinsella D, Schreeves K (2005) Gas Narcosis. In:  
459 Richardson D, Kinsella D, Schreeves K (eds) *The encyclopedia*  
460 *of recreational diving*. PADI, Rancho Santa Margarita, pp 20–23
- 461 Rostain JC, Lavoute C, Risso JJ, Vallee N, Weiss M (2011) A review  
462 of recent neurochemical data on inert gas narcosis. *Undersea*  
463 *Hyperb Med* 38:49–59
- 464 Rota-Baterlink RA (1999) The diagnostic value of automated  
465 threshold perimetry. *Cur Opin Ophtalmol* 10:135–139
- 466 Salib Y, Plourde G, Alloul K, Provost A, Moore A (1992) Measuring  
467 recovery from general anaesthesia using critical flicker fre-  
468 quency: a comparison of two methods. *Can J Anaesth* 39:  
469 1045–1050
- 470 Schillaci C, Fazio O (1967) Critical fusion frequency. (Its changes  
471 after ingestion of alcohol). *Boll Ocul* 46:772–782
- 472 Schwartz-Bloom RD, Sah R (2001) Gamma-aminobutyric acid(A) neu-  
473 rotransmission and cerebral ischemia. *J Neurochem* 77:353–371
- Seki K, Hugon M (1976) Critical flicker frequency (CFF) and  
subjective fatigue during an oxyhelium saturation dive at 62  
ATA. *Undersea Biomed Res* 3:235–247
- Sharma P, Singh S, Sharma BC, Kumar M, Garg H, Kumar A, Sarin  
SK (2011) Propofol sedation during endoscopy in patients with  
cirrhosis, and utility of psychometric tests and critical flicker  
frequency in assessment of recovery from sedation. *Endoscopy*  
43:400–405
- Truszczynski O, Wojtkowiak M, Biernacki M, Kowalczyk K (2009)  
The effect of hypoxia on the critical flicker fusion threshold in  
pilots. *Int J Occup Med Environ Health* 22:13–18
- Tytla ME, Trope GE, Buncic JR (1990) Flicker sensitivity in treated  
ocular hypertension. *Ophthalmology* 97:36–43
- Vallee N, Rostain JC, Risso JJ (2009) How can an inert gas  
counterbalance a NMDA-induced glutamate release? *J Appl*  
*Physiol* 107:1951–1958
- Vallee N, Rostain JC, Risso JJ (2010) A pressurized nitrogen  
counterbalance to cortical glutamatergic pathway stimulation.  
*Neurochem Res* 35:718–726
- Wernberg M, Nielsen SF, Hommelgaard P (1980) A comparison  
between reaction time measurement and critical flicker fusion  
frequency under rising nitrous oxide inhalation in healthy  
subjects. *Acta Anaesthesiol Scand* 24:86–89
- Wlodarczyk A, McMillan PF, Greenfield SA (2006) High pressure  
effects in anaesthesia and narcosis. *Chem Soc Rev* 35:890–898
- Zhang J, Su Y, Oury TD, Piantadosi CA (1993) Cerebral amino acid,  
norepinephrine and nitric oxide metabolism in CNS oxygen  
toxicity. *Brain Res* 606:56–62

UNCORRECTED